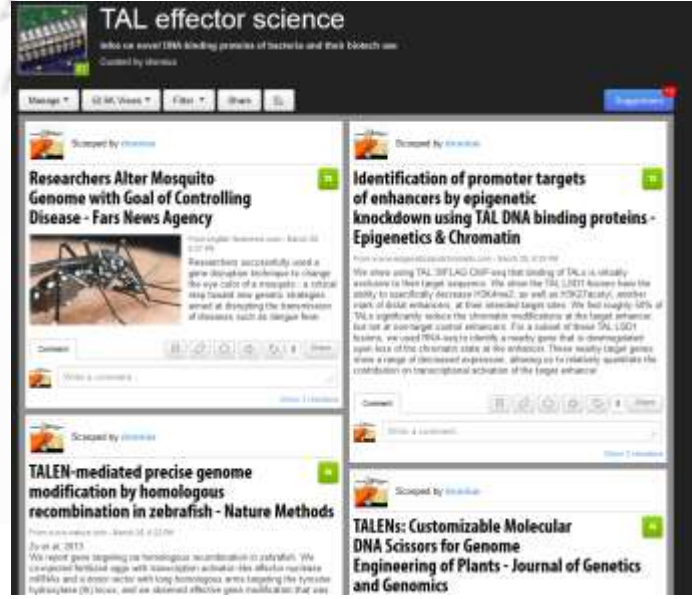


# Targeted variation of genomes using TAL effectors

Sebastian Schornack

<http://www.scoop.it/t/tal-effector-science>



Read:  
Schornack et al (2013) Annu Rev Phytopathol. 51:18.1–18.24  
Vojtas (2013) Annu. Rev. Plant Biol. 64:327–50  
Boch Bonas (2010) Annu Rev Phytopathol. 48:419-36

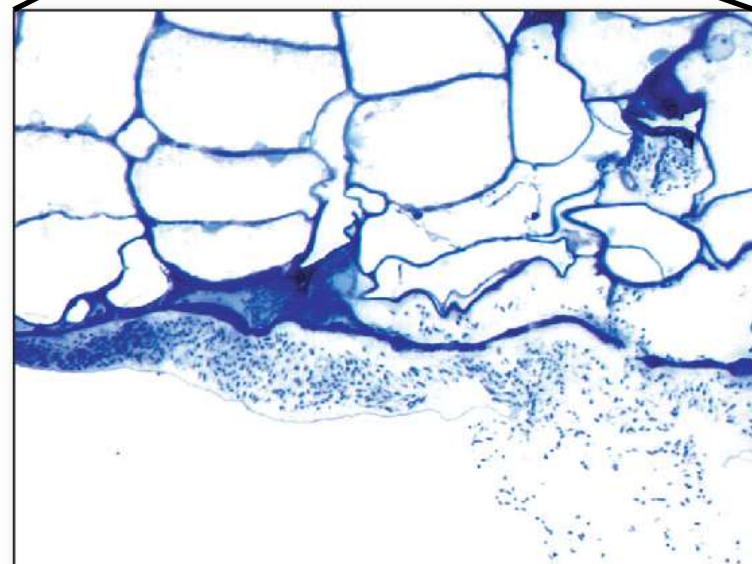
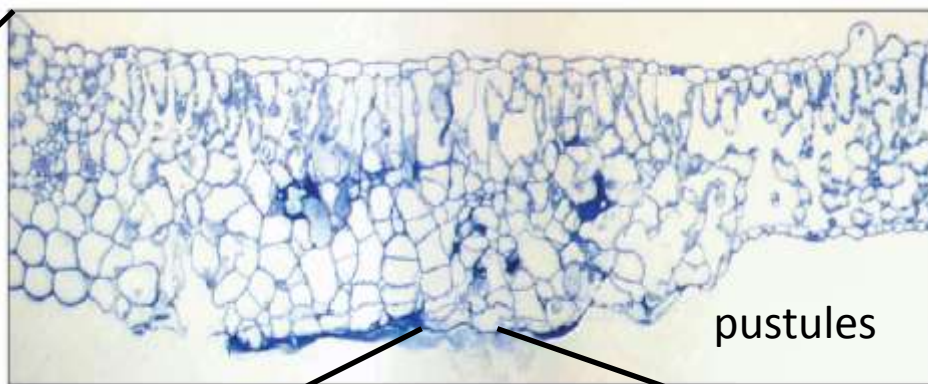
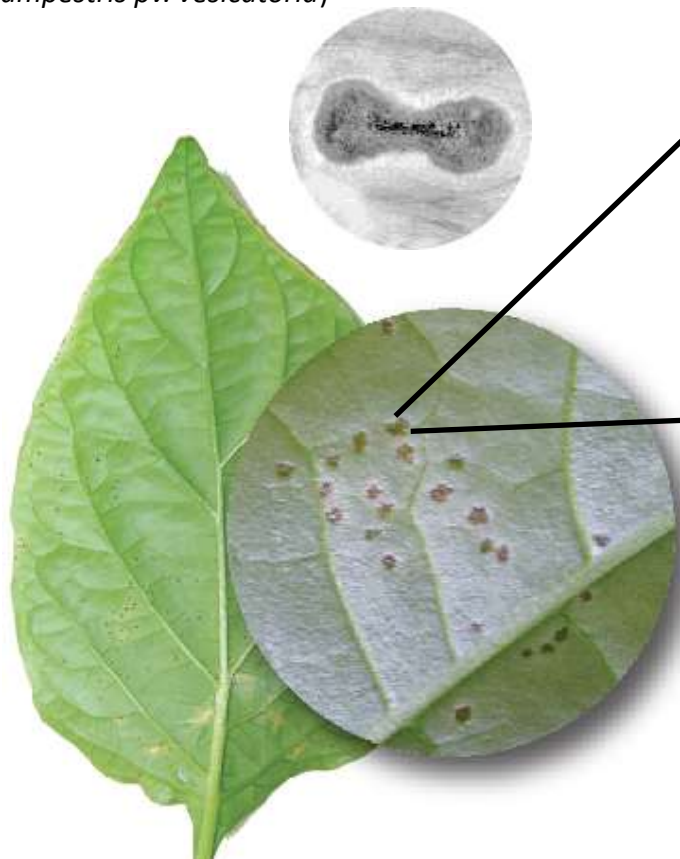
# Humans modify the genomes of animals and plants



By application of mutagens (chemicals, irradiation) and selective breeding

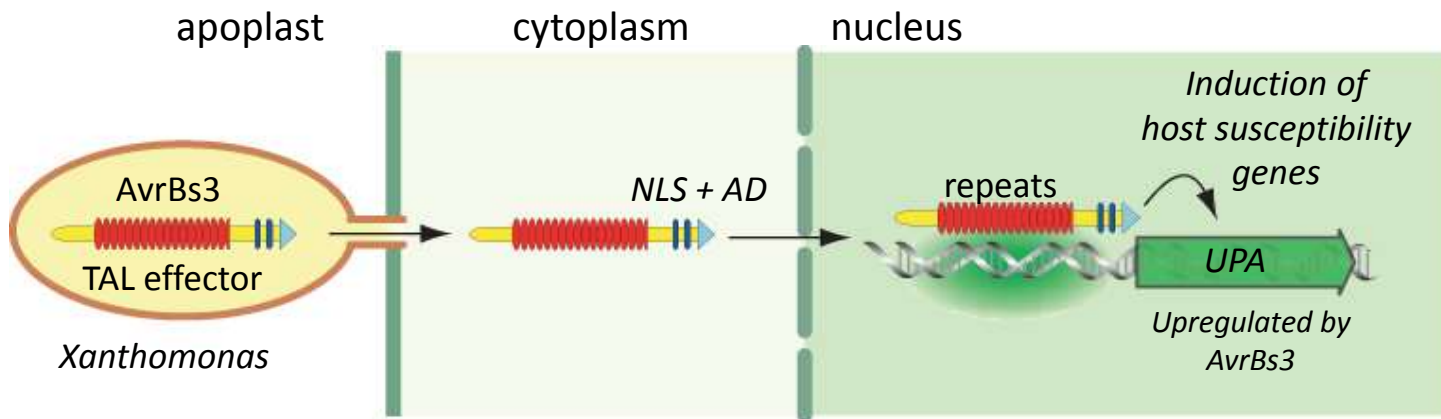
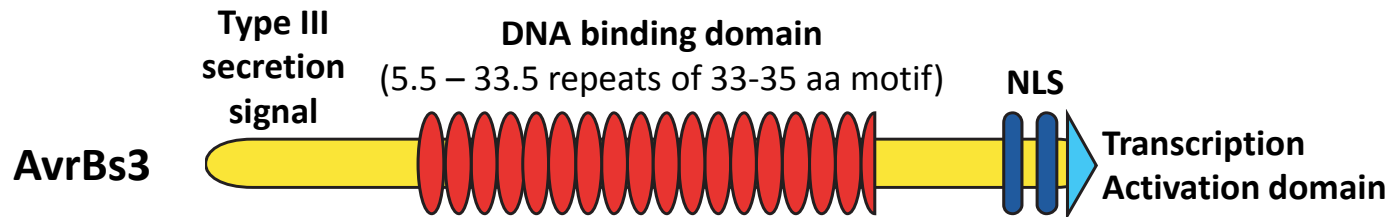
# *Xanthomonas* bacteria reprogram plant cells to grow

*Xanthomonas euvesicatoria*  
(*X. campestris* pv. *vesicatoria*)



Bacterial spot disease in pepper

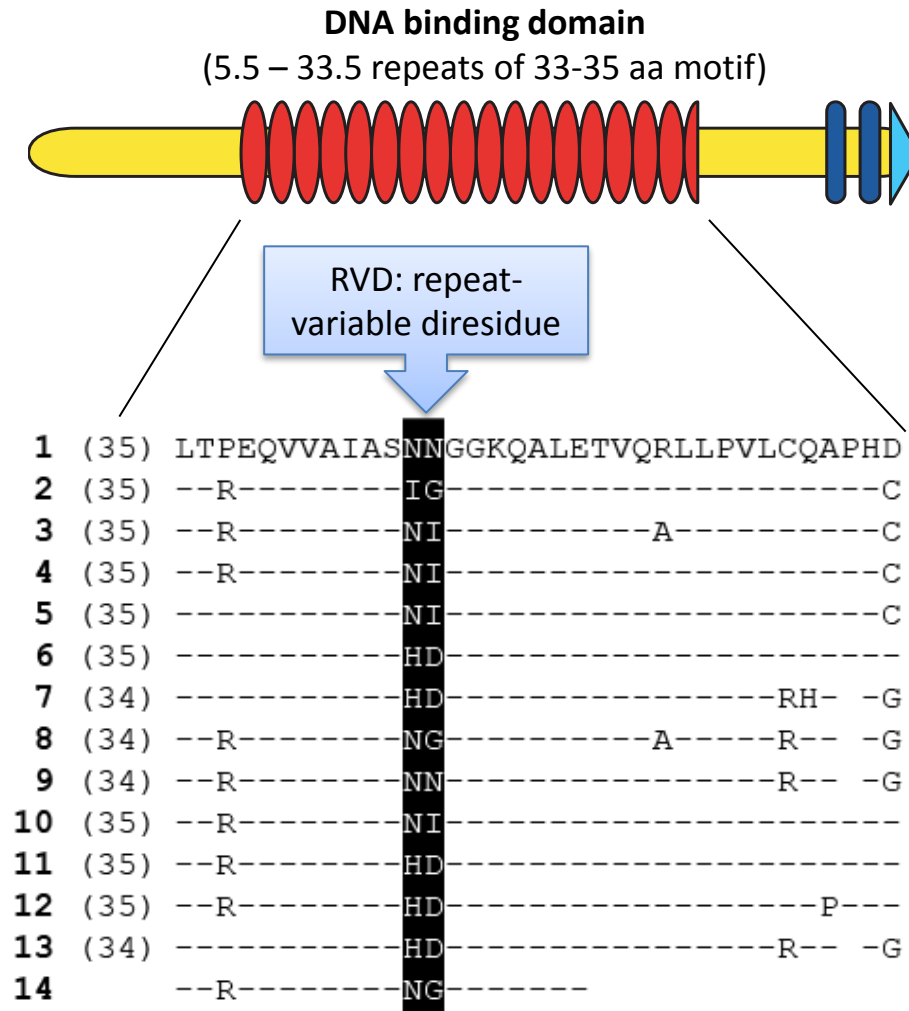
# TAL effectors = Transcription Activator-Like effectors



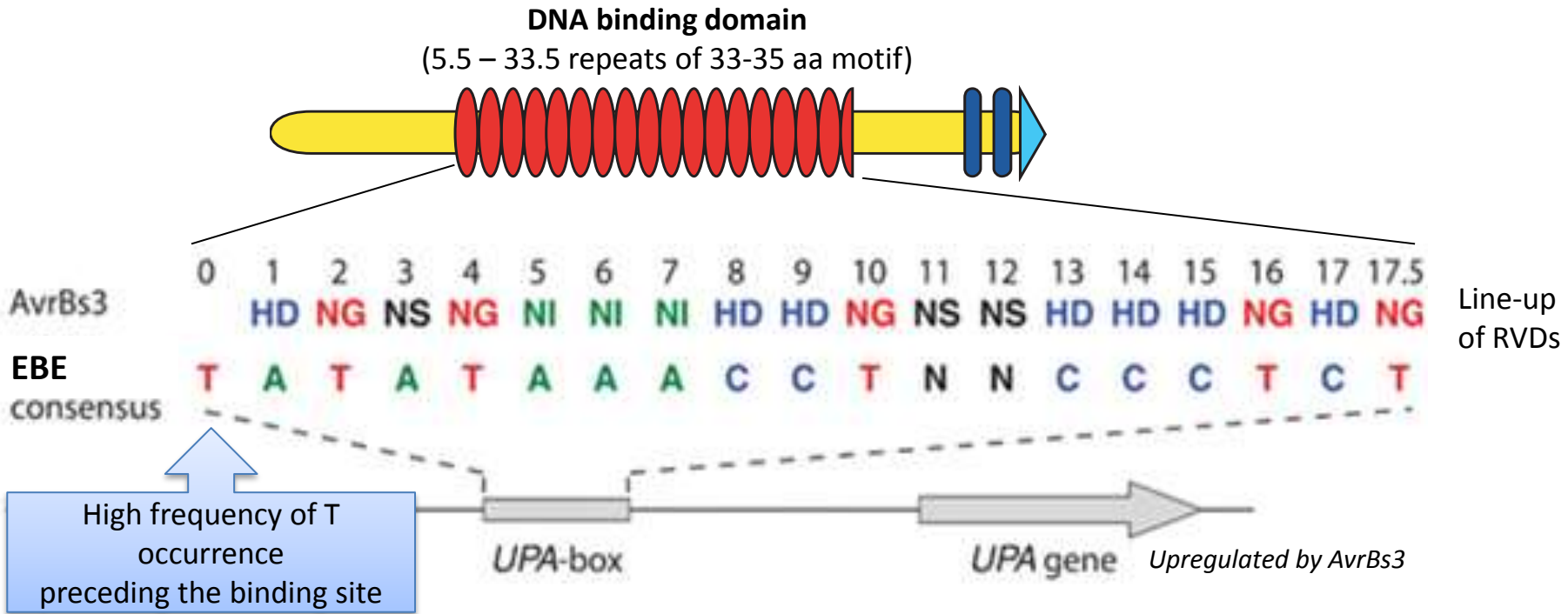
Bind promoter sequences in the host plant

Activate the expression of plant genes that aid bacterial infection.

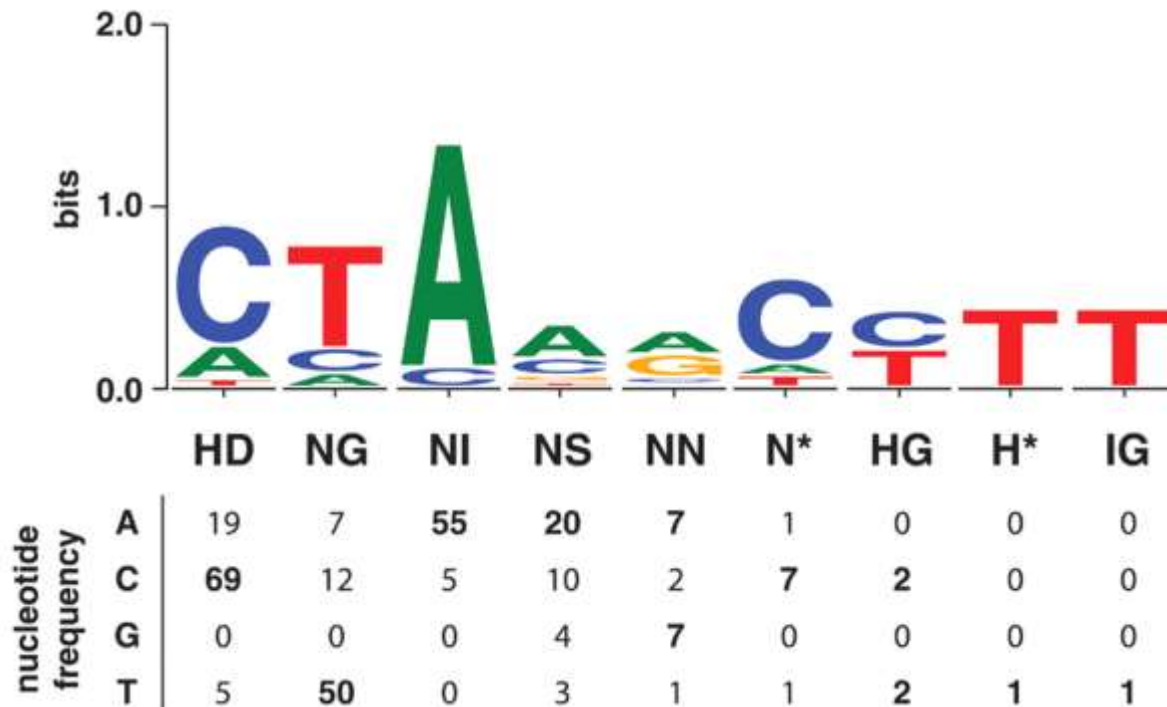
# TAL effectors - hypervariable 12<sup>th</sup> and 13<sup>th</sup> amino acids



# TAL effectors – linear RVD-DNA base relationship



# TAL effectors – The DNA binding code



NI = A

HD = C

HG = G

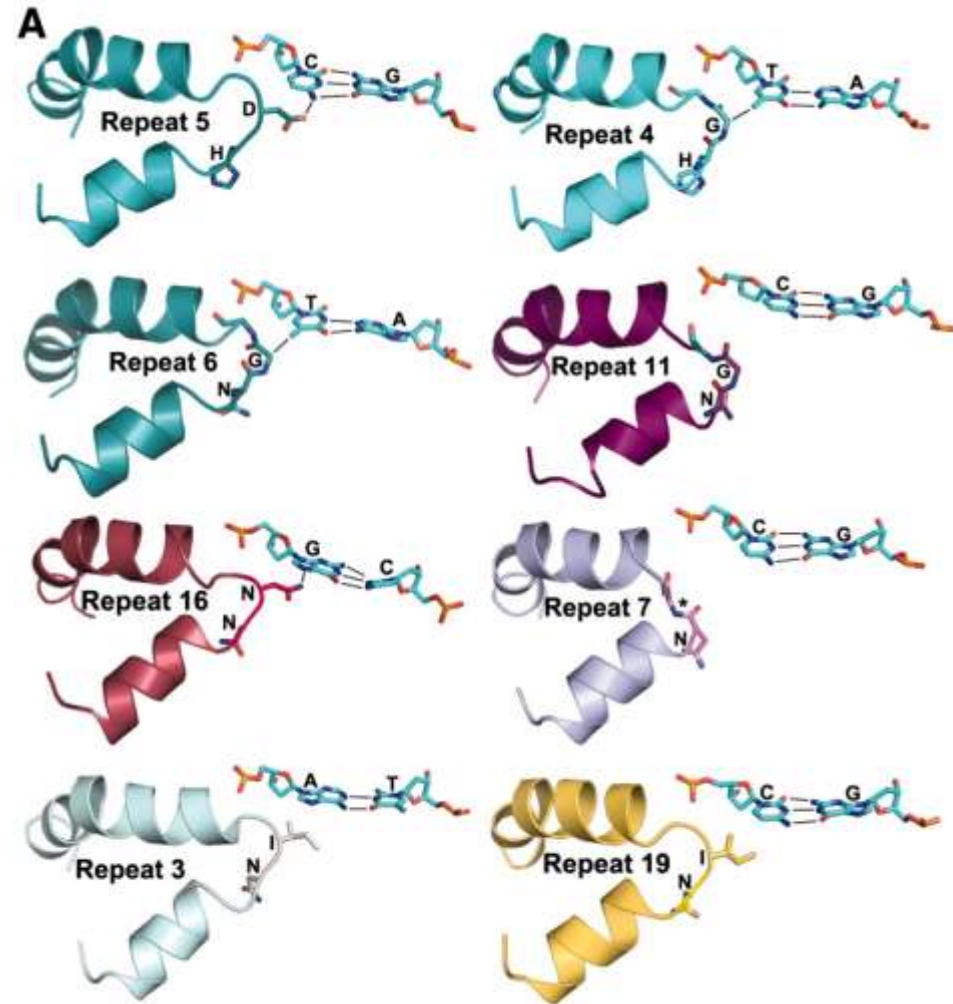
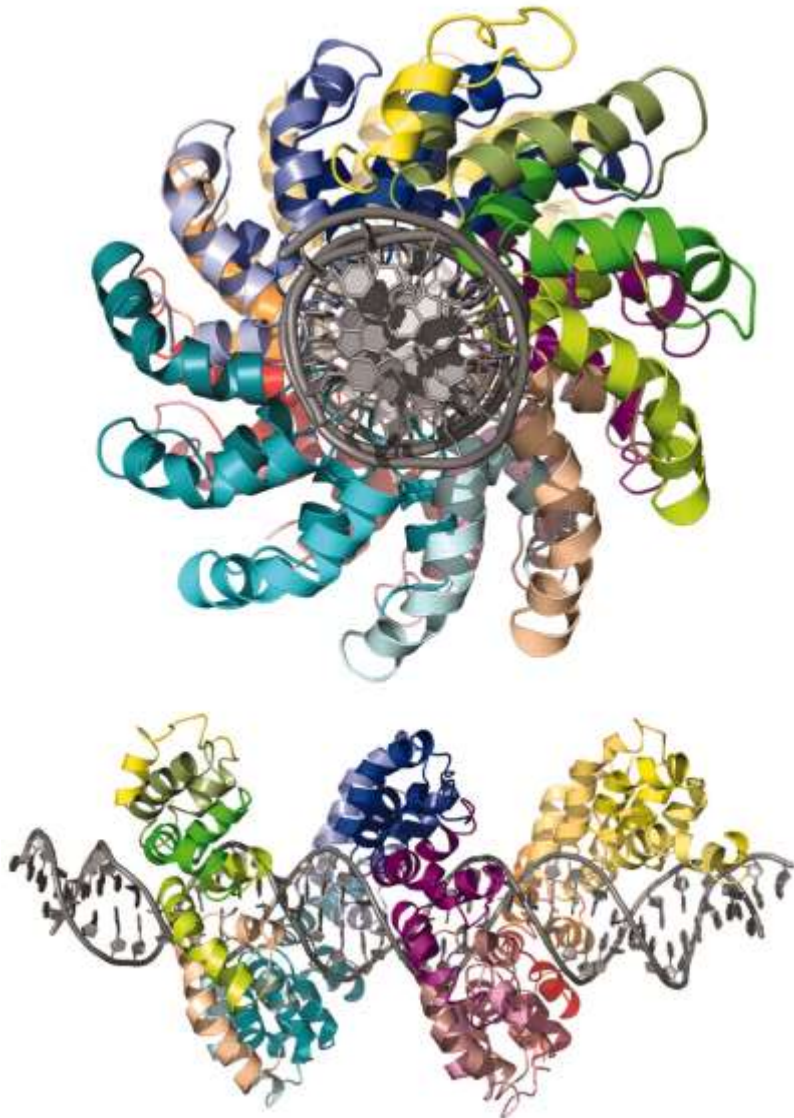
NG = T/mC

NN = G/A

NS = A/C/G/T

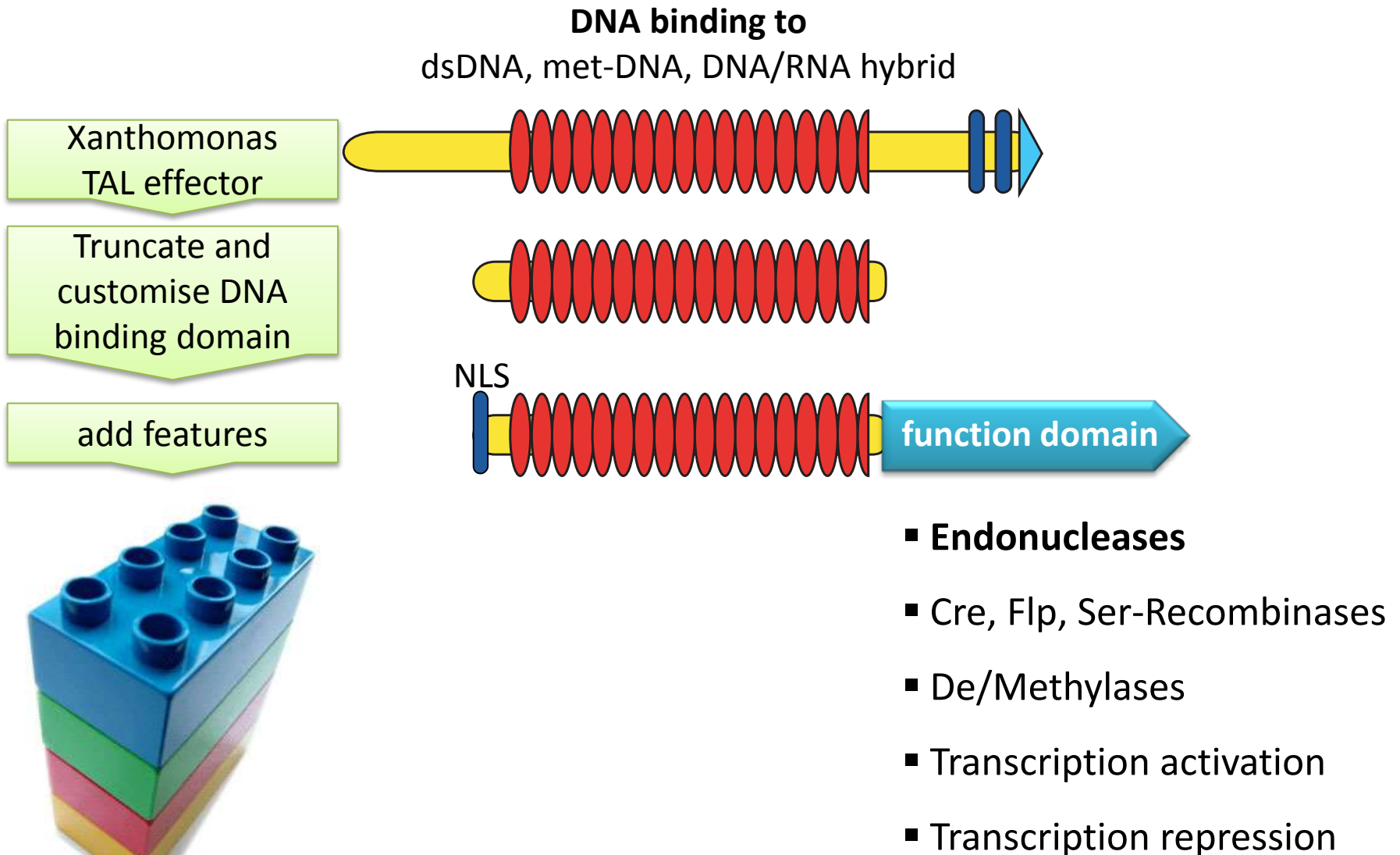
N\* = C/mC

# Repeats pair 1-on-1 with bases of the +DNA strand





# Customising the tool



# How to build/get a custom TAL effector?

## 1. Identify optimal DNA regions in a given target DNA sequence



TALeffectors.com

TAL effector nucleotide targeter - <https://boglab.plp.iastate.edu/>  
 Collectis search tool: <http://talen-hit.collectis-bioresearch.com/search>  
<http://baolab.bme.gatech.edu/Research/BioinformaticTools/assembleTALSequences.html>

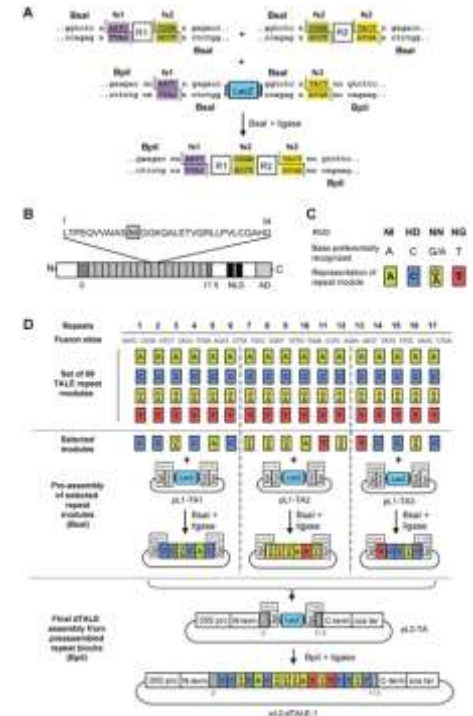
[http://www.talendesign.org/mojohand\\_main.php](http://www.talendesign.org/mojohand_main.php)

## 2a. Build it from single repeats using available modular cloning kits [Golden Gate cloning / Cut-ligation cloning]



Bogdanove/Voytas Kit  
 Joung Kit  
 Zhang Toolbox

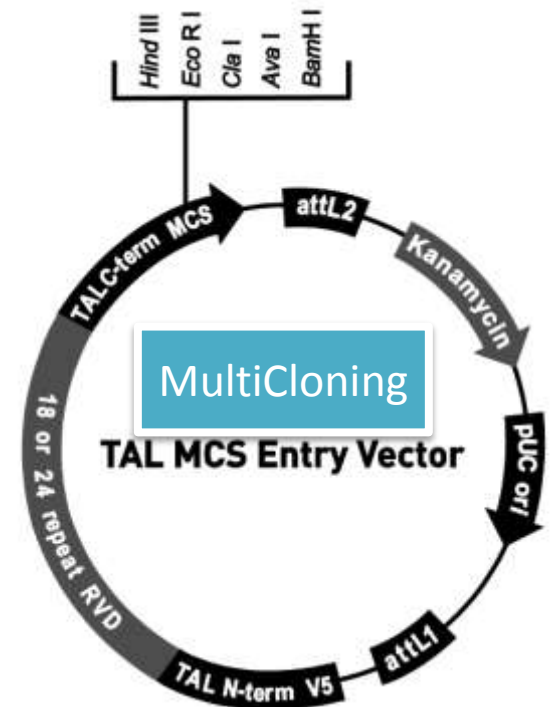
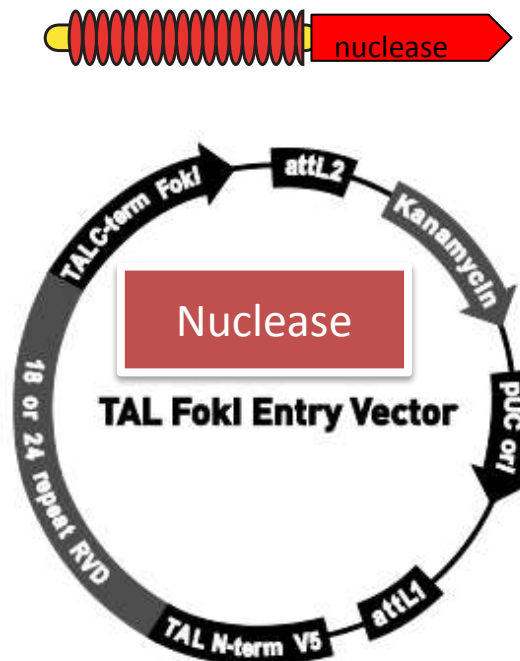
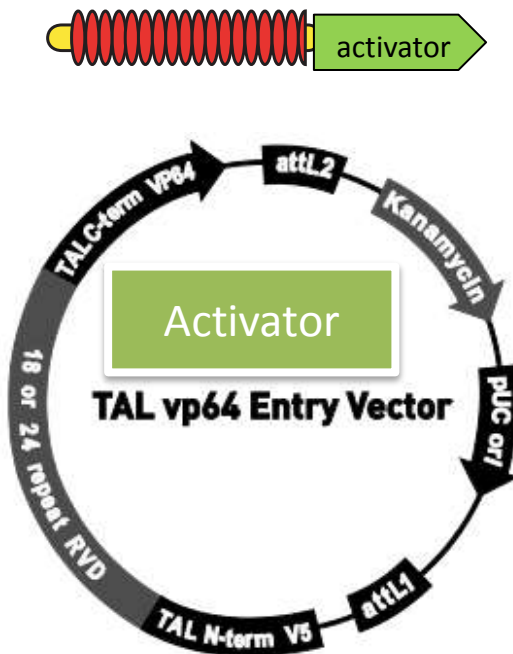
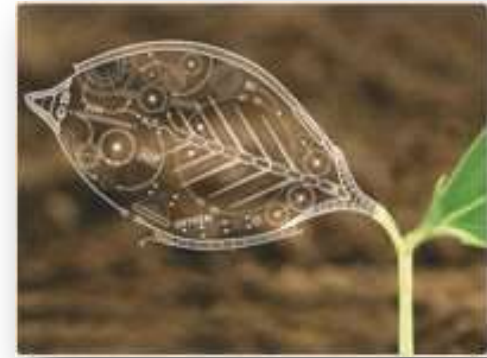
Lahaye Kit  
 Boch Kit



# How to build/get a custom TAL effector?

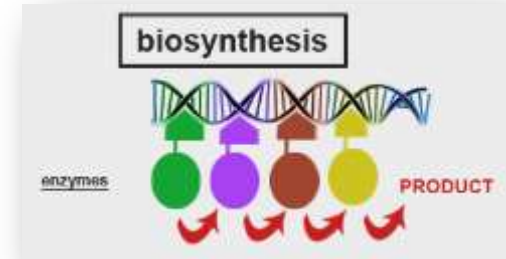
## 2b. Order it from a company

Life Technologies/GeneArt® TALs



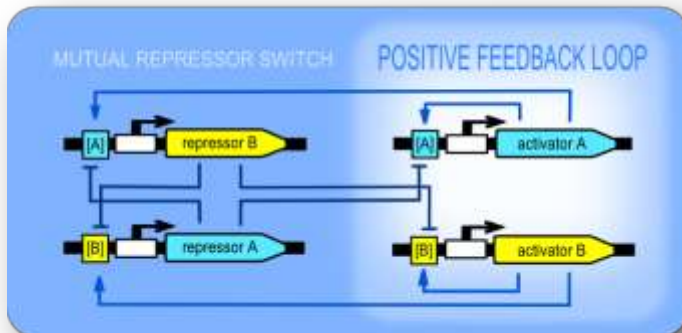
# Applications

DNA binding domains to tether enzyme cascades  
(iGEM team Slovenia with ZFN)



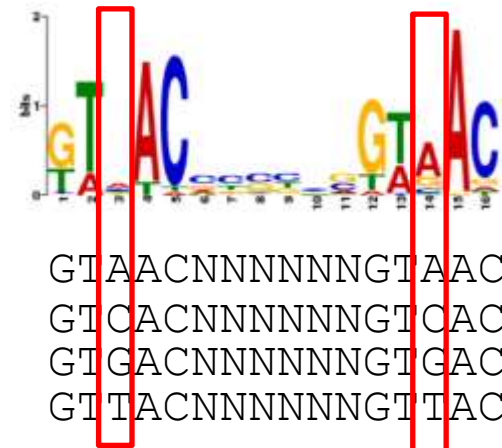
<http://2010.igem.org/Team:Slovenia/PROJECT/introduction>

Synthetic signal switches/circuits

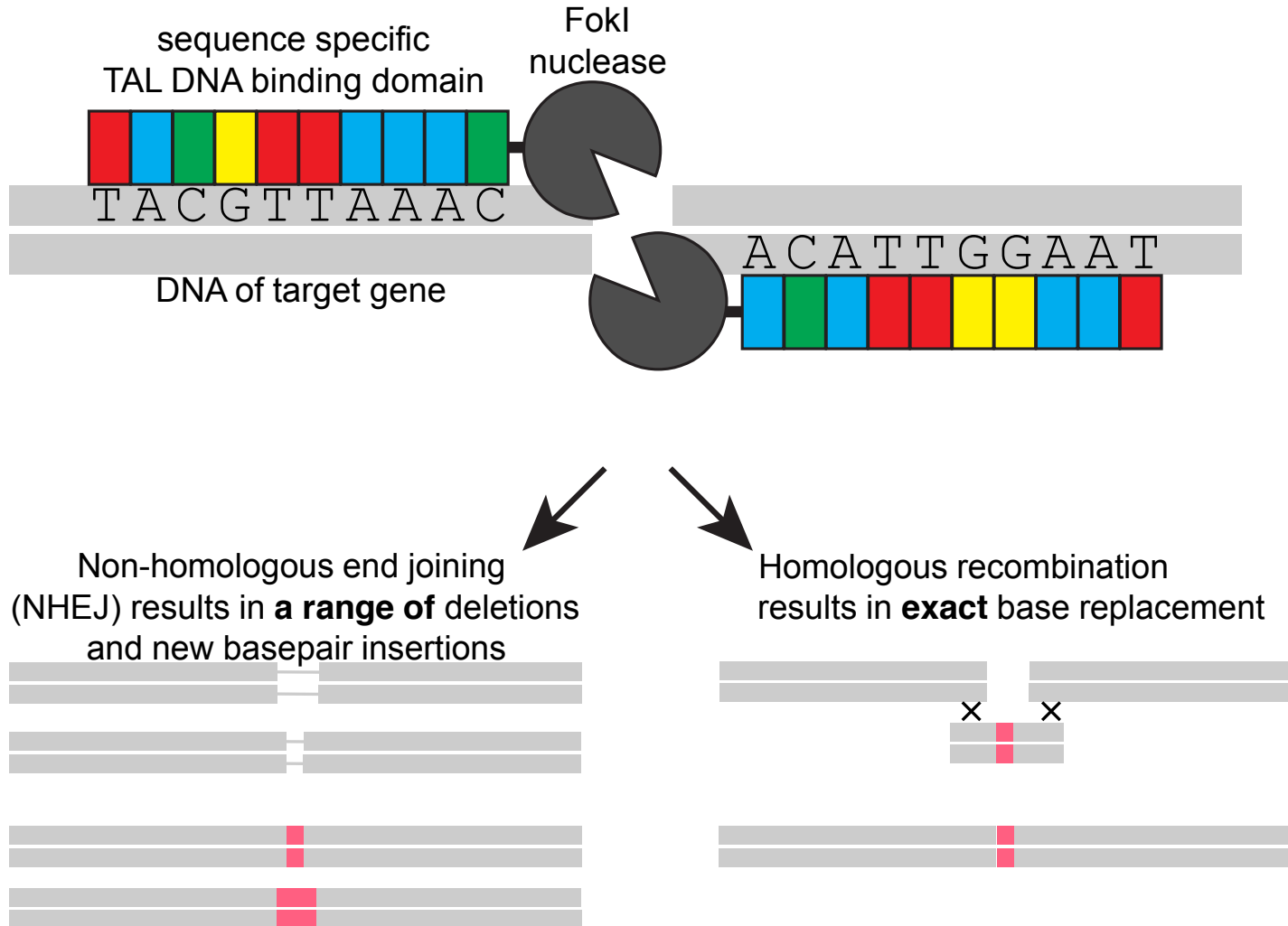


<http://2012.igem.org/Team:Slovenia/TheSwitchPositiveFeedbackLoopSwitch>

Subsampling of transcript pools

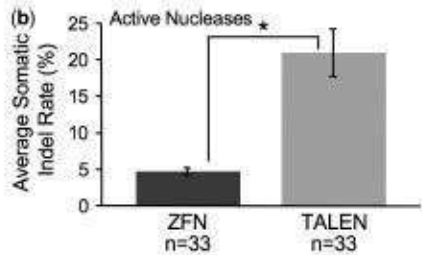


# TAL effector nucleases / TALENs



# TAL effector nucleases are customisable and specific

## Zinc finger nucleases



Chen et al, 2013, Nucl.AcidsRes.

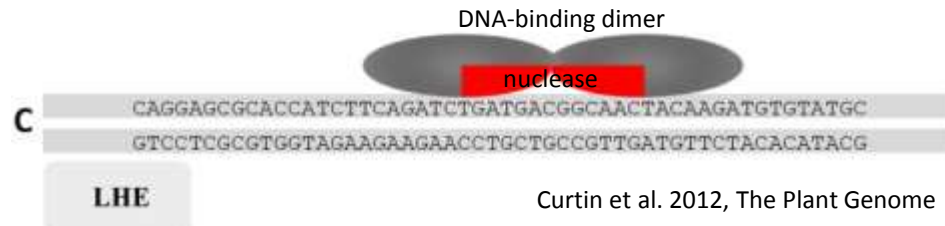
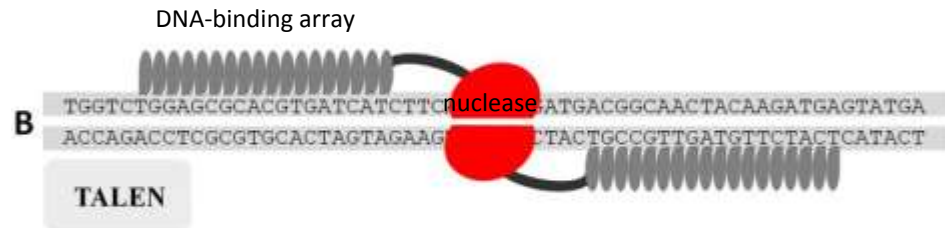
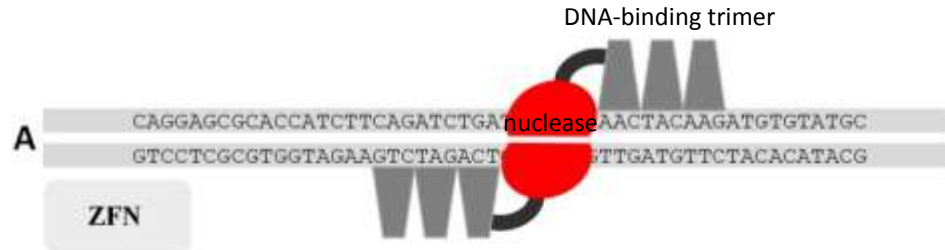
1-on-3  
Customisation  
requires specificity  
tests

## TAL effector nucleases

1-on-1  
Perfectly customizable

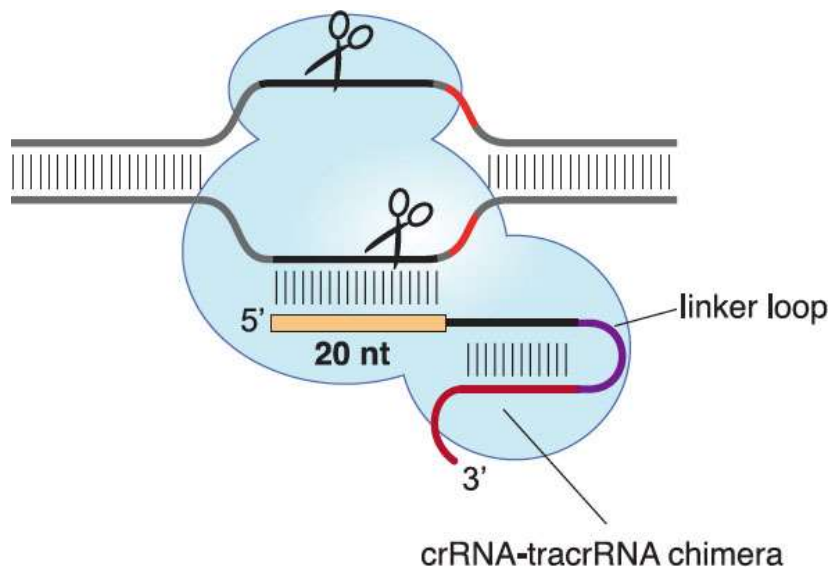
## Meganucleases/ Homing endonucleases

1-on-motif  
Complicated  
customization



Curtin et al. 2012, The Plant Genome

# A new kid on the block: CRISPR/Cas9 nuclease



- fast, easily programmed , inexpensive
- Requires that the target sequence is followed by a 3'-NGG (the so-called "protospacer-adjacent motif").
- limited to 12-bp off the RNA + GG PAM motif, so effectively ~14bp of specificity
- this enables unique targeting of ~40% of human genome

Advantage over ZFNs or TALENs → multiplexing. Simply add multiple guide RNAs.

## Unknowns:

Specificity for single site?

Cas9 fused to other domains possible?

Efficiency?

RNA scaffold differences between systems?

Sequence requirements within the Protospacer adjacent motif

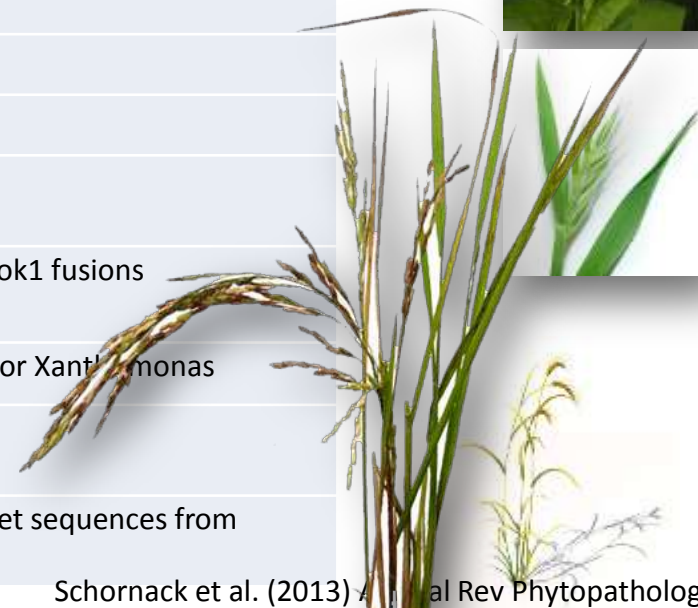
See also: [www.addgene.org/CRISPR/guide/](http://www.addgene.org/CRISPR/guide/)

# Scientific applications in plants

Successfully applied in bacteria, fungi, algae, higher plants, insects, frogs, fish, mouse, rat, pigs, cows

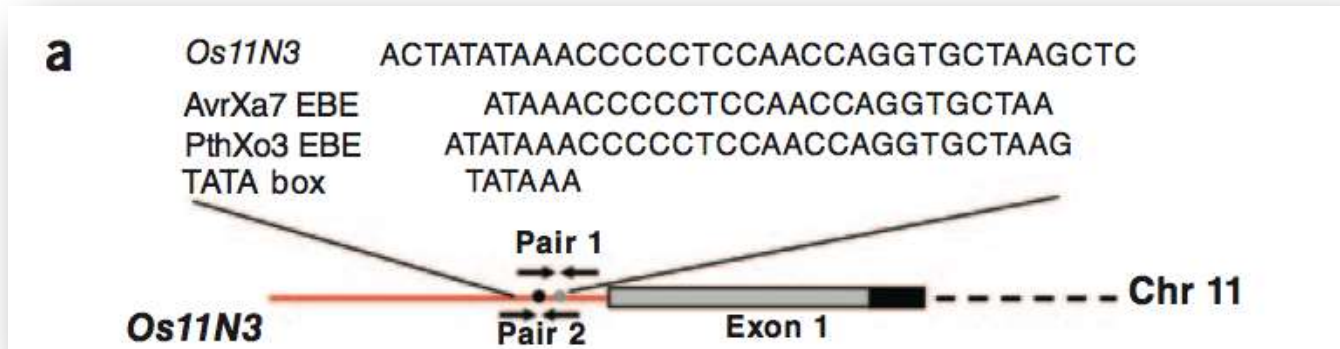
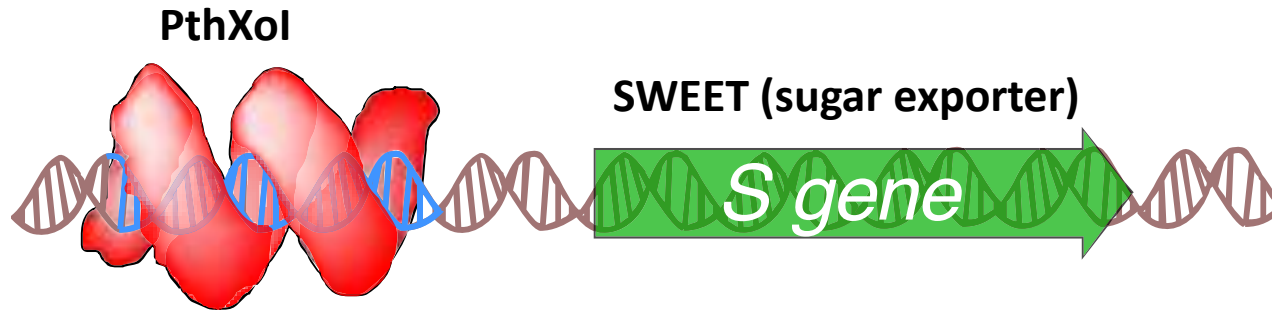
See also: <http://eendb.zfgenerics.org/een.php>

	Organism	Specific application
<b>Activator</b>	Arabidopsis	Transcriptional activation of endogenous KNAT1 and EGL3 genes with TALEs directed to their promoters
	Tobacco	Transcriptional activation of reporter gene
<b>Repressor</b>	Arabidopsis	RD29A promoters in reporter constructs and endogenous genes repressed with EAR repressor domain fusions
	Yeast	Repression of reporter genes
<b>Nuclease</b>	Arabidopsis	NHEJ-induced deletions with Fok1 fusions
	Tobacco	NHEJ based repair of inactive reporter gene
	Tobacco cells	NHEJ and HR based modification of ALS genes
	Brachypodium	NHEJ-induced modifications and large deletions with Fok1 fusions
	Rice	NHEJ based modification of a host susceptibility gene for Xanthomonas
	barley	NHEJ-induced deletions with Fok1 fusions
	Yeast	Repair of lacZ reporter gene constructs containing target sequences from various organisms



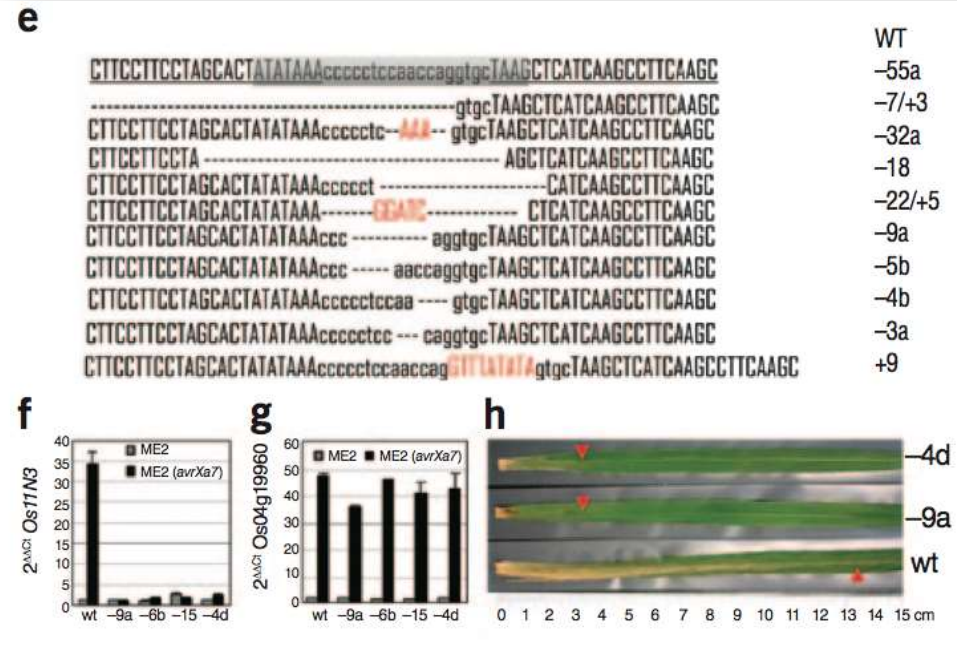
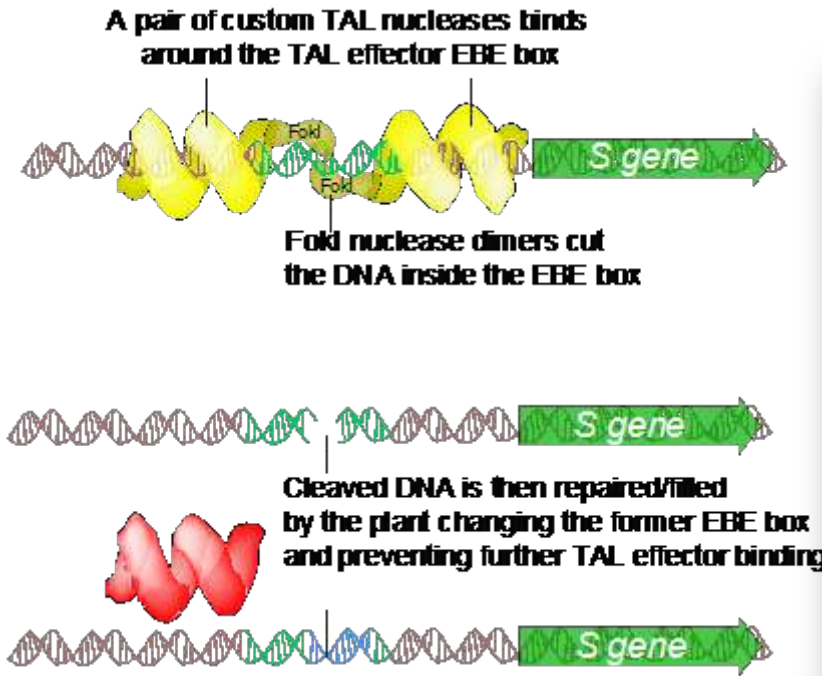


# Generation of disease resistant rice by removing DNA



Li et al. (2012) Nature Biotechnology

# Generation of disease resistant rice by removing DNA



Li et al. (2012) Nature Biotechnology

# Plants with broad *Xanthomonas* resistance

Synthetic EBE boxes in a decoy promoter trap for TAL effectors of multiple pathogens



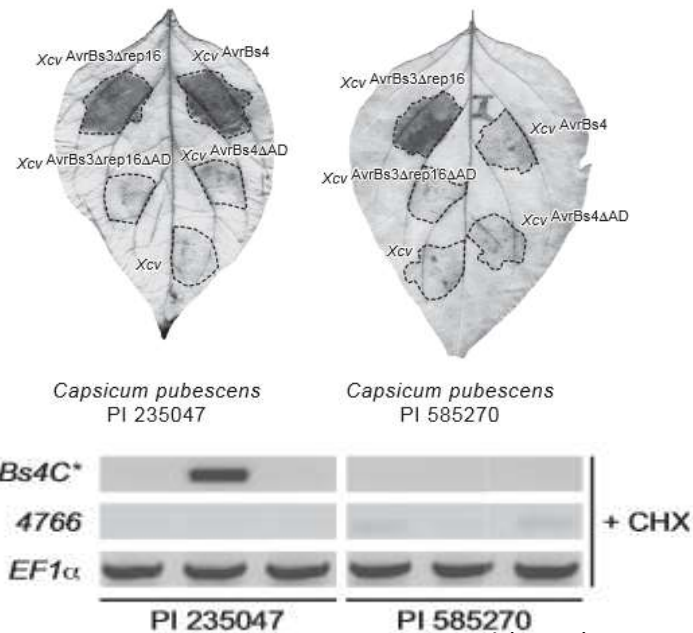
Schornack et al. (2013) Annual Rev Phytopathology

We need more executors (E genes)

Rice *Xa27*

Pepper *Bs3*

Pepper *Bs4P*



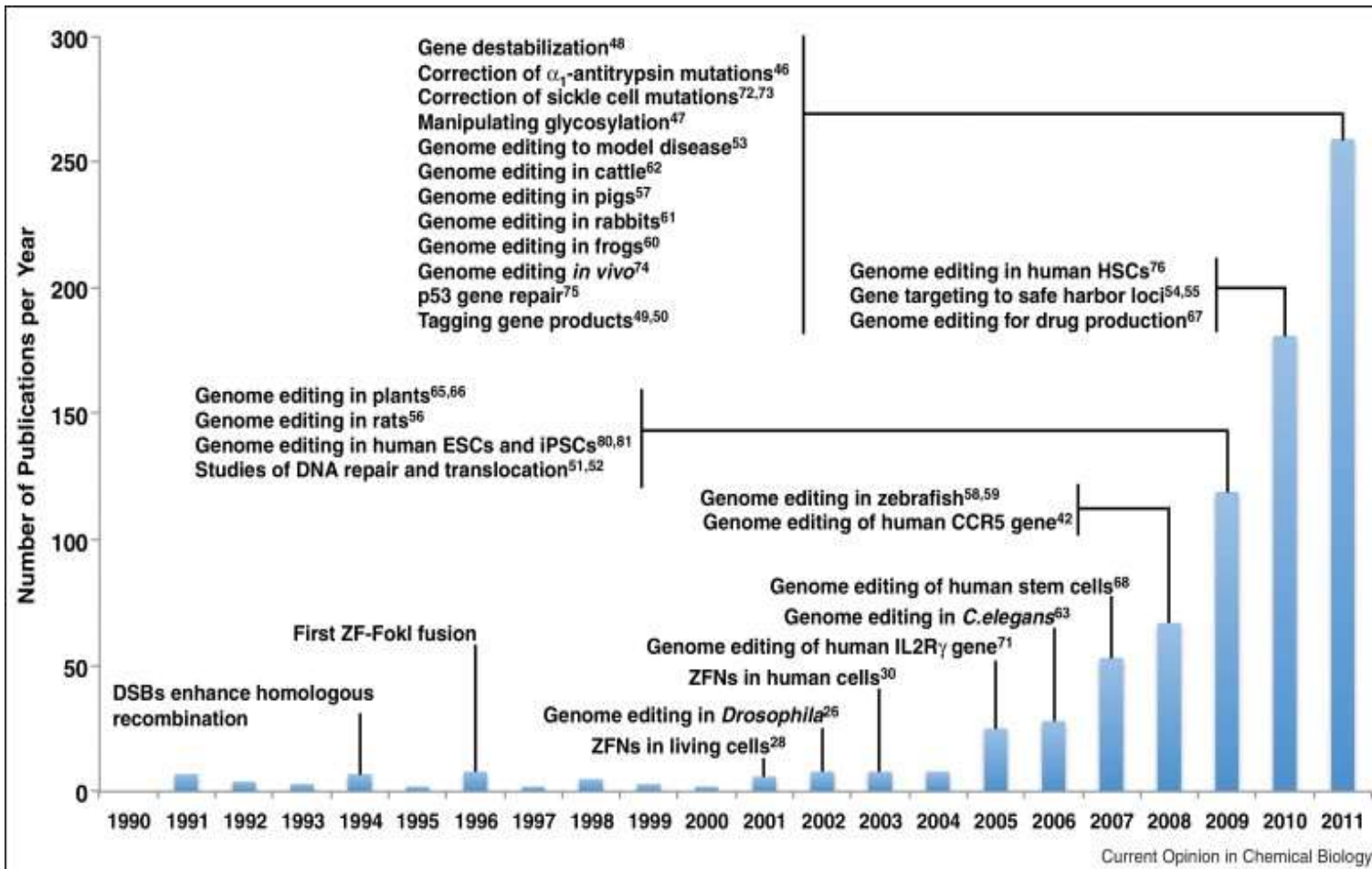
Strauss et al (2013), PNAS

# Genome editing – an accepted next-gen biotechnology?

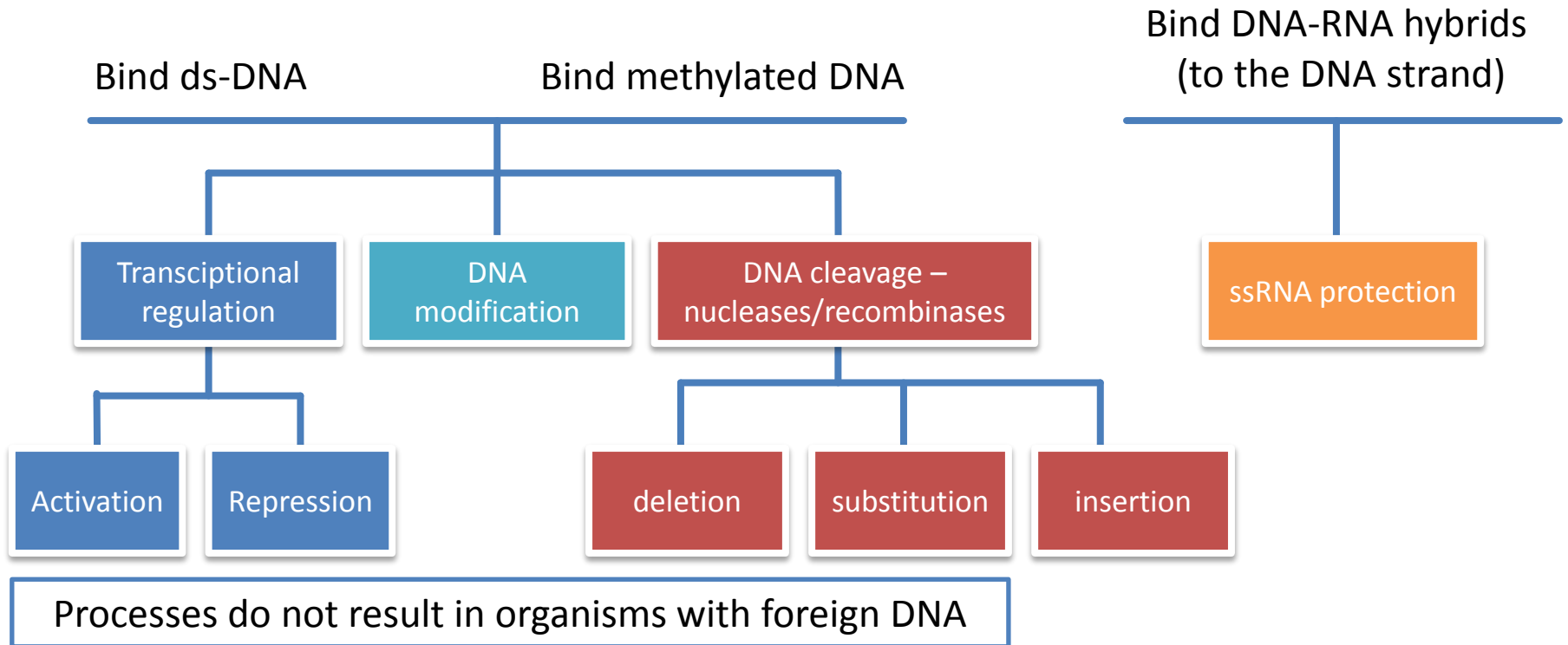


# Genome editing – an accepted next-gen biotechnology?

## Targeted variation: Site-specific alteration of DNA-anchored information



# Possibilities for targeted variation



# TAL effector technology – How will it be regulated?



## Position statement of the german ZKBS – June 2012

**A segment must contain at least 20 nucleotide pairs (NP) to lead to a recombinant nucleic acid.**

**An intentional change of less than 20 NP cannot be distinguished with sufficient confidence from random occurrence of this sequence.** Certain sequences of less than 20 NP [...] are indistinguishable from the genetic changes caused by conventional mutagenesis or natural mutation (random occurrence) (Cao et al., 2011).

The mutations induced by mutagenesis methods are according to § 3 no. 3b. Sentence 2 letter a GenTG (mutagenesis) **no genetic modifications.**

# Netherlands Commission on Genetic Modification (COGEM)

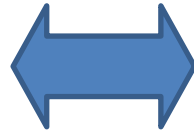


the issue of whether the modification process or the characteristics of the modification product should be the determining factor

## modification process

Process based legislation  
Europe

Product: GMO



## characteristics of the modification product

Product based legislation  
America, Canada, others

Product: Not GMO

### **Transcription factors do not involve genetic modification**

If ...introduced into a cell in protein form, there will be no alterations to the DNA sequence in the hereditary material ...There would therefore seem to be no genetic modification involved. Under this line of reasoning these activities do not fall under the EU directives.

Some medicines on the European market have a similar effect and do not fall under the GMO regulations. An example is Vidaza (azacitidine), which blocks the synthesis of DNA and RNA.

### **the use of [sequence specific] nucleases in protein form is largely comparable to the use of chemical mutagens**

the question is whether activities involving the use of nucleases in protein form should be covered by the GMO regulations, or not.



# Acknowledgements



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2Blades

H. Scholze, G. Minsavage, J. Jones, R. Stall, A. Bogdanove, V. Nekrasov, S. Kamoun, K. Peter, A. Strauss, P. Roemer, T. Jordan, D. Studholme

