

# How to use LAS AF

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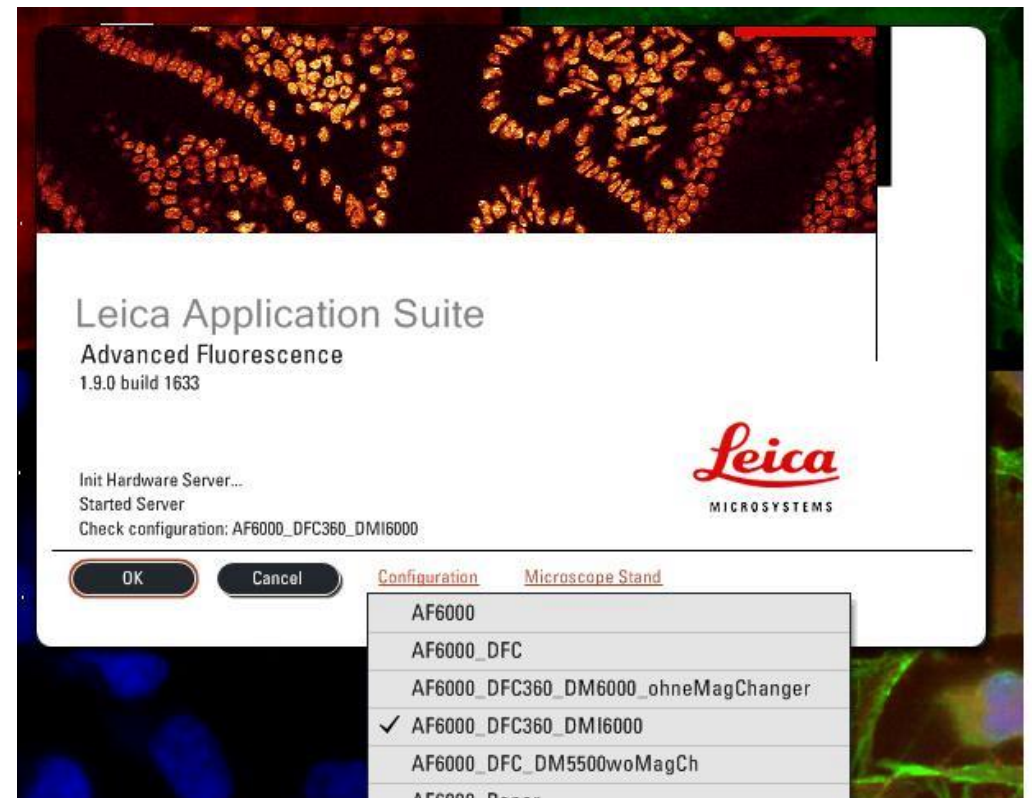
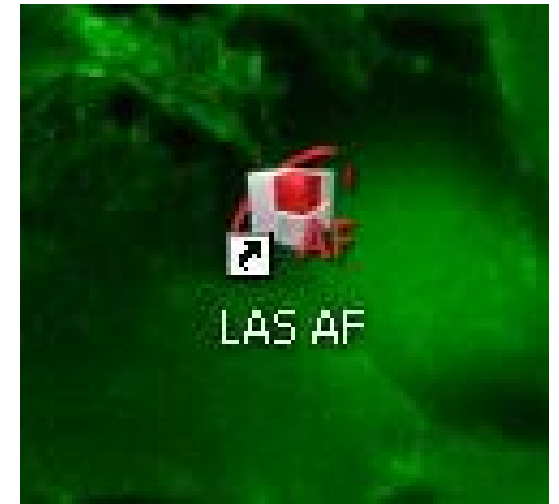
- § Installation
  - § Workflow
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  - § Acquire (Image Viewer, z-stack, time lapse, stage)
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# Installation:

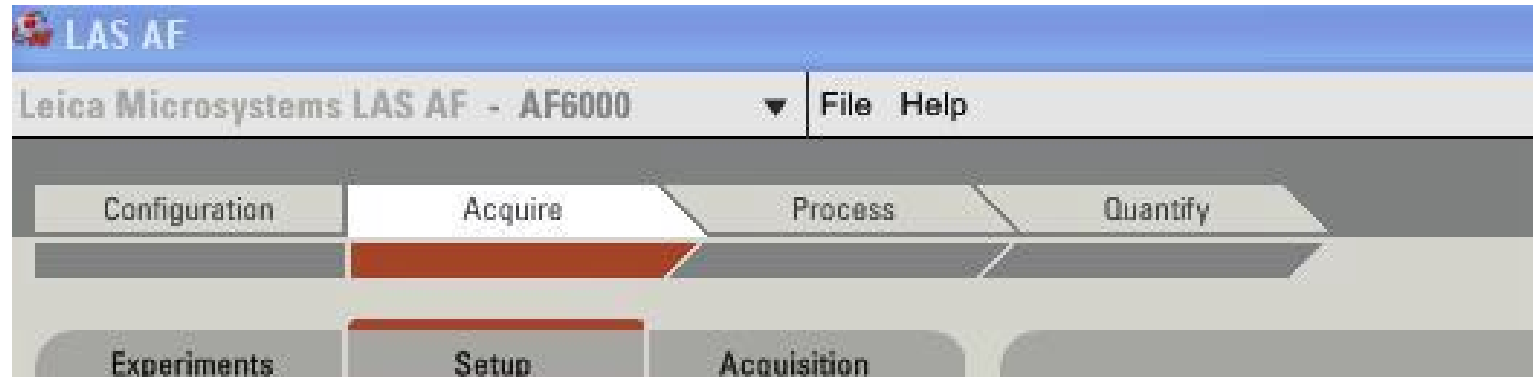
- § Installation CD
  - § 2 updates / year: FTP-server Leica Mannheim
  - § Installs / updates Leica LAS (Basis software)
  - § Firmware upgrade of microscope may be necessary
  - § Hardware configuration and data are safe
  - § Full software is installed: dongle protection
  - § LAS AF lite: freeware software (Viewer, Export)
  - § Review software with modules is available
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# Starting the software:

- § Select the correct configuration
- § Microscope, camera etc. must be switched on
- § System starts with the settings of the last session
- § Stage initialization not necessary, if no motor stage functions for this session

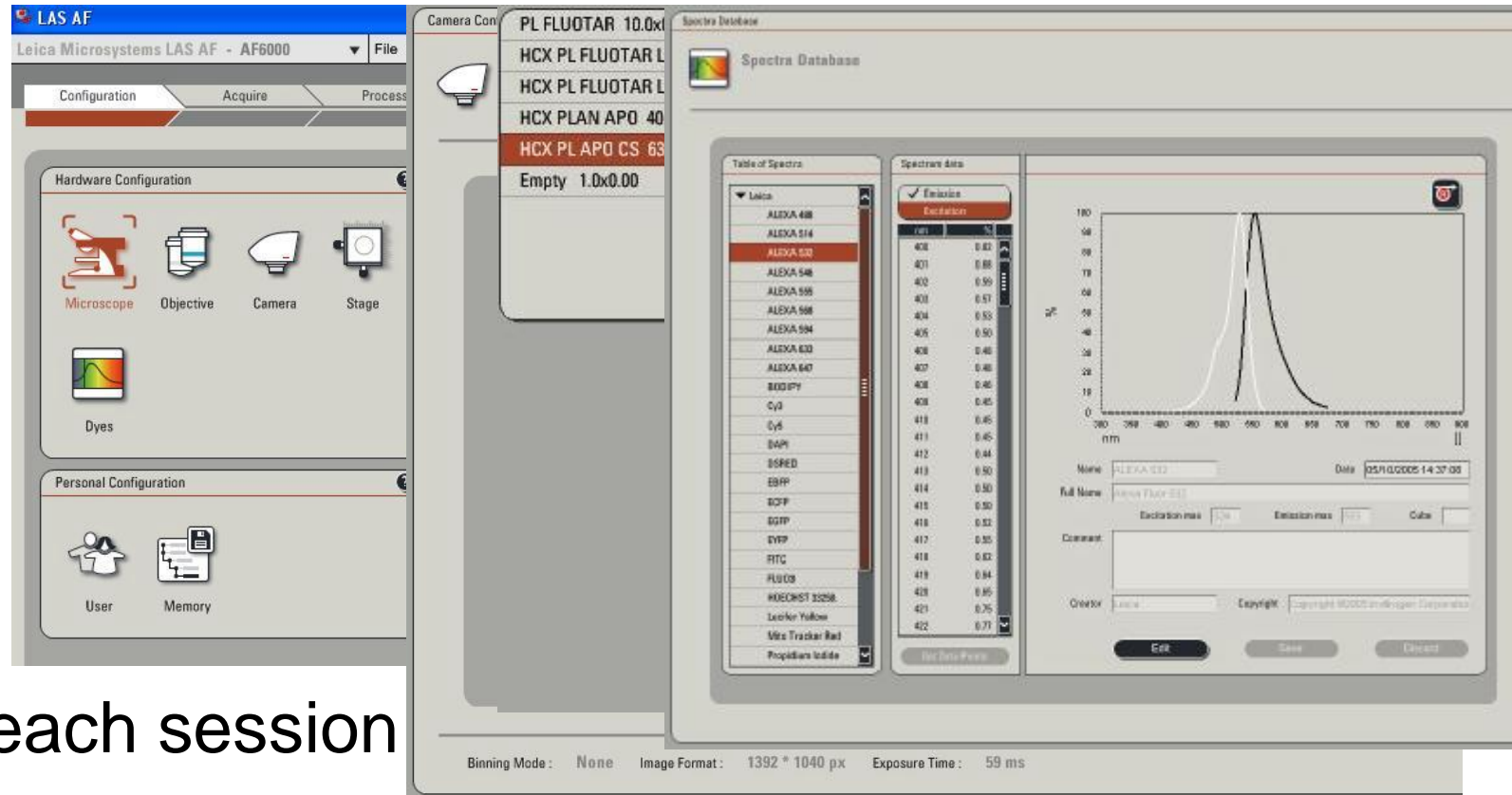


# Workflow:

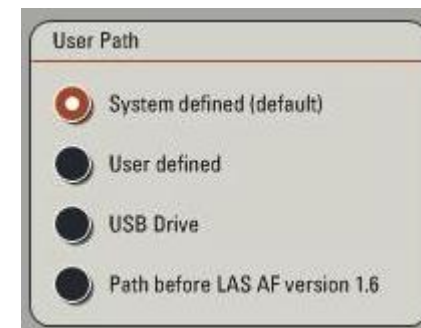


- § Directs the user
- § System starts in “Acquire”

# Configure:



- § No need for each session
- § Change camera settings if necessary (c-Mount, 8bit – 16 bit)
- § Display of objective parameters
- § List of fluorochromes
- § User settings



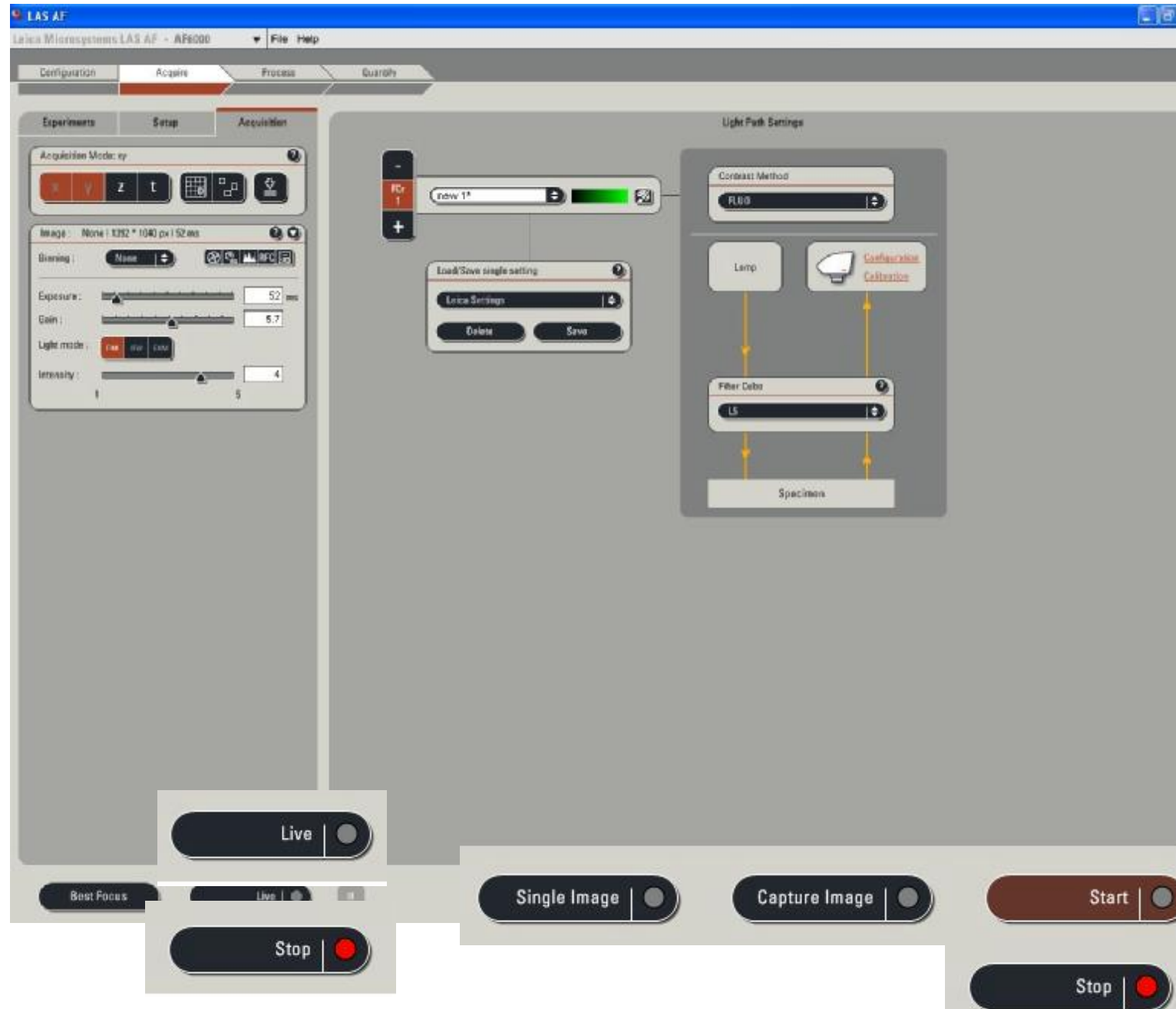
# Acquire:

- § Go to “Acquire” sub-menu
- § Define **Light Path**
- § Select contrast method
- § Select filter cube
- § Select look-up-table (LUT)
- § Go to “LIVE”
- § Adjust camera settings
- § Save channel (right mouse click)
- § Add more channels



# Acquire:

- § Camera Settings
- § LIVE on/off:  
Preview (frozen image)
- § SINGLE IMAGE:  
1 channel only
- § CAPTURE:  
1 set of channels
- § START:  
1 experiment  
(z-stack, time lapse,..)





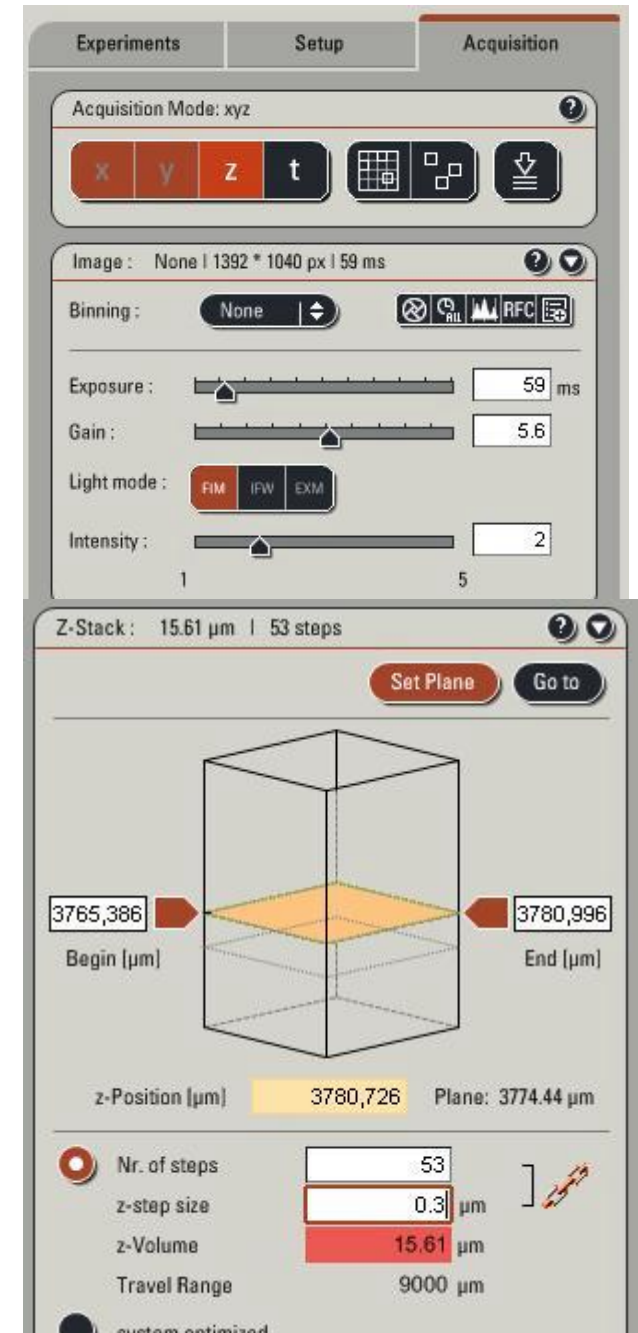
# Image Viewer:

The screenshot displays the Leica LAS AF software interface, which is divided into several sections:

- Top Bar:** Shows the application name "LAS AF" and the menu "Leica Microsystems LAS AF - AF6000" with "File" and "Help" options.
- Navigation Tabs:** Includes "Configuration", "Acquire" (the active tab), and "Process".
- Sub-Tabs:** Under "Acquire", there are "Experiments", "Setup", and "Acquisition" sub-tabs.
- Acquisition Mode:** Set to "xy".
- Image Information:** Shows "Image : None | 1392 \* 1040 px | 59 ms".
- Settings:**
  - Binning:** Set to "None".
  - Exposure:** Set to 59 ms.
  - Gain:** Set to 5.6.
  - Light mode:** Options for "FIM", "IFW", and "EXM".
  - Intensity:** Set to 2, with a scale from 1 to 5.
- Main View:** A large window displaying a fluorescence microscopy image of a biological specimen. The image shows a network of green and blue structures. A red arrow points to a specific feature. A scale bar at the bottom indicates a length of 40 μm.
- Toolbars:** A vertical toolbar on the left side of the main view contains various icons for image manipulation, such as zoom, pan, and measurement.

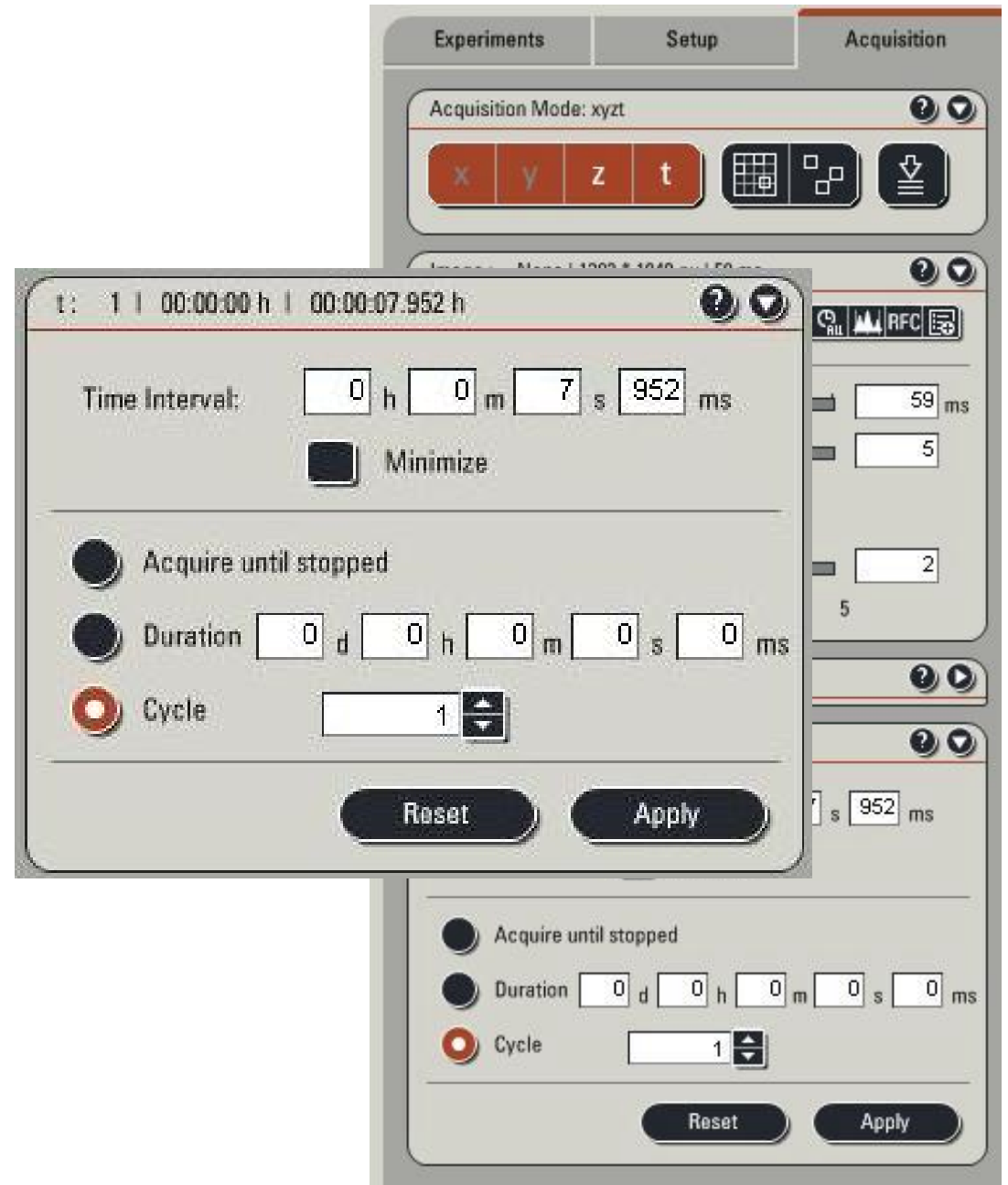
# Acquire z-stacks:

- § Select Z
- § Define “PLANE” (in focus)
- § Focus to upper and lower border and select
- § Select “System optimized” or define step size individually
- § START
- § Setup: first channel then z or vice versa



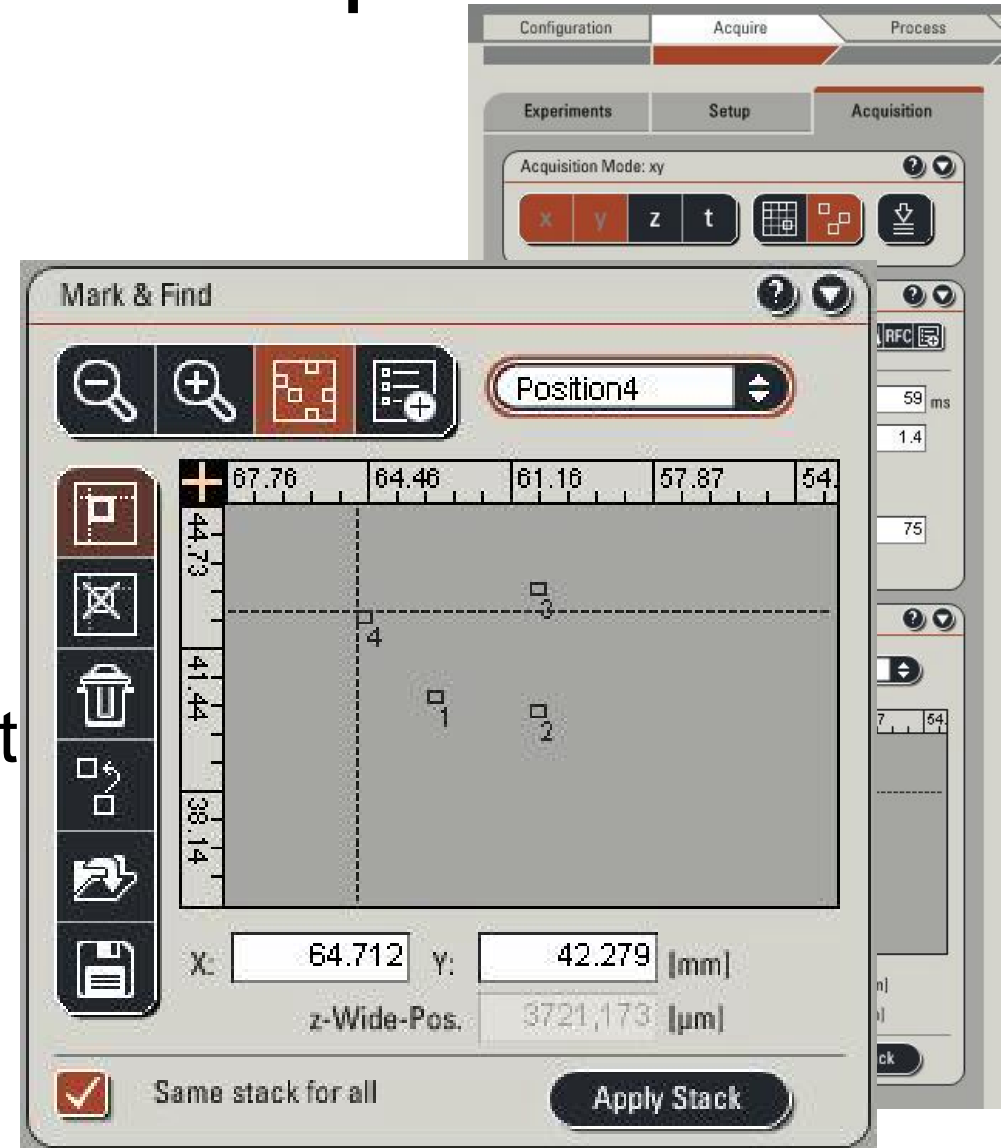
# Acquire time lapse:

- § Select T
- § Select DURATION or CYCLES or ACQUIRE UNTIL STOP
- § Select Interval
- § START
- § May be combined with Z



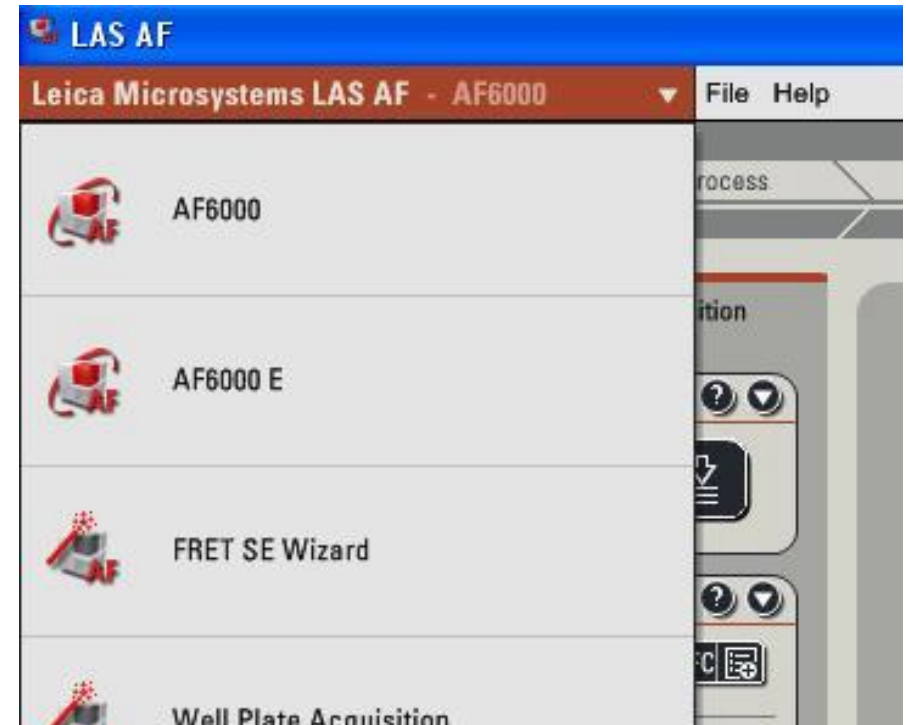
# Acquire multi position time lapse:

- § Select Stage settings
- § Select positions and check in stage control box
- § Positions are available in XYZ
- § Start Time Lapse Experiment



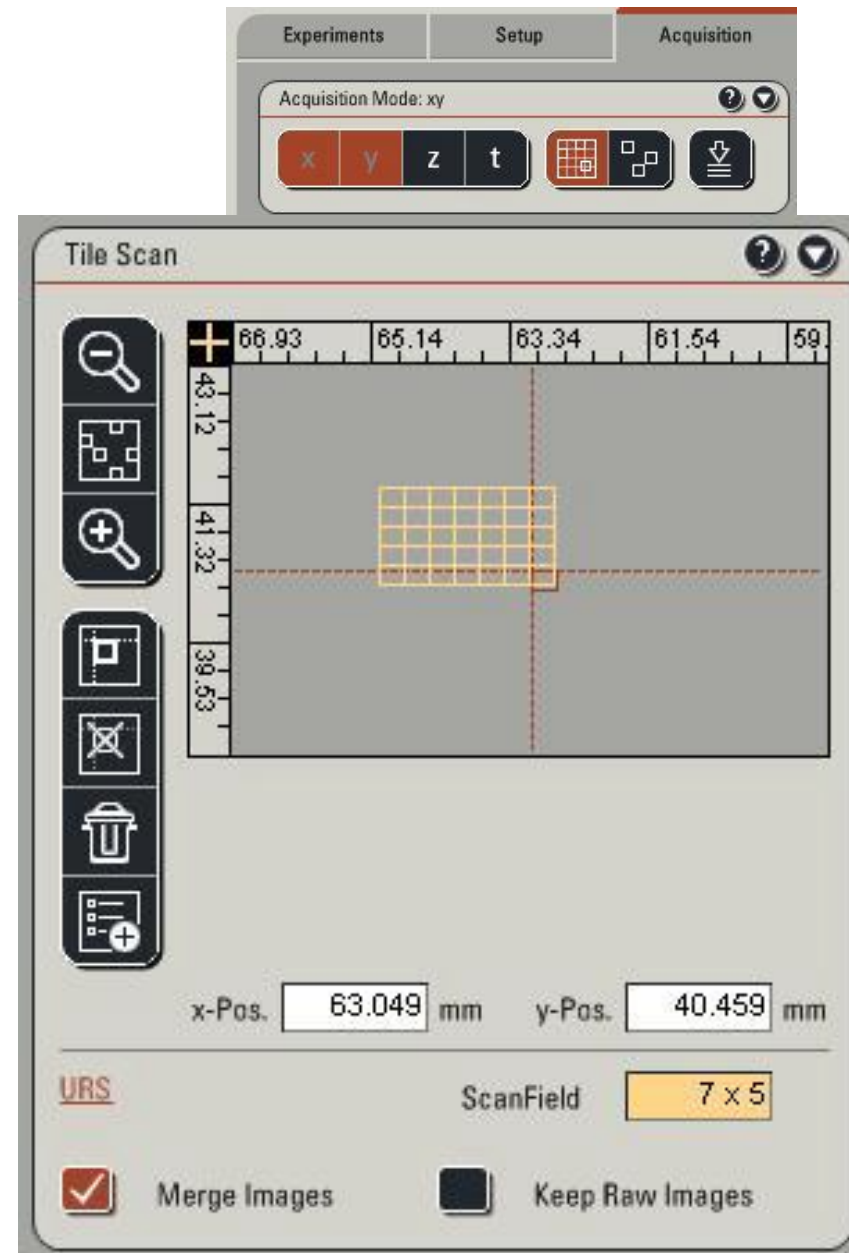
# Live Data Mode:

- § Optional Module
- § Definition of Jobs and Macros
- § Definition of Loops, Pause, Trigger,..
- § Tool for complex experiments



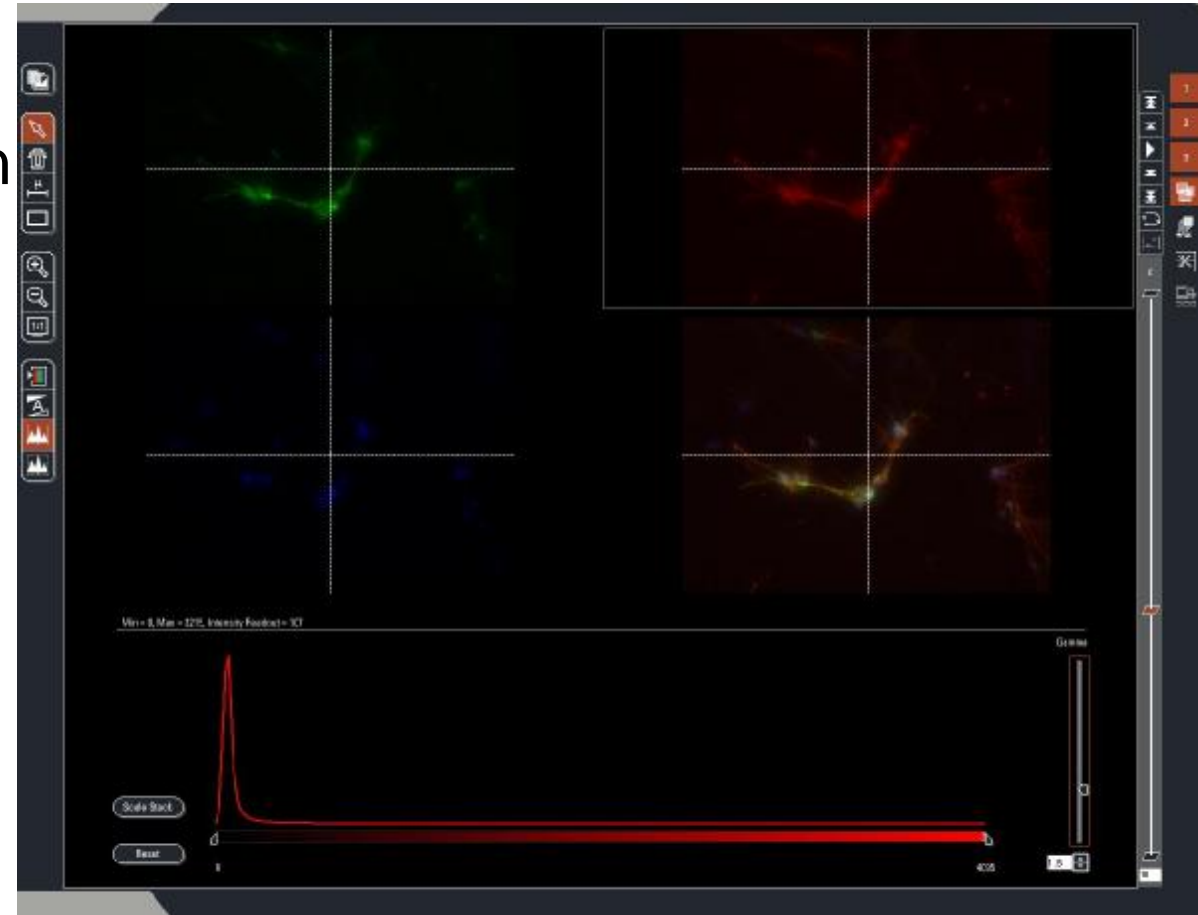
# Acquire Tile Scan:

- § Select Stage settings: Tile Scan
- § Select upper left and lower right position
- § Save individual images ?
- § Special stage settings ?
  
- § Start Acquisition
  
- § No Stitching: precision of the motor stage



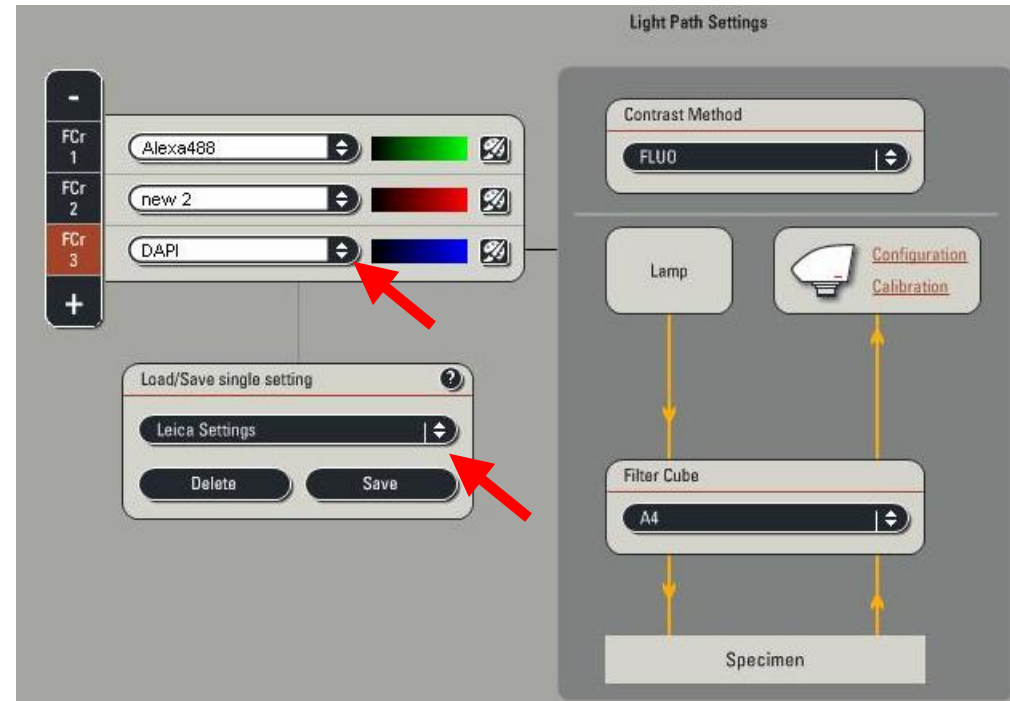
# Image Viewer: Gallery, Maximum Projection, sections

- § For complex experiments
- § Maximum Intensity Projection for z-Stacks and time lapse experiments
- § XZ- and YZ-Sections for visualization of z-stacks
- § Gallery
- § Rescaling of 16 bit
- § Autoscaling
- § Gamma value
- § Colour (LUT)

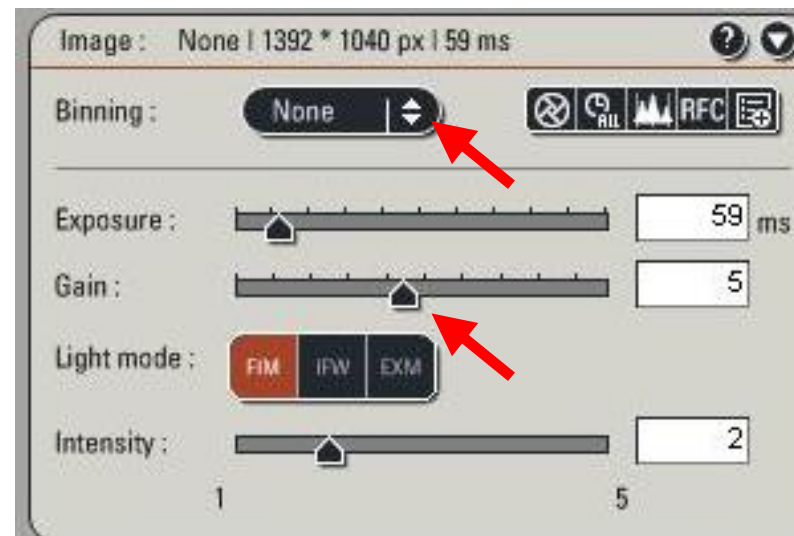


# Acquisition:

§ Save settings:  
each channel or  
each set of channels



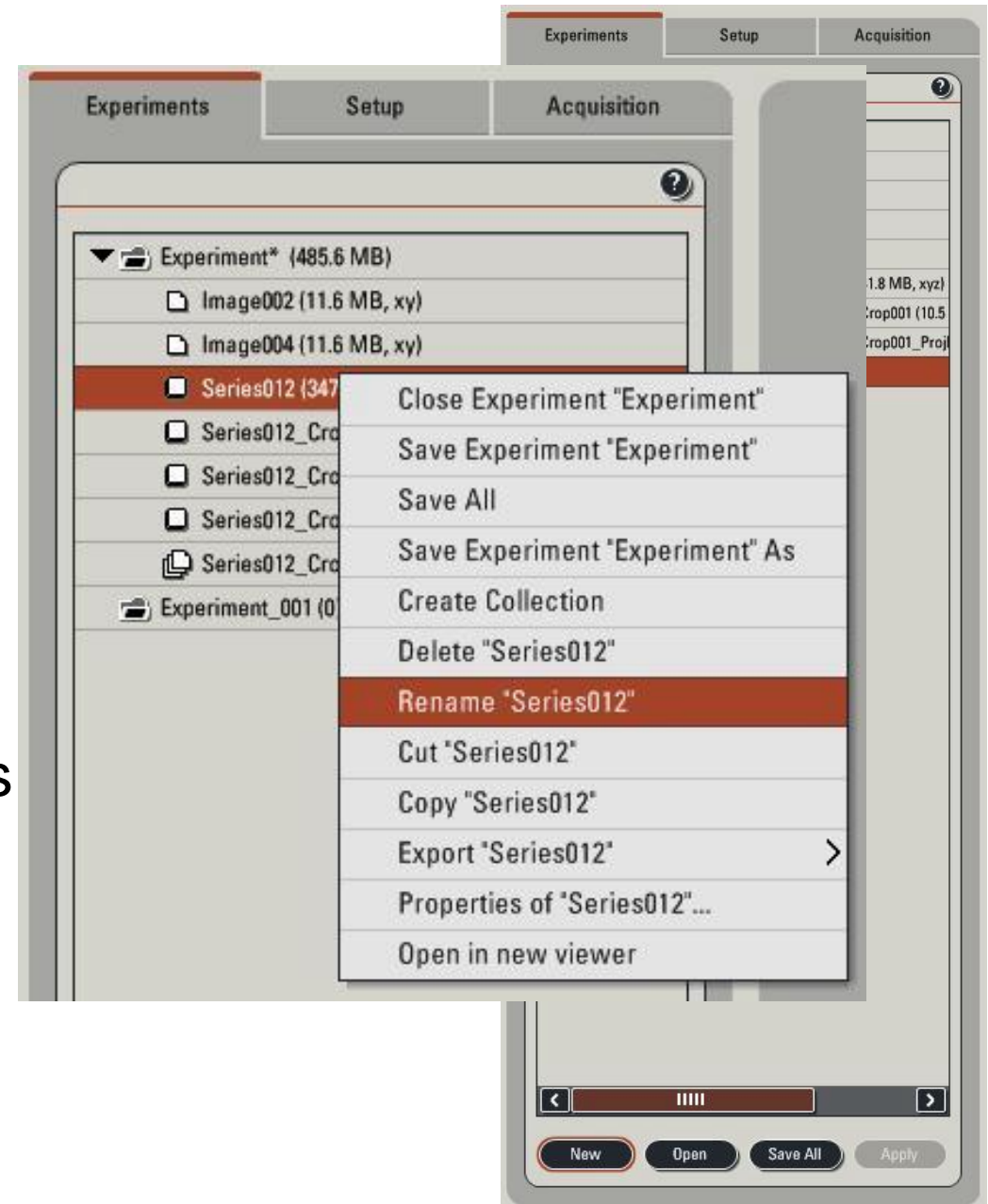
§ Camera settings:  
Binning and  
recommended gain  
(half maximum)





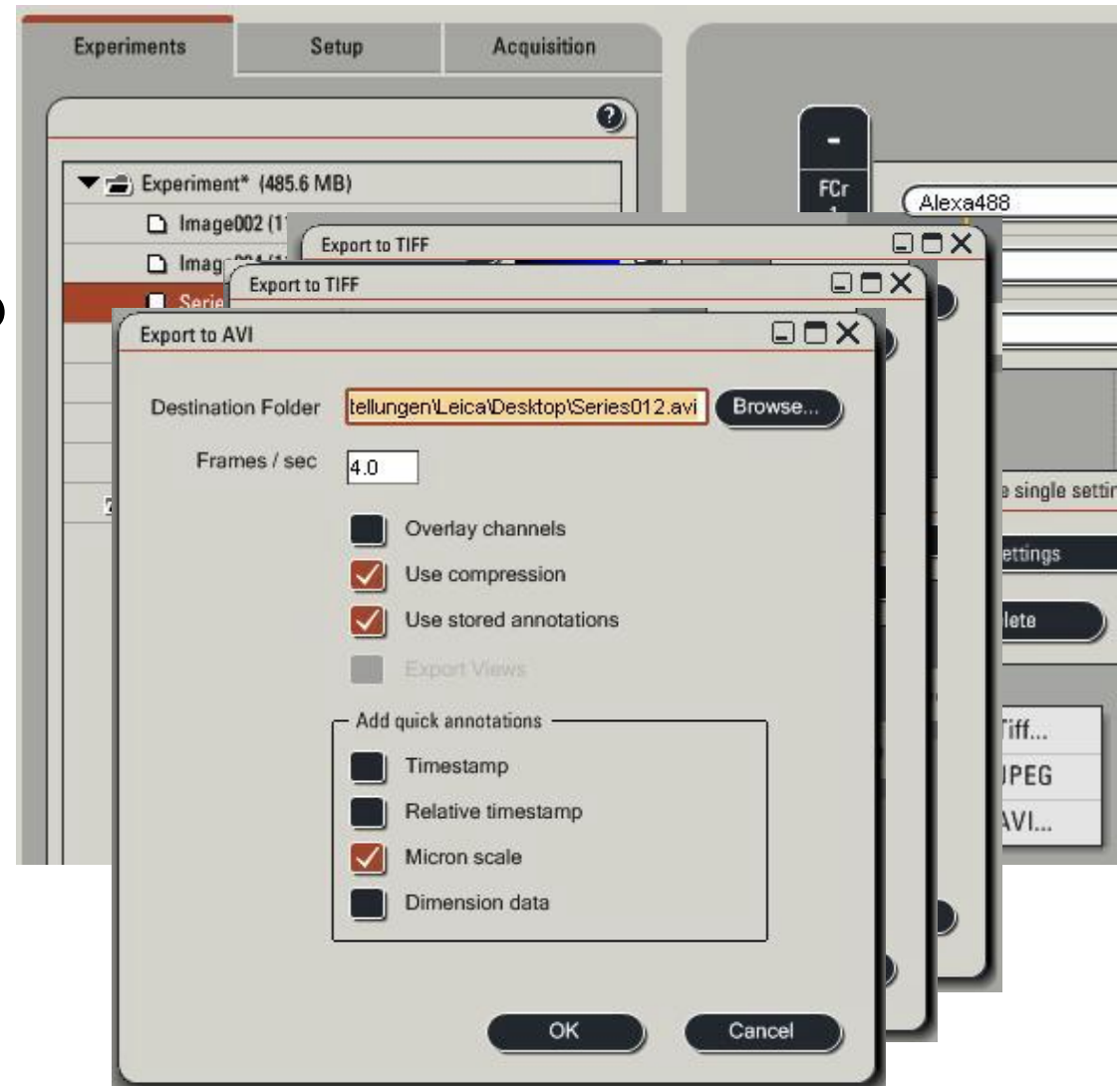
# Data Management:

- § All image sets automatically in Data Container
- § Save “Experiments”: LIF-format
- § LIF can be opened with LAS AF-lite (free of charge) on any PC-system
- § Moving data sets with Drag&Drop between experiments
- § Rename, Delete, Copy-Paste,... With right mouse click
- § Export TIF, JPEG, AVI



# Export:

- § Channel images or overlay
- § Export many data sets in 1 step
- § TIF: Raw images or as currently displayed in Image Viewer
- § Export annotations, Scale bar, Time Point,...
- § 1 file per image
- § AVI: select frame rate and compression



# Metadata:

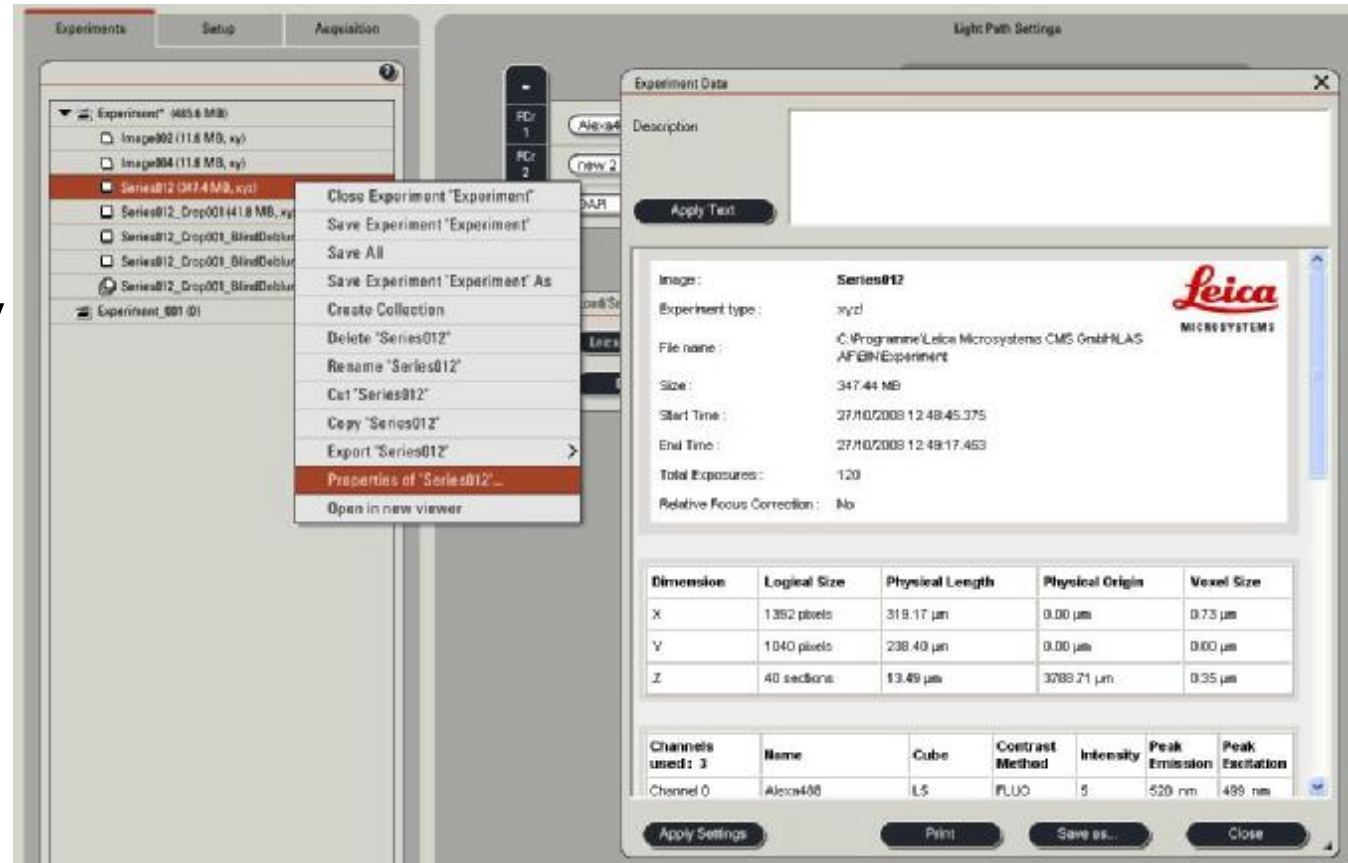
§ All relevant microscope and camera settings are saved automatically

§ Metadata visible as “Properties” in the experiment list

§ Properties can be exported as XML

§ Metadata are exported automatically with channel images

§ Complete experiment can be reproduced with “Apply settings” in the “Properties”-window

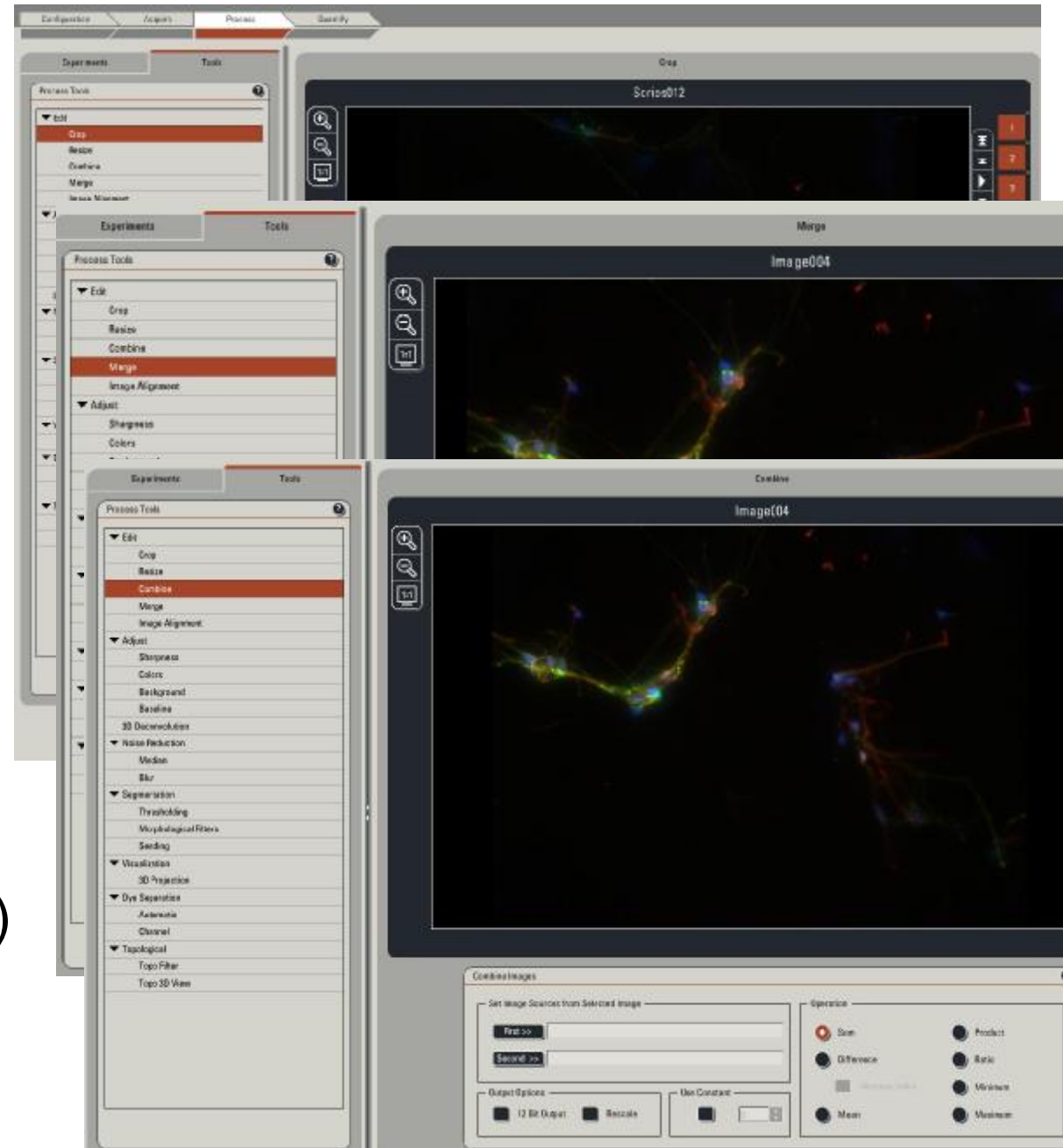


# How to make Life easy:

- § Copy successful image set into an “Experiment” called “My templates”
  - § Each user can open this experiment, select the appropriate data set and go to “Properties – Apply settings”
  - § Appropriate settings are available, adjust exposure time, objective lens, z-stack definition and start imaging
  - § Keep a reasonable list in definition of Channels and Settings
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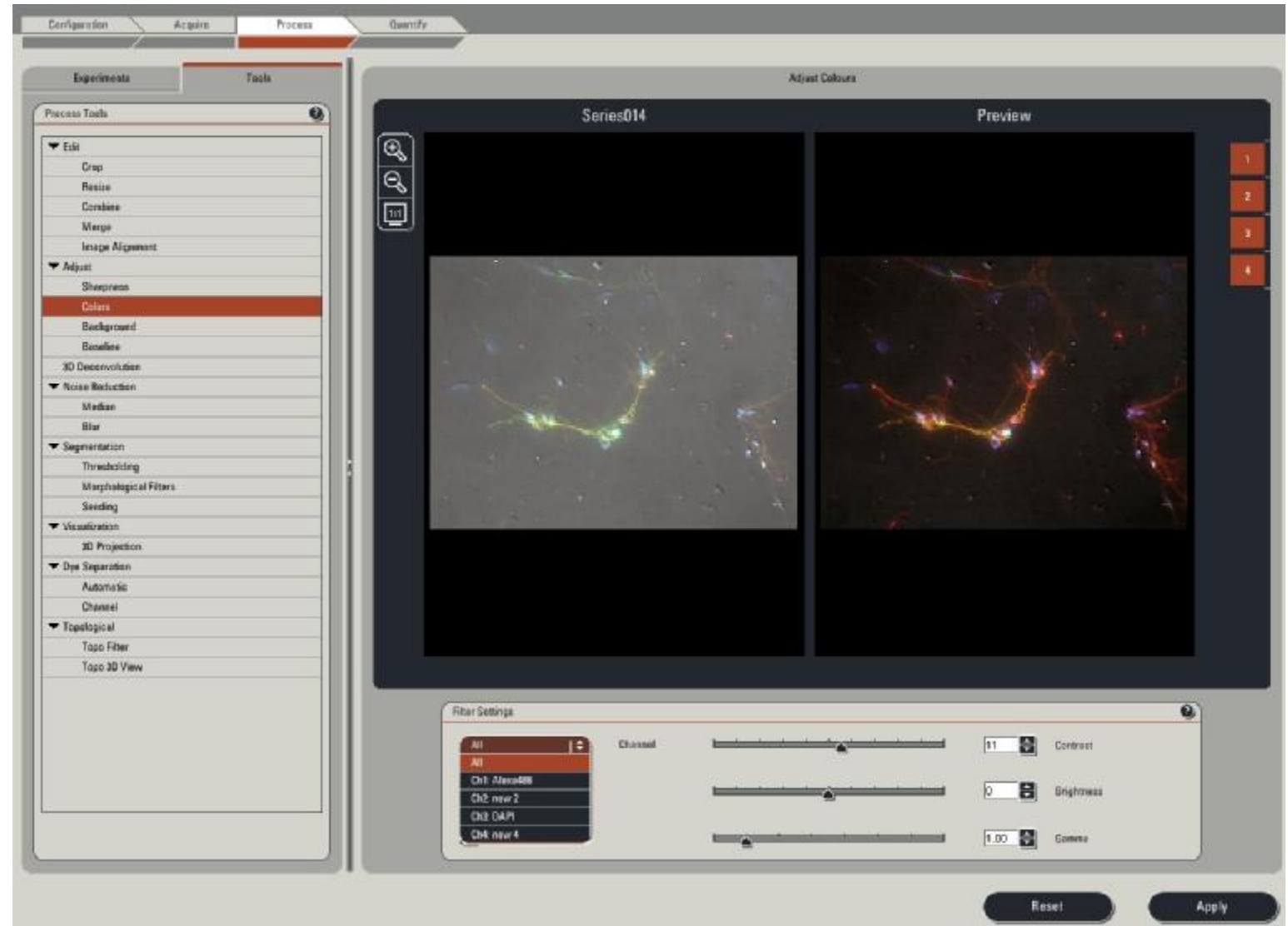
# Process:

- § Crop: Select subsets (ROI, z-levels, time points, channels)
- § Merge: Glue together (channels, time points, z-positions, in X or in Y)
- § Combine: Arithmetics
- § Adjust Colours: Mixing the channels (also included in LASAF lite)



# Process:

- § Adjust Colours:  
Mixing the channels  
(also included in LAS AF lite)

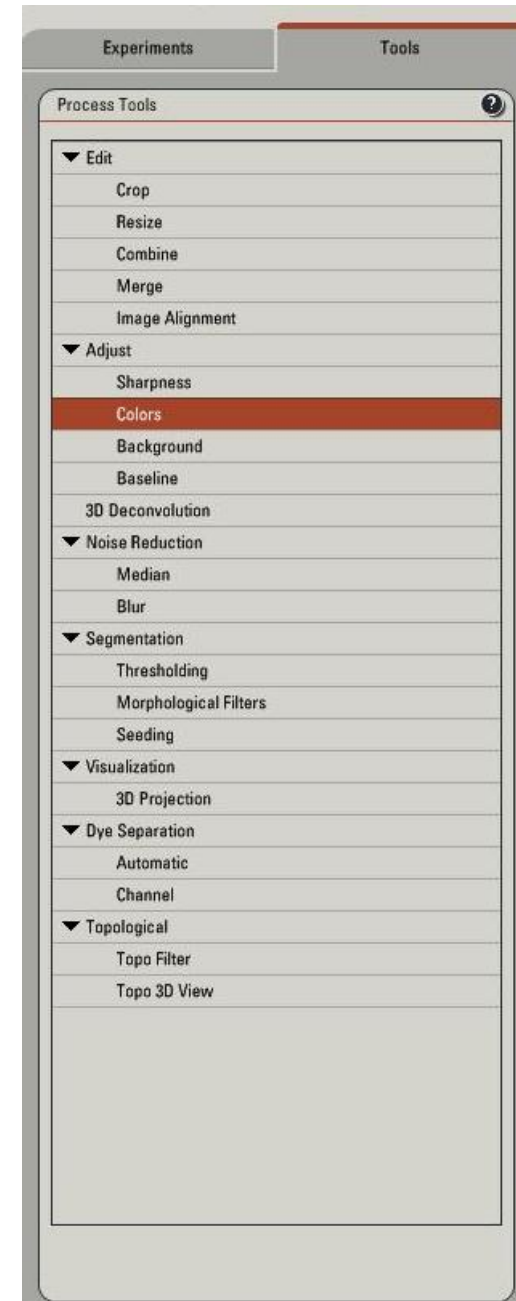


# Process:

- § Noise reduction
- § Sharpness
- § Background subtraction
- § .....

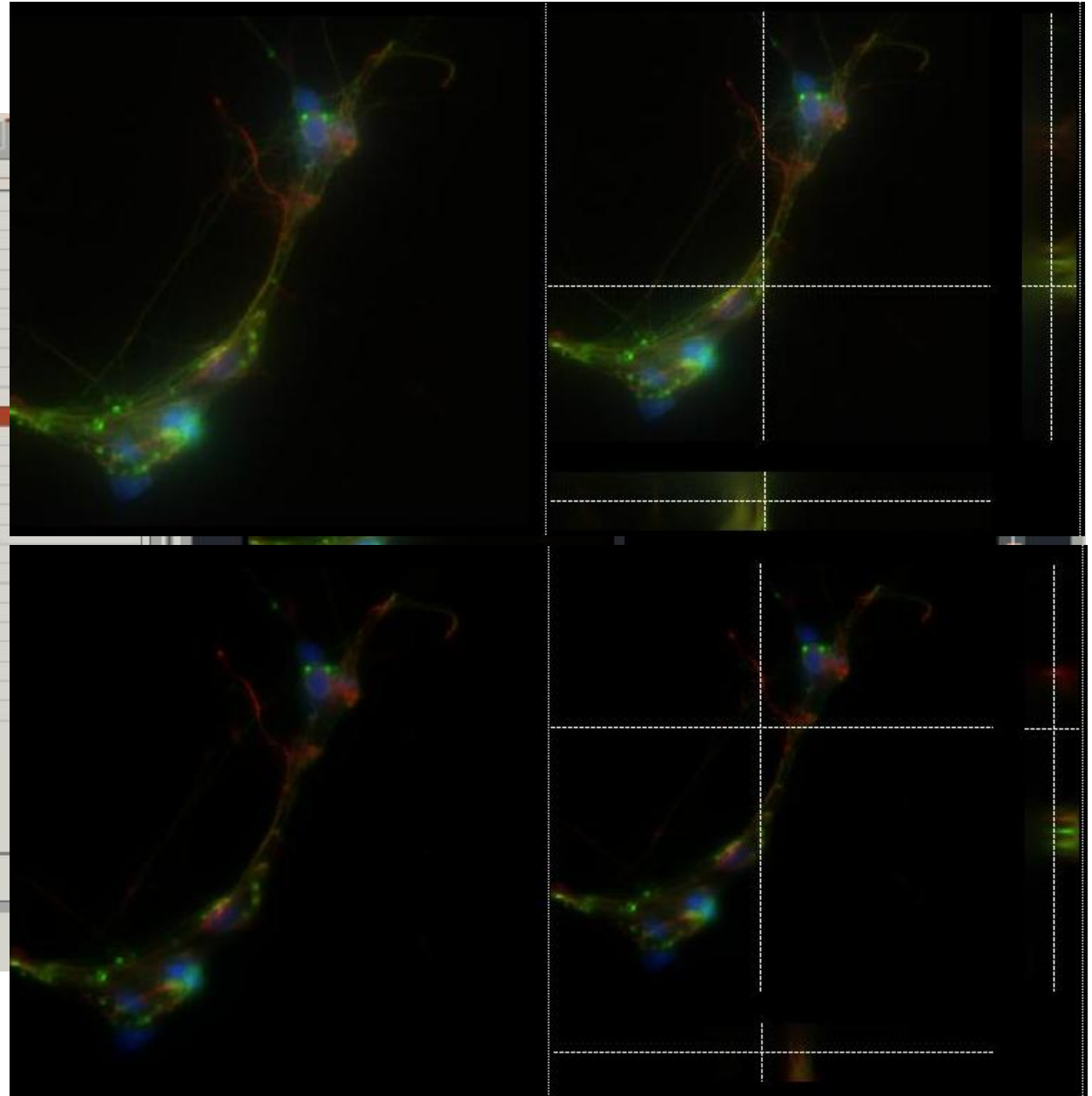
## Optional:

- § Deconvolution
- § 3D Visualization
- § Dye Separation



# Deconvolution:

- § All data automatically loaded
- § Recommendation: Blind, 5 iterations, deselect "fast"





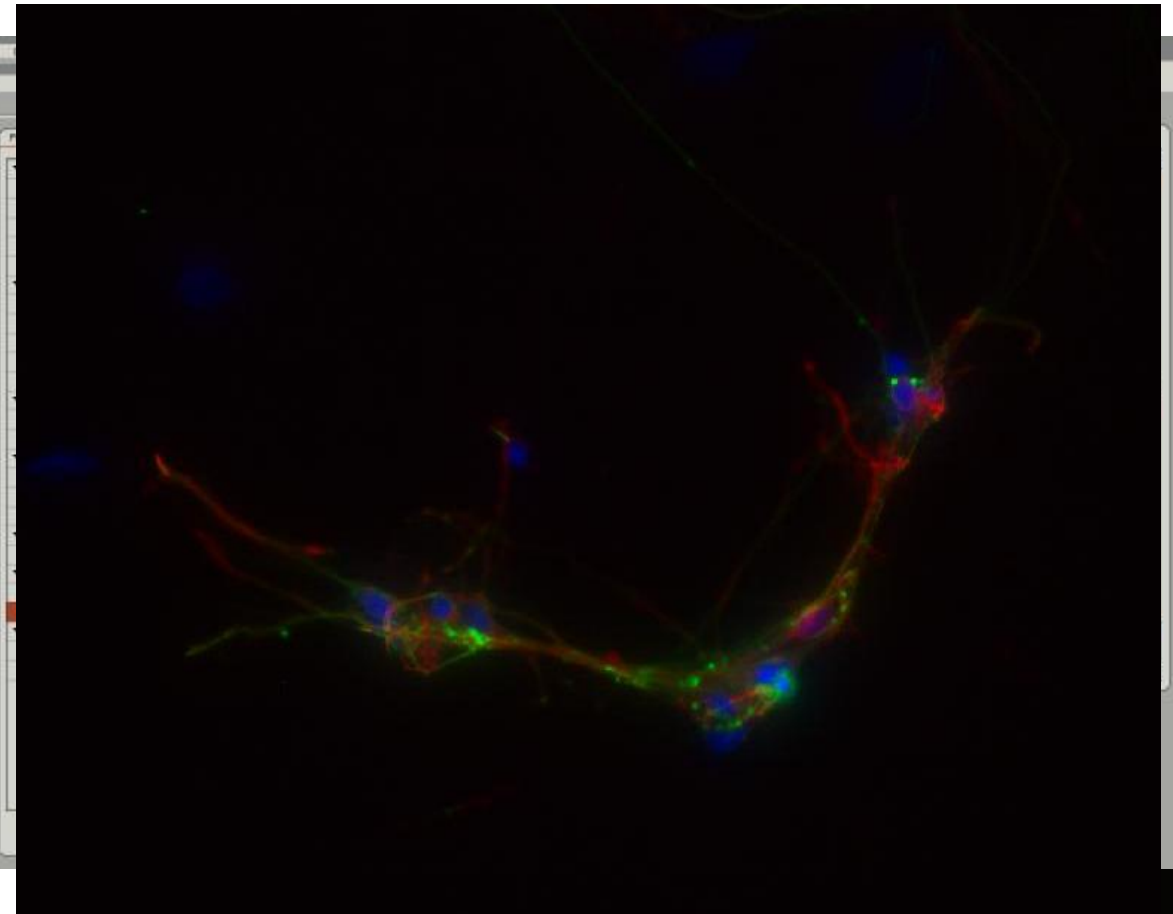
# 3D Visualization:

- § Create movies (tilting)
- § “colours” tool also for “projection series”



# Dye Separation:

- § Correction of Cross talk
- § Select areas of one fluorochrome only

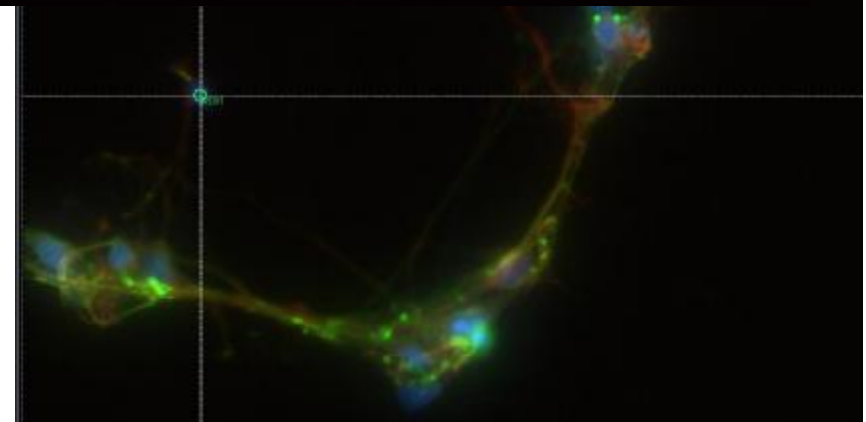


Edit Matrix

Matrix

	Ch1	Ch2	Ch3
Dye1	1	0	0
Dye2	0	1	0
Dye3	0	0	1

Reset Matrix Save Load Apply



# Quantification:

- § Profiles
- § Areas
- § Measurement “live”
- § Export as images or as Excel
  
- § No automatic detection !

