



Microbial Adhesion and Molecular Interactions



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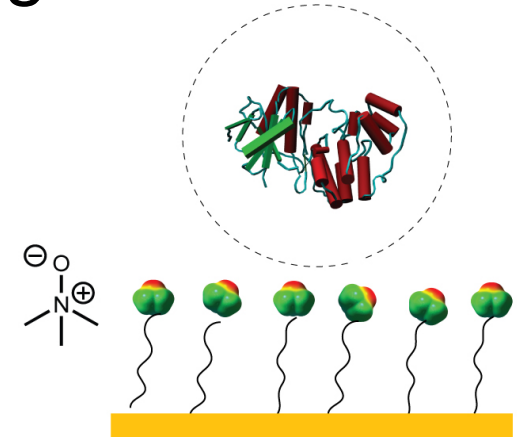
Bioelectrical Engineering Hub
Warwick

Overview

- SiO₂, PS, PEGA beads ... (phage-λ genomic display library)

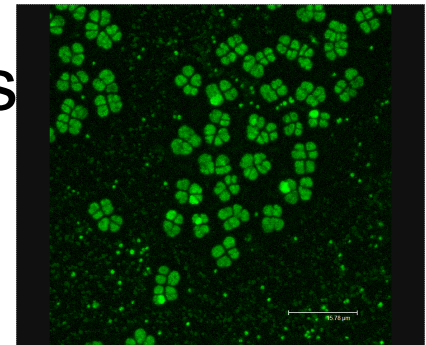
Langmuir, 2006, 22, 8144

Langmuir, 2013, 29, 2961



- Au, Si planar substrates with monolayers (lysozyme, fibrinogen)
- 1 μm microspheres, 100-200 μm beads (*Methanosarcina barkeri*)

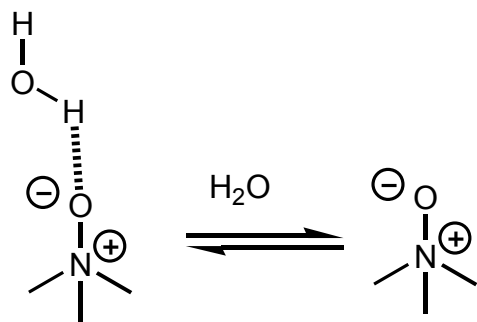
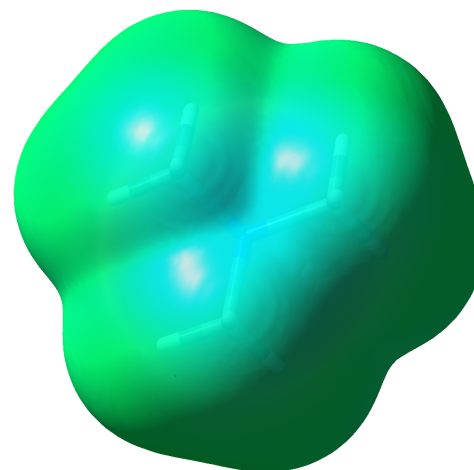
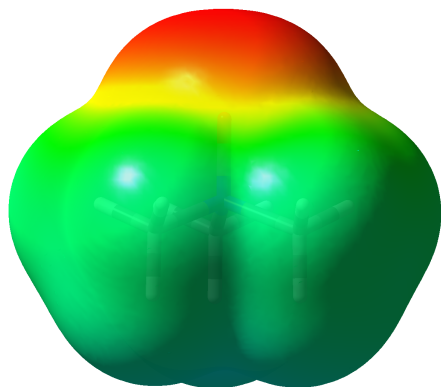
D Dobrzanska, *PhD Thesis*, University of Warwick 2014



- polyurethane foam (cattle slurry, fruit and veg anaerobic digester sampling)

D Dobrzanska, *PhD Thesis*, 2014

Trimethylamine *N*-oxide: a 'kosmotrope'



$\mu = 5.07$ D (calc.) 5.02 D (expt., benzene)
cf. 1.85 D for water, 0.61 D Me_3N

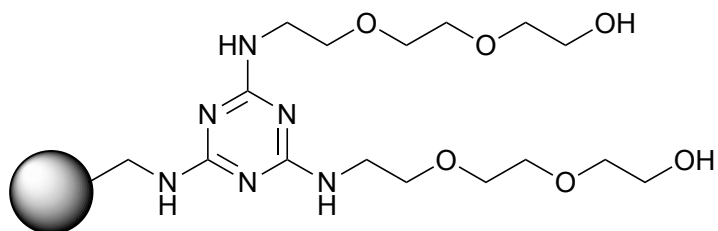
$\text{p}K_a$ ($\text{Me}_3\text{N}^+\text{O}-\text{H}$) = 4.5 (R P Bell, W C E
Higginson *Proc. Royal Soc.* **1949**, 197, 141)

$\text{p}K_a$ ($\text{Me}_3\text{N}^+-\text{H}$) = 9.76 (H K Hall, Jr. *J. Am.*
Chem. Soc. **1957**, 79, 5441).

Calculation performed at MP2/aug-cc-pVDZ level using *Gaussian03* T R Walsh

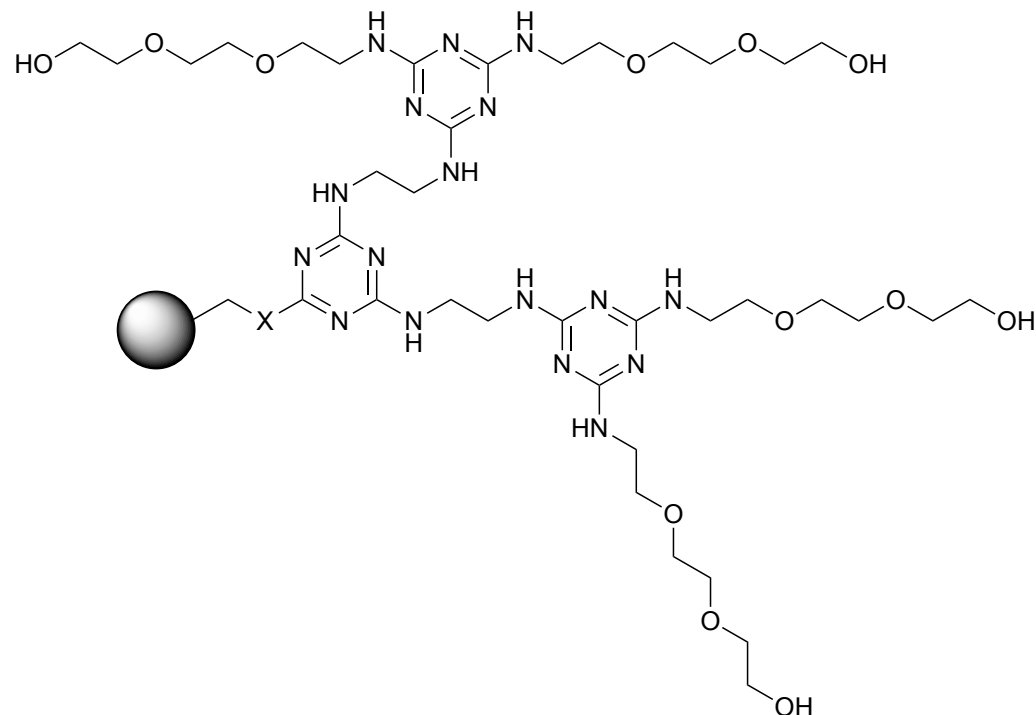
Counteracts denaturing effect of urea on proteins, stabilises macromolecular structure
D W Bolen, I L Baskakov *J Mol Biol* **2001**, 310, 955

New substrates prepared: dendrimeric OEG



1a SynPhase™ Lanterns

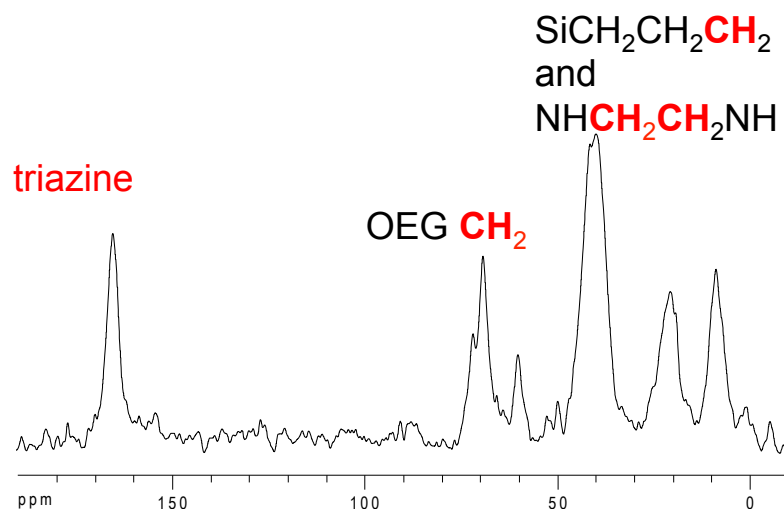
1b Aminopropyl silica



2a X = NH; SynPhase™ polyamide Lanterns

2b X = NH; Aminopropyl silica

2c X = O; Wang resin

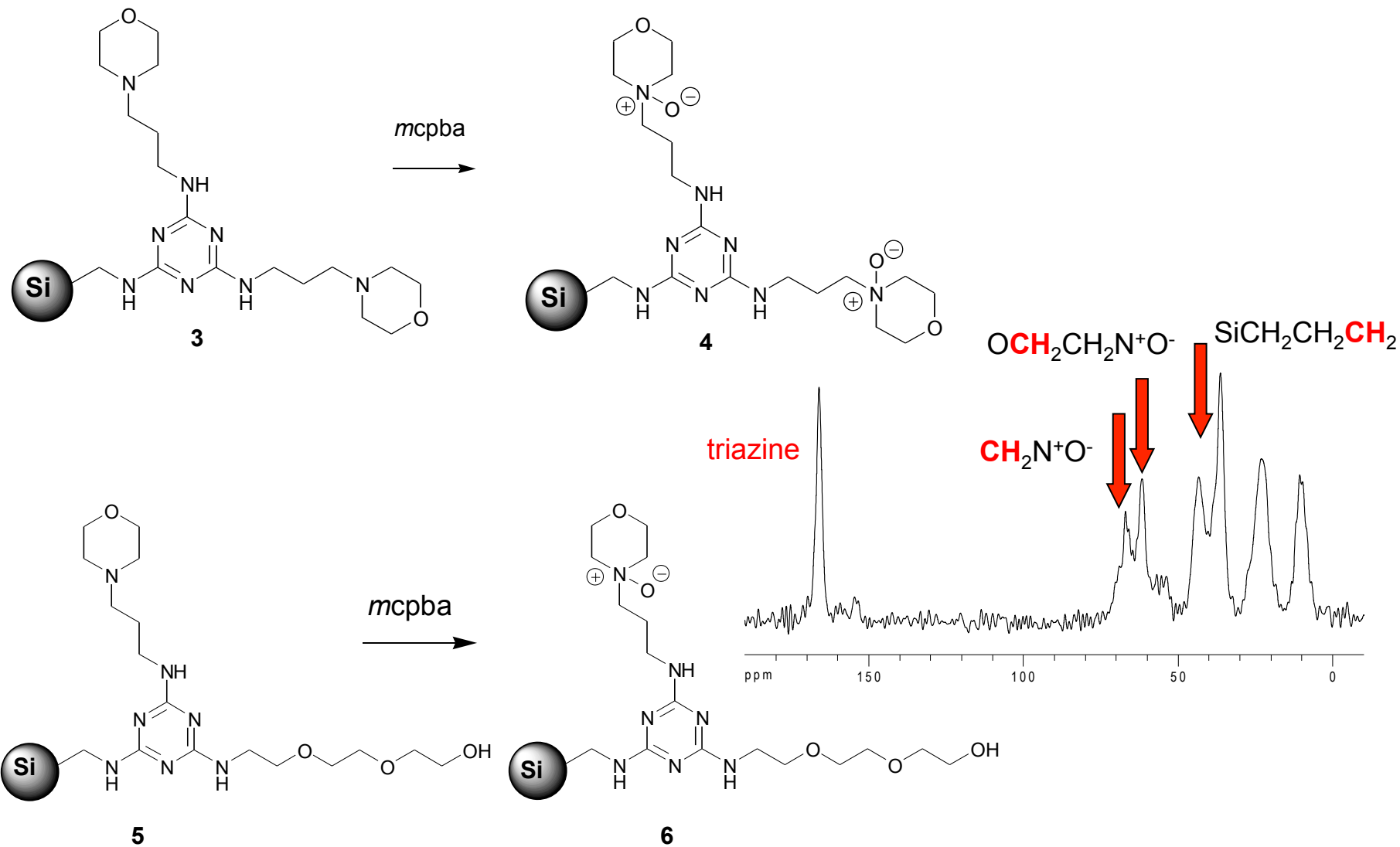


^{13}C CP-MAS NMR

S J Dilly, S J Carlisle, A J. Clark, A R Shepherd, S C Smith, P C Taylor, A Marsh *J Pol Sci: Pol Chem (Part A)* **2006**, *44*, 2248

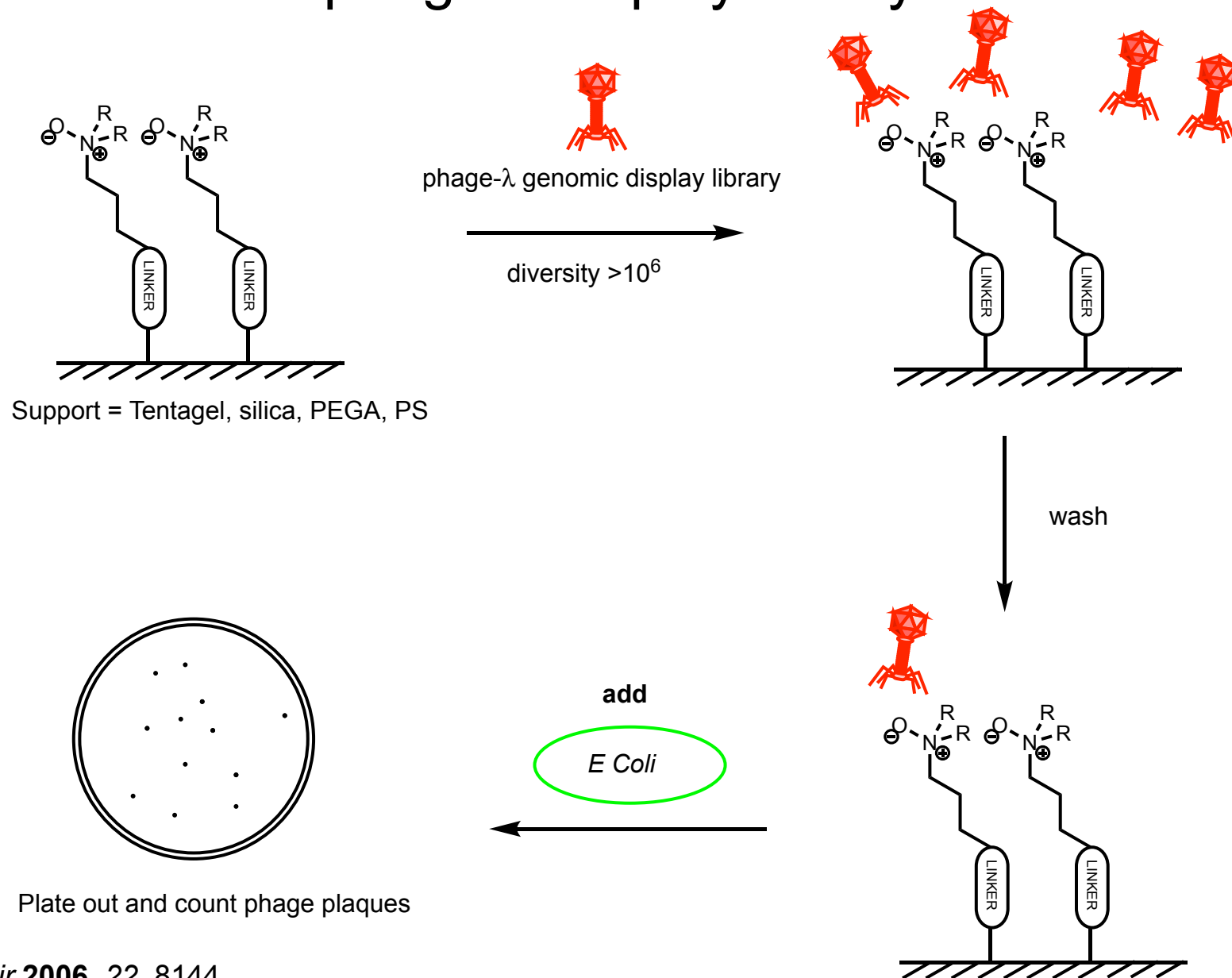
Dilly SJ, Beecham MP, Brown SP, Griffin JM, Clark AJ, Griffin CD, Marshall J, Napier RM, Taylor PC, Marsh A. *Langmuir* **2006**, *22*, 8144.

New substrates prepared: tertiary amine oxides

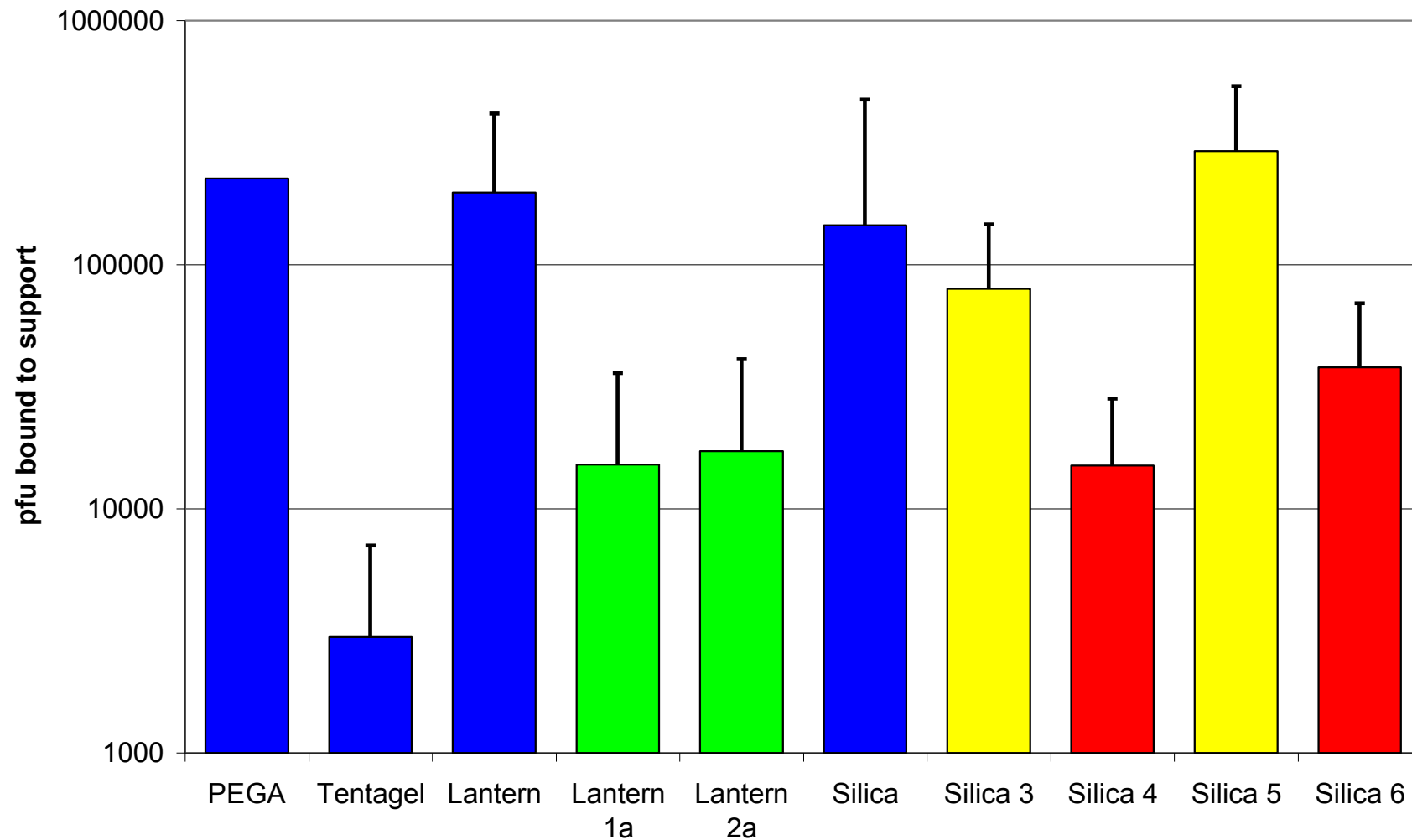


Dilly SJ, Beecham MP, Brown SP, Griffin JM, Clark AJ, Griffin CD, Marshall J, Napier RM, Taylor PC, Marsh A. *Langmuir* **2006**, 22, 8144.

Testing non-specific adhesion using plant genome phage- λ display library

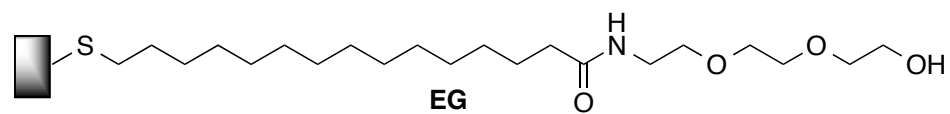
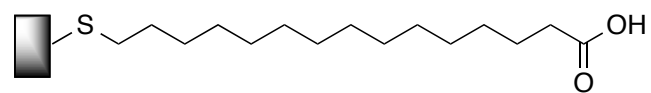
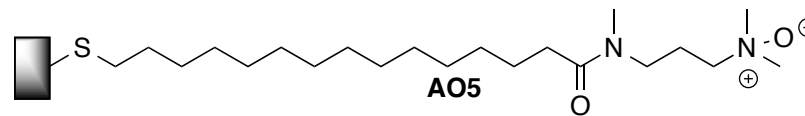
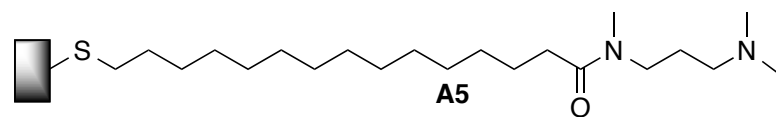
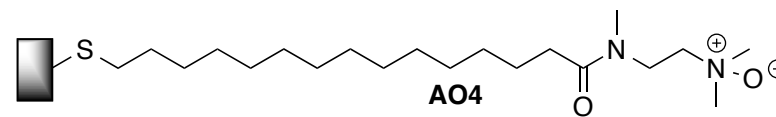
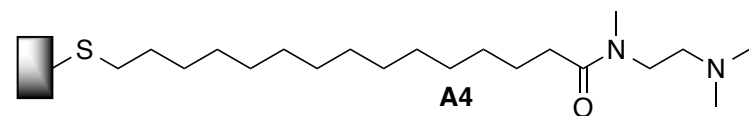
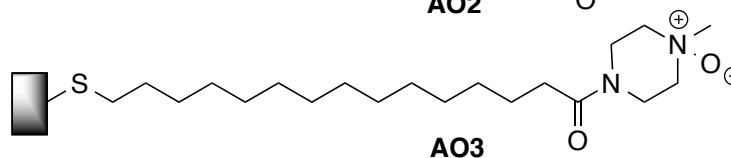
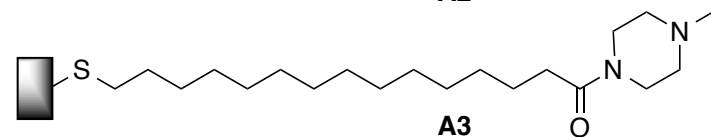
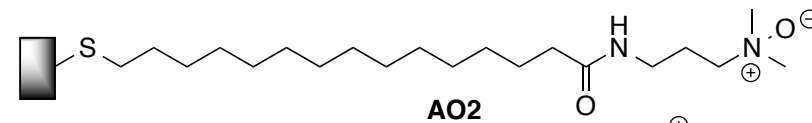
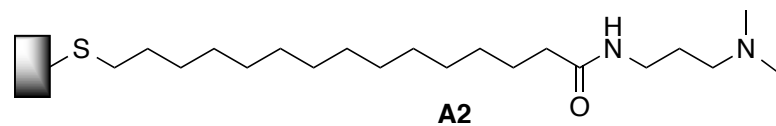
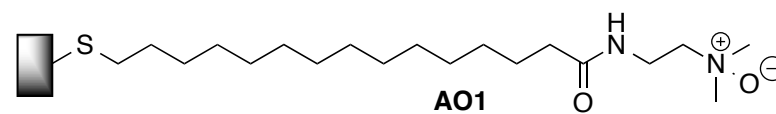
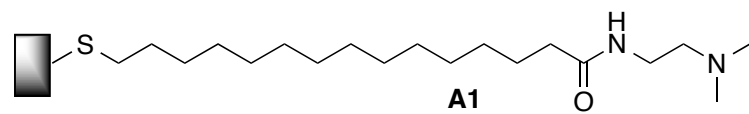


Results of non-specific protein adhesion tests

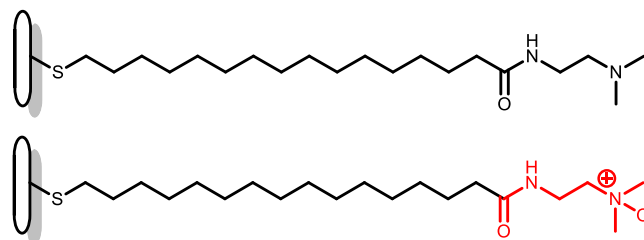


- Low binding levels desirable

Dilly SJ, Beecham MP, Brown SP, Griffin JM, Clark AJ, Griffin CD, Marshall J, Napier RM, Taylor PC, Marsh A. *Langmuir* **2006**, 22, 8144.

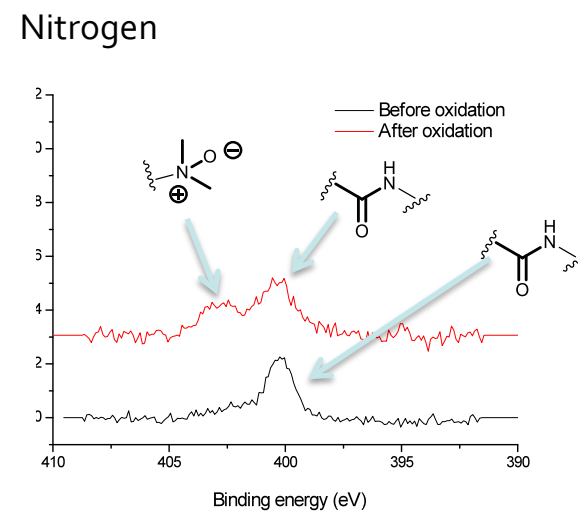
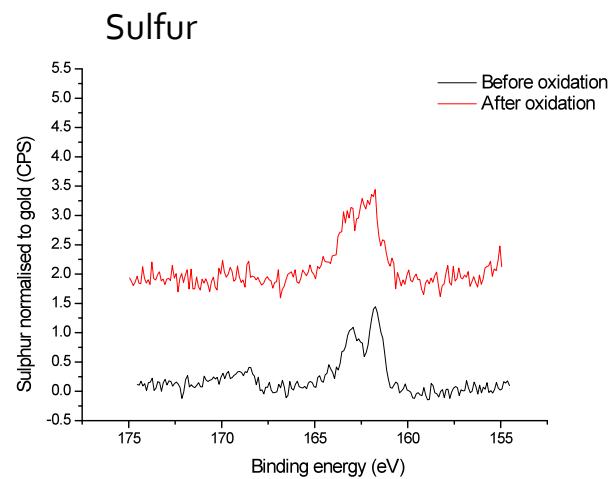
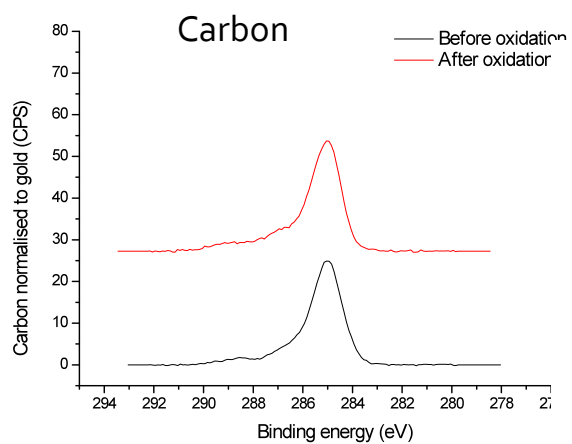


XPS and water contact angle

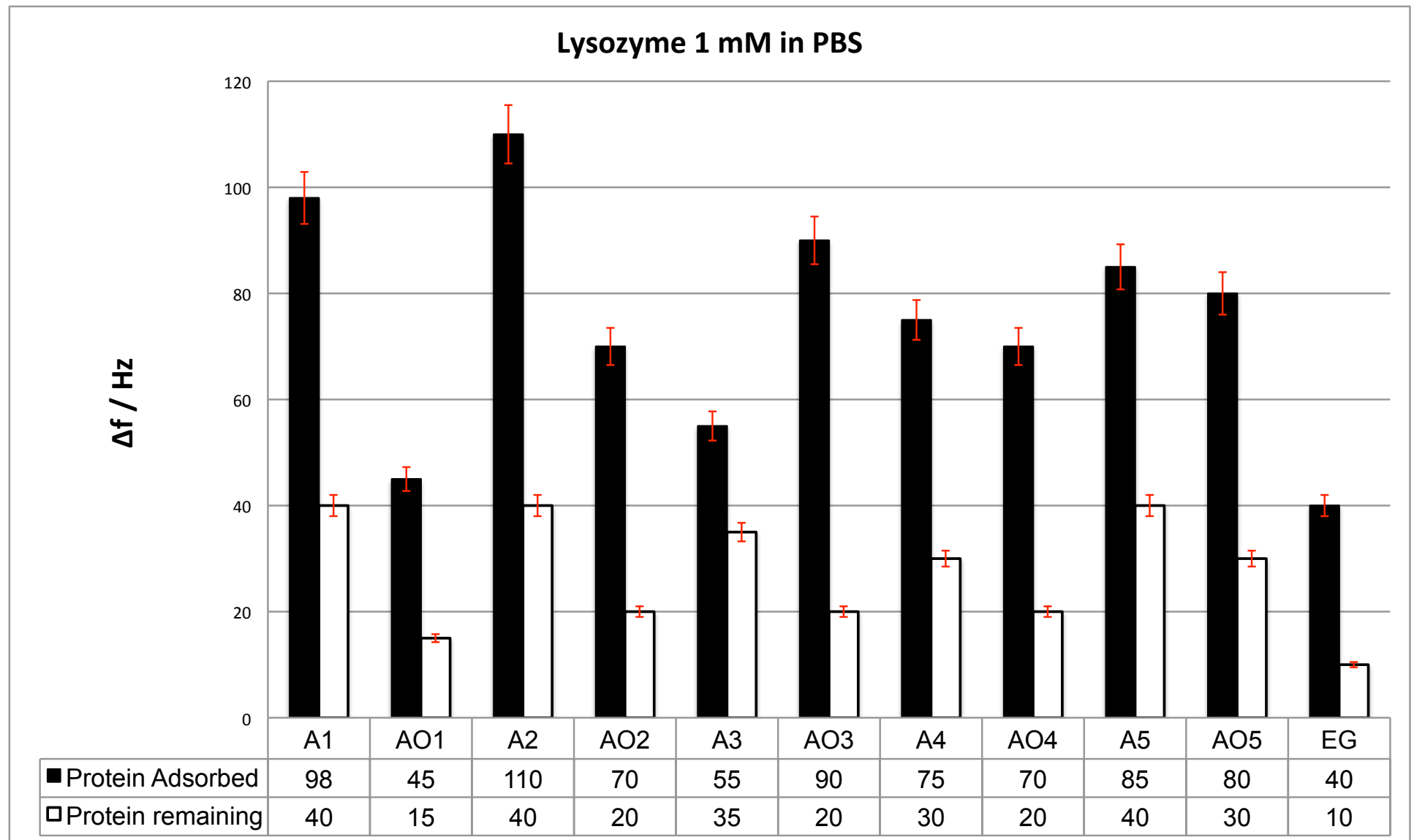


average contact angle/° 34±3

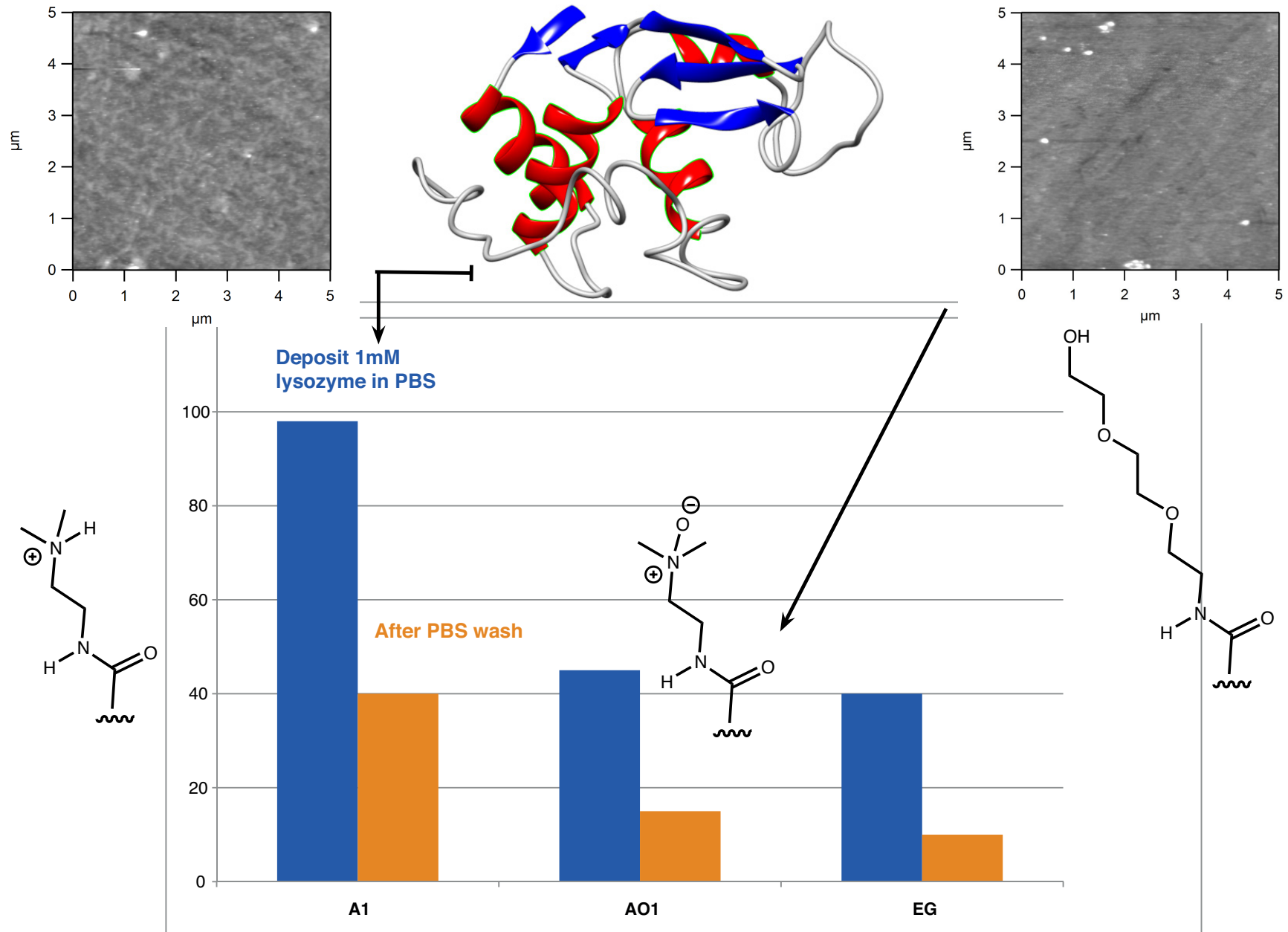
average contact angle/° 25±3



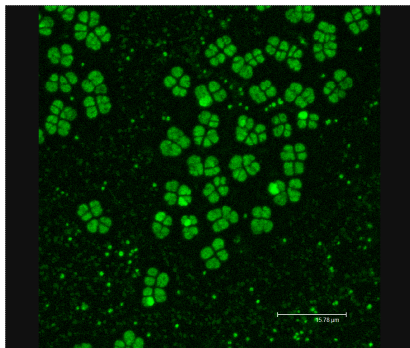
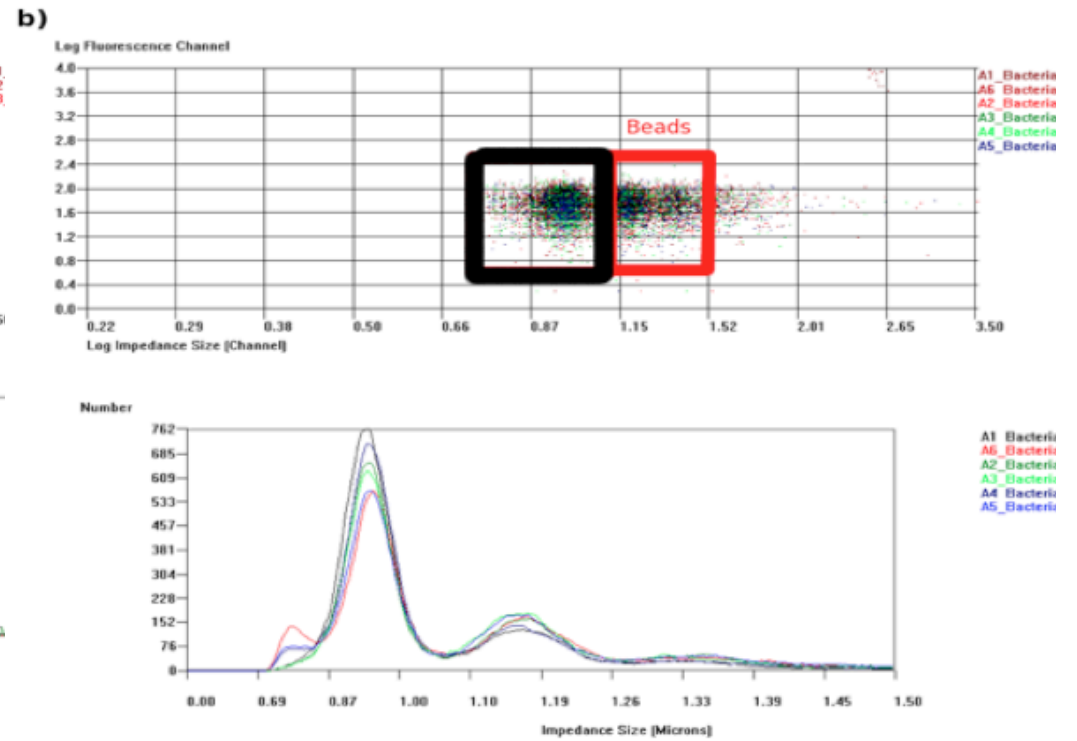
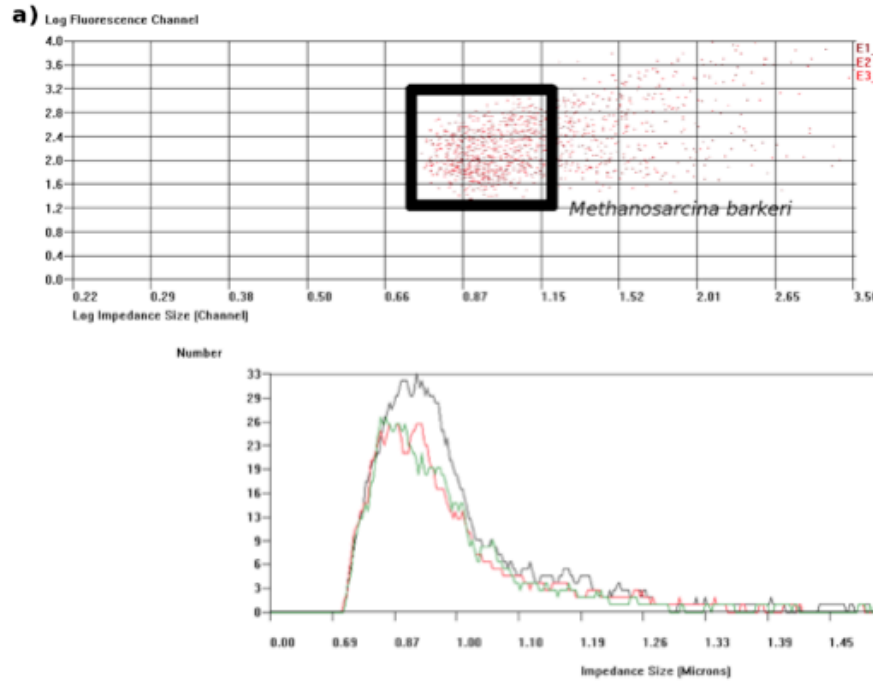
QCM study of lysozyme and fibrinogen adhesion to surfaces



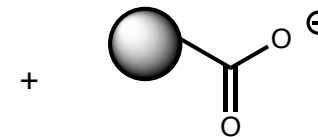
Tertiary amine *N*-oxide derivatised gold surfaces to control protein adhesion



Flow cytometry: *M. barkeri* and 1 μm PS beads

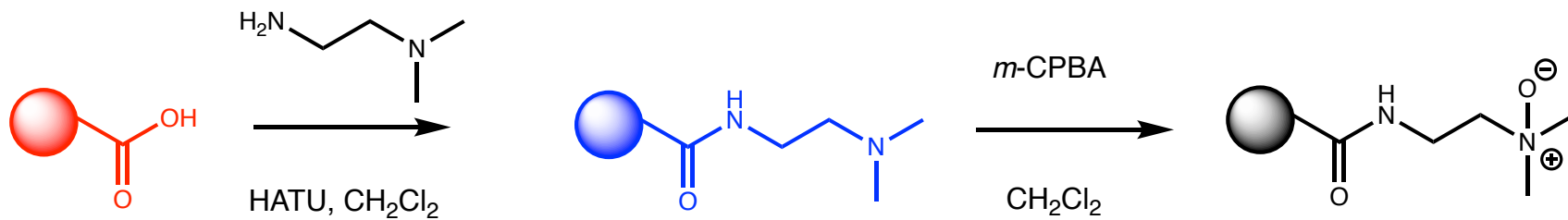
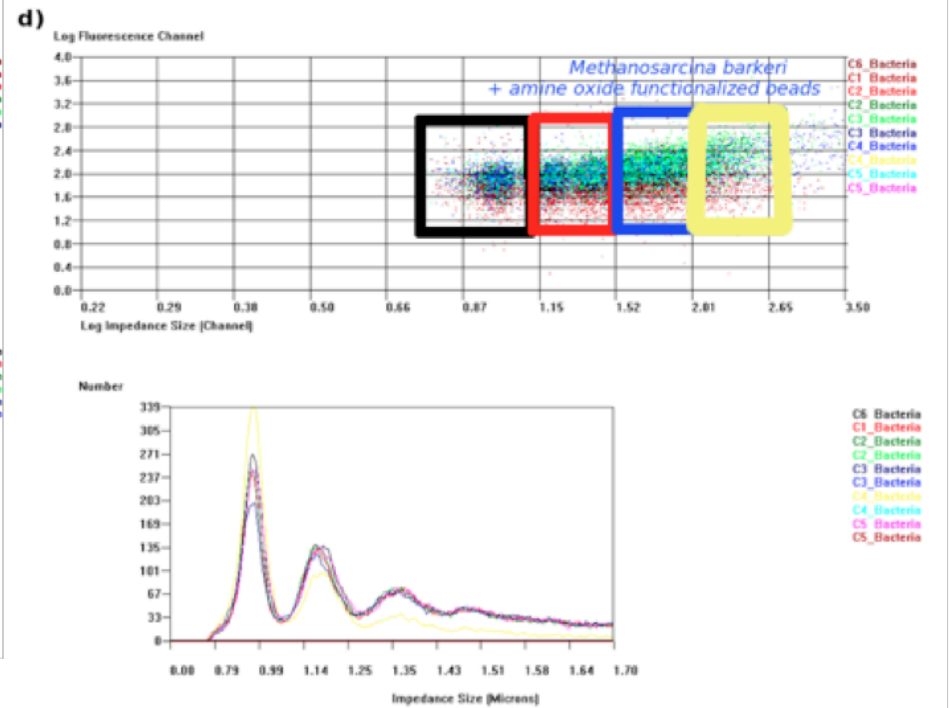
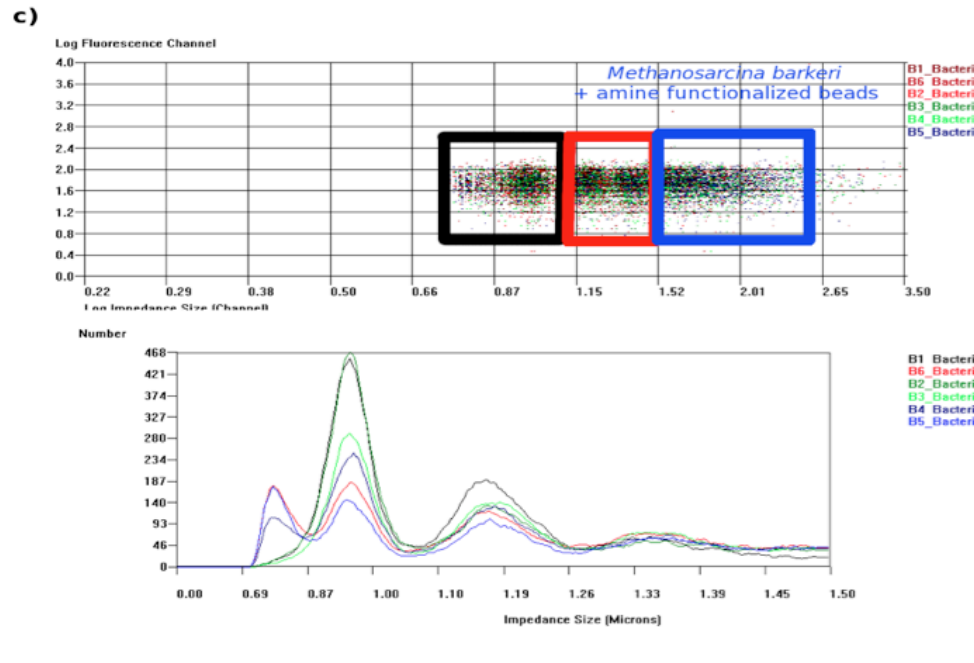


Methanosarcina barkeri

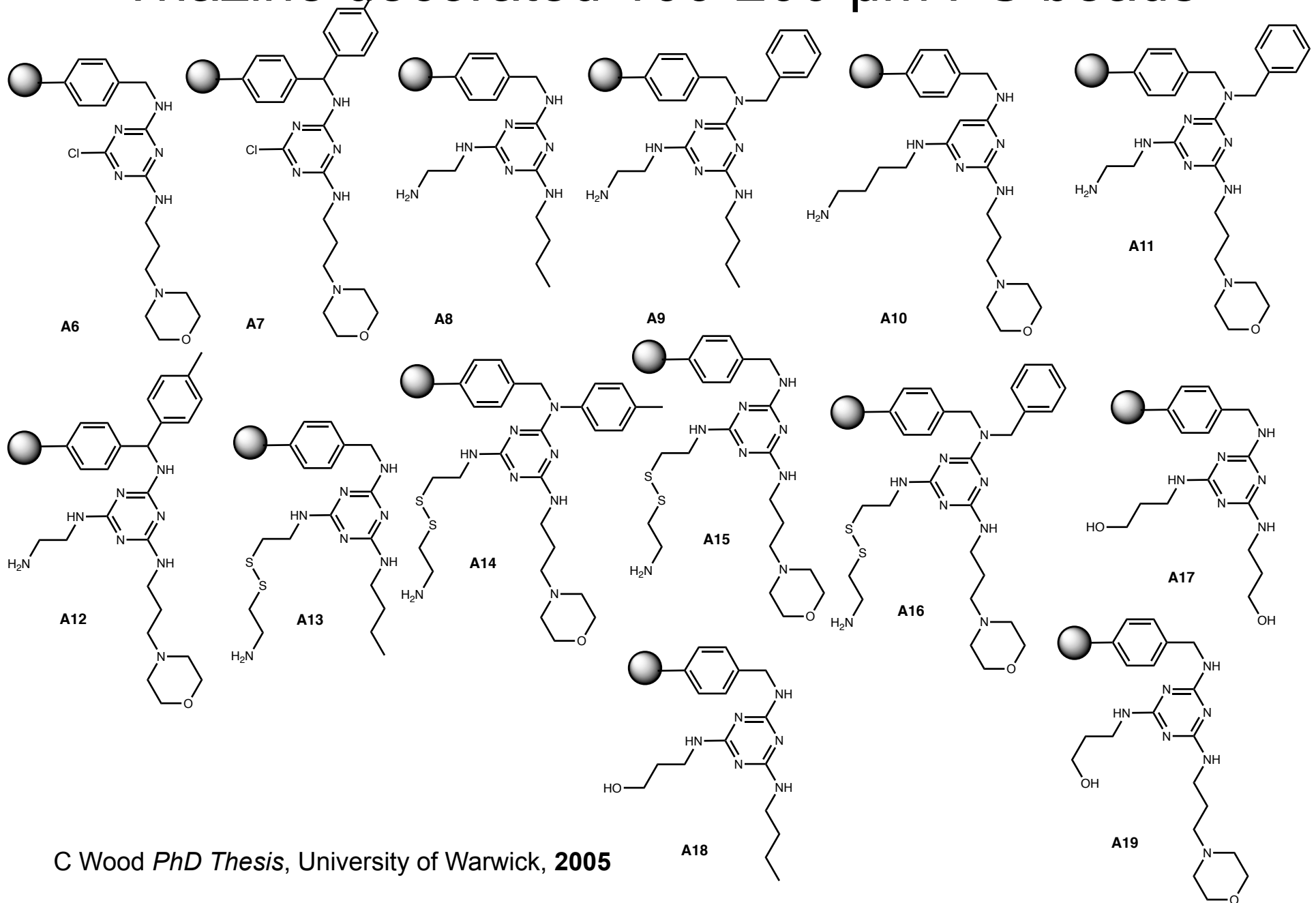


1 μm polystyrene beads

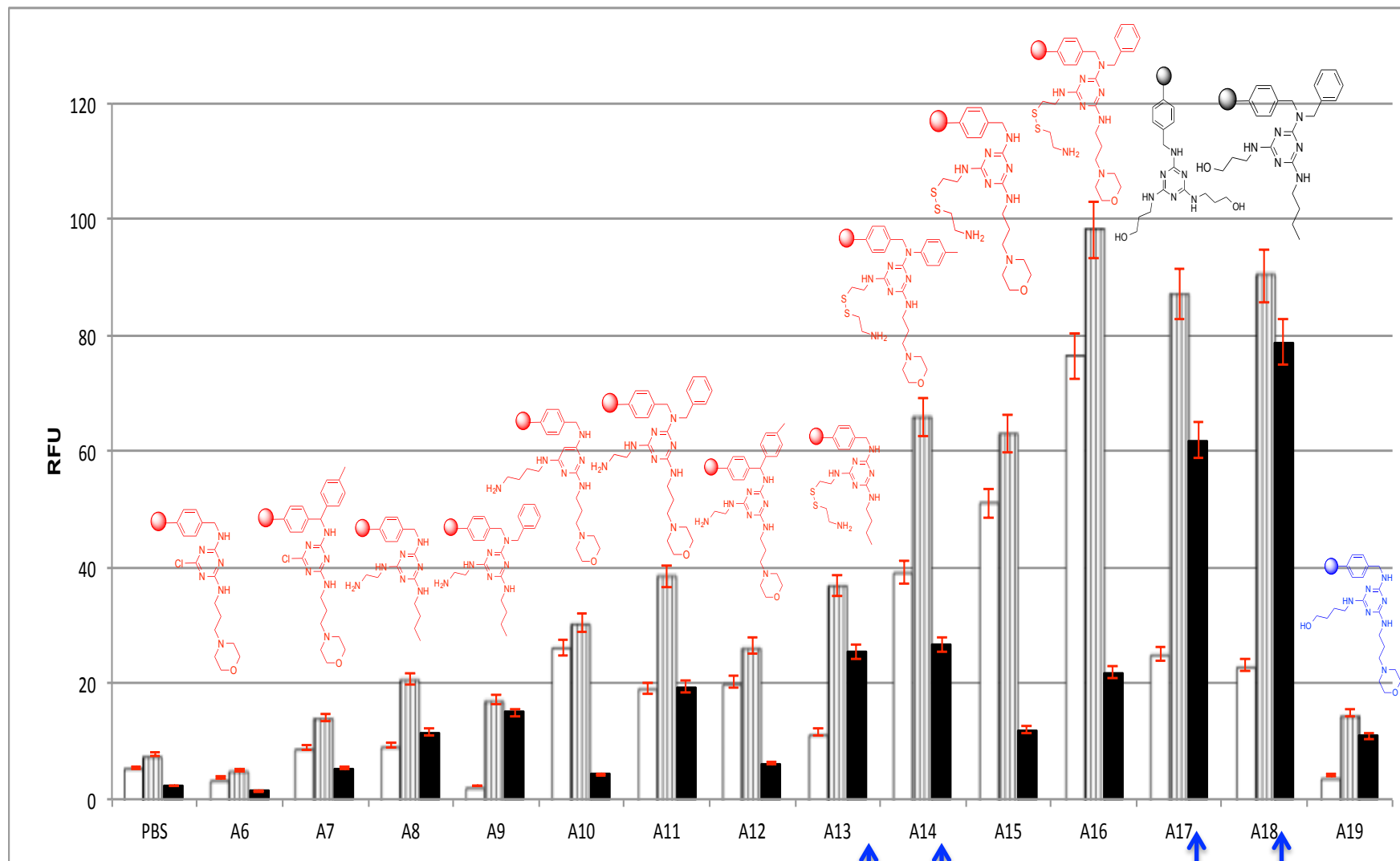
Dimethylamino- and amine *N*-oxides



Triazine decorated 100-200 μm PS beads



Fluorescence plate reader assay



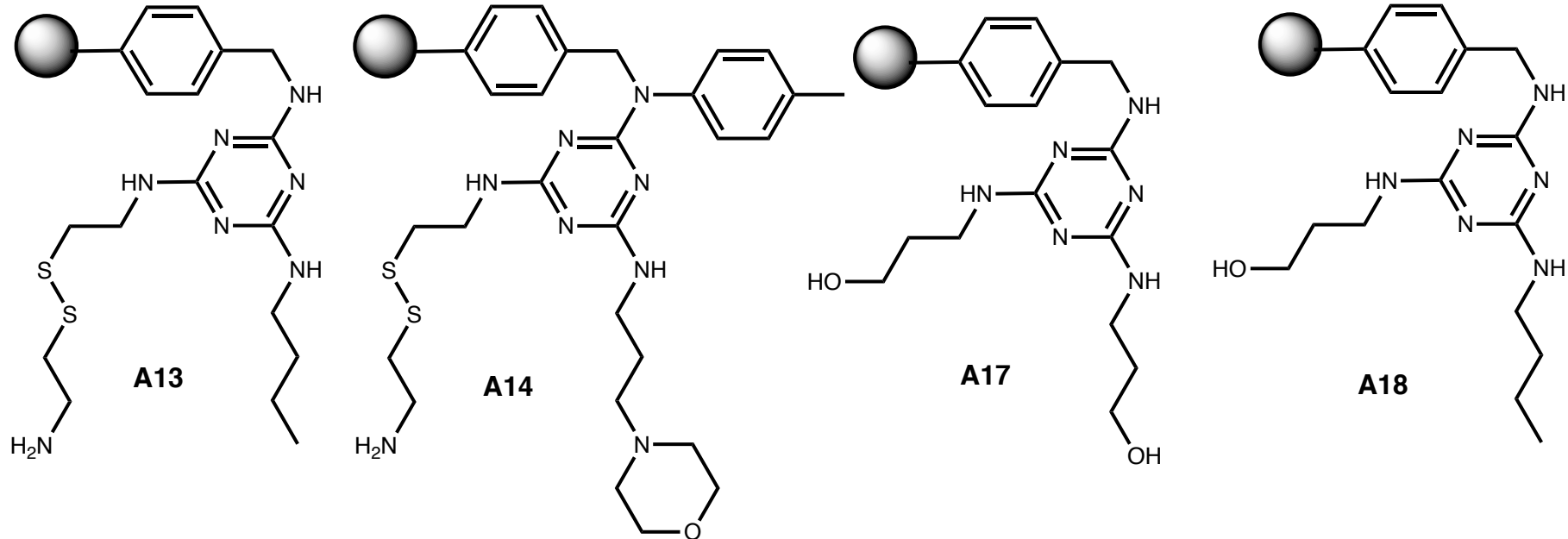
Fluorescence intensity of:

(a) **White** PS beads in PBS buffer after 12h, anaerobic chamber.

(b) **Grey** *M. barkeri* with PS beads in PBS buffer

(c) **Black** *M. barkeri* intrinsic fluorescence by subtraction *i.e.* proportional to concentration

PS beads binding *M. barkeri*



Calc. pKa 9.6 (NH_3^+)

9.6 (NH_3^+)

8 (triazine NH^+)

6.2 (triazine NH^+)

at pH 7.4

7 (morpholine NH^+)

8 (triazine NH^+)

Best interactors neutral hydrophobic with $-\text{OH}$

Methanogen cell envelope structures

Plausible interaction with hydrophobic cell envelope. Possibility of more specific molecular interactions with glycoproteins.

For overview of archaea cell envelope structure see:

Putative cell envelope and adhesion proteins on the outer layer of *Methanobrevibacter ruminantium* Leahy, S C *et al.* PLOS ONE 2010 5: e8926

Methanosarcina acetivorans cell envelope M. A. Arbing *et al.* *Proc. Natl. Acad. Sci USA* **2012**, 109, 11812-11817

Supports used for Anaerobic Digester biofilm stabilization

Fixed support	Advantage	Disadvantage
PVC sheet media	<ul style="list-style-type: none"> - easy install - low cost - no loss of material 	<ul style="list-style-type: none"> - fouling if rag removal inadequate
Fabric web	<ul style="list-style-type: none"> - easy install - no loss of material 	<ul style="list-style-type: none"> - prone to brandling worm blooms
Dispersed support		
Polypropylene cylinders	<ul style="list-style-type: none"> - excellent mixing - high surface area 	<ul style="list-style-type: none"> - media loss - maintenance of aeration system
Polyurethane foam	<ul style="list-style-type: none"> - excellent mixing - high surface area 	<ul style="list-style-type: none"> - media loss - maintenance of aeration system

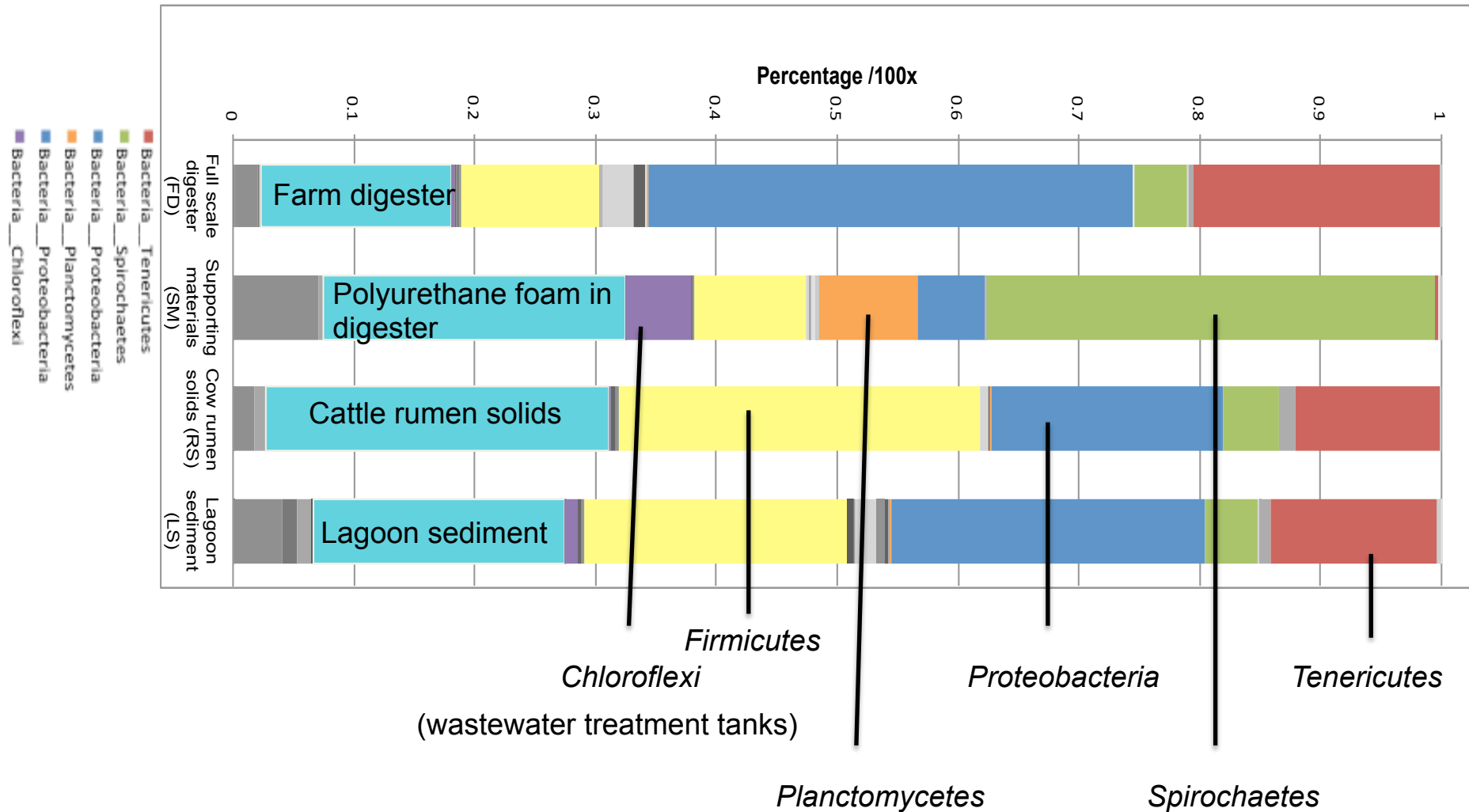
High-throughput 16S rRNA sequencing of surface bound microbes

Extracted from microbes:

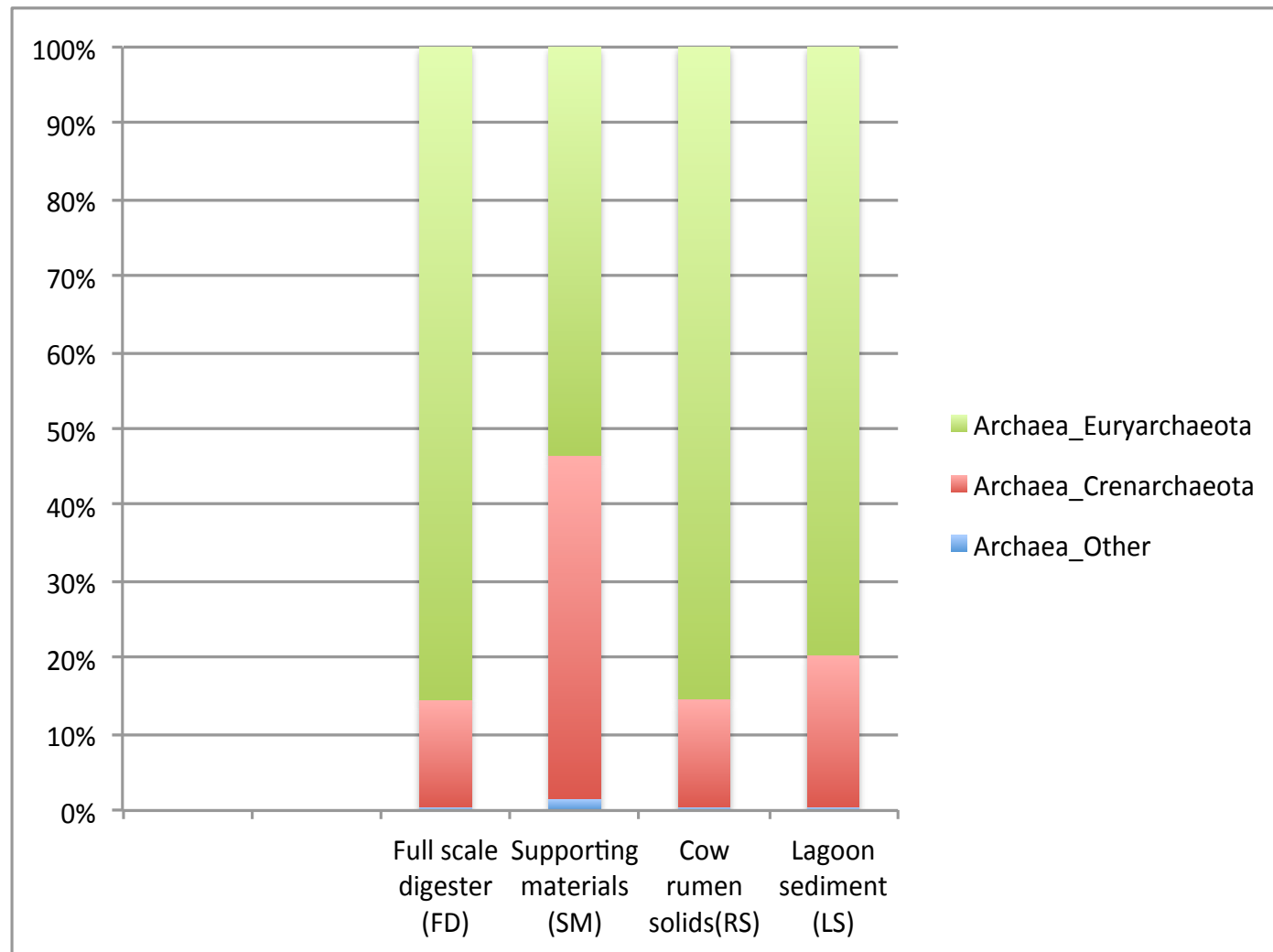
- In farm scale digester liquid
- On polyurethane foam surfaces suspended in farm scale digester
- Cattle rumen solids
- Cattle slurry lagoon

Sequences searched using QIME pipeline against greengenes 16S RNA database to identify *bacteria* and *archaea*

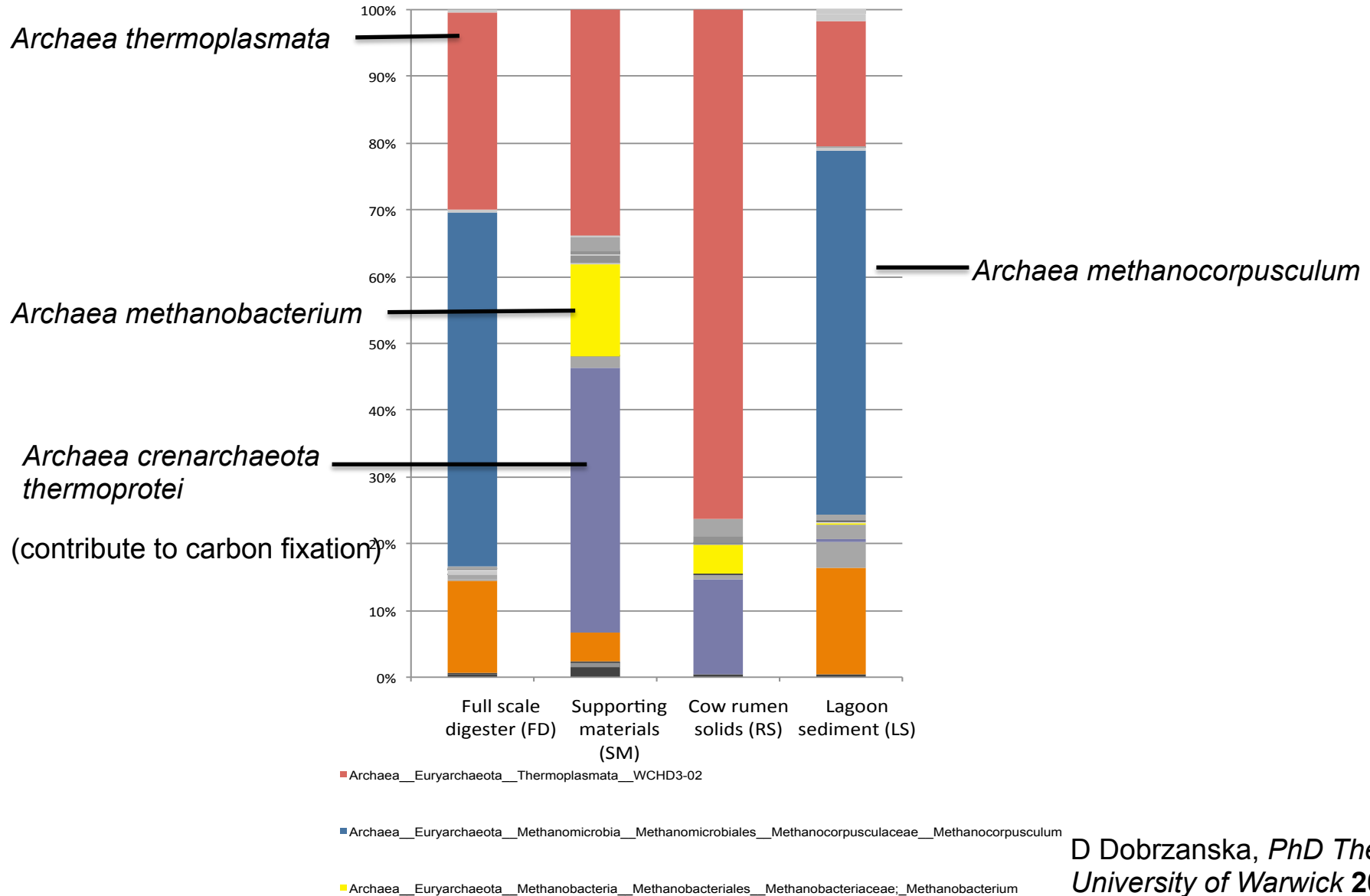
Bacteria from cattle slurry at phylum level using 16S rRNA sequencing



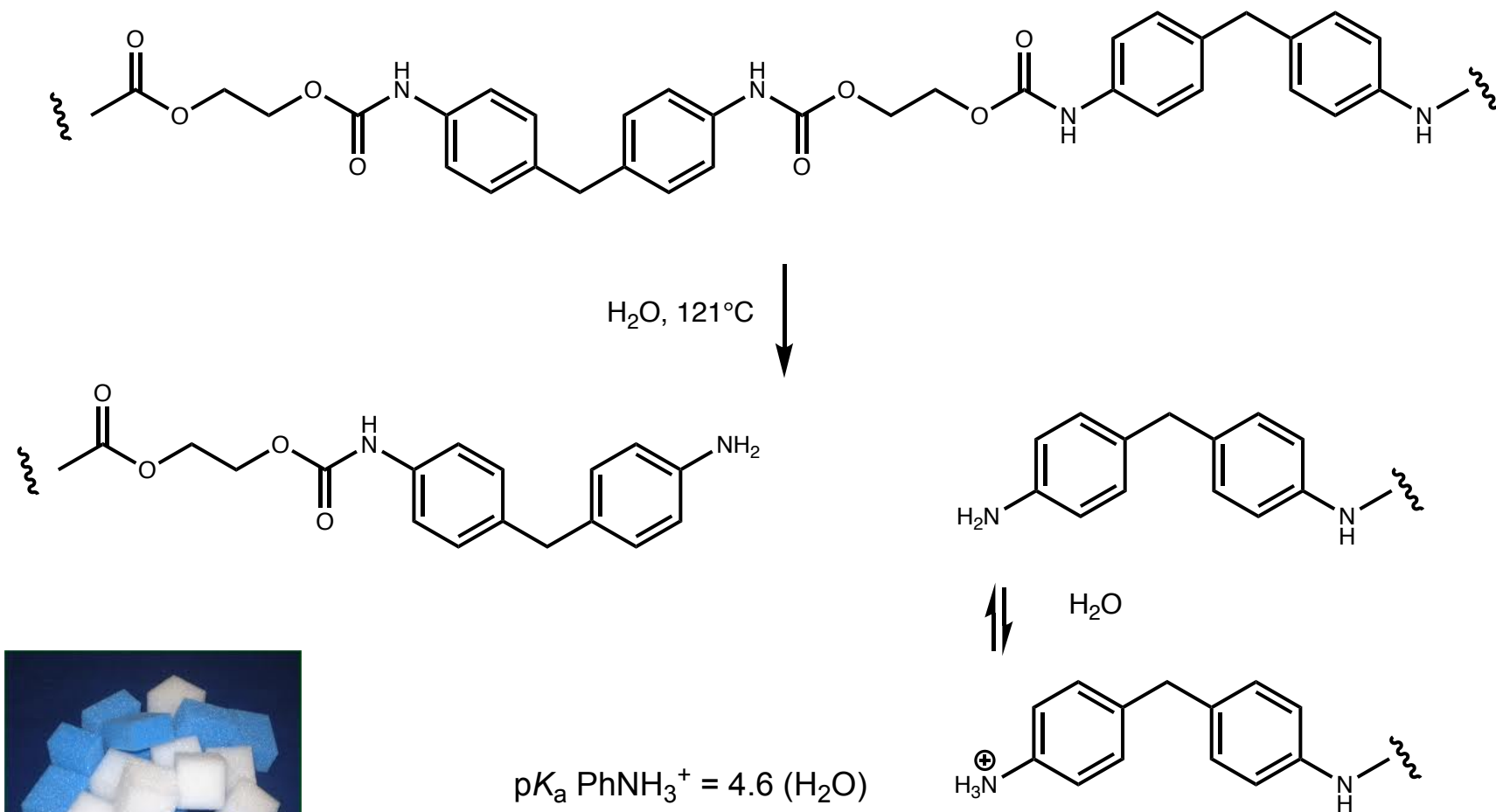
Archaea at phylum level from 16S rRNA sequencing



Archaea at genus level from 16S rRNA sequencing

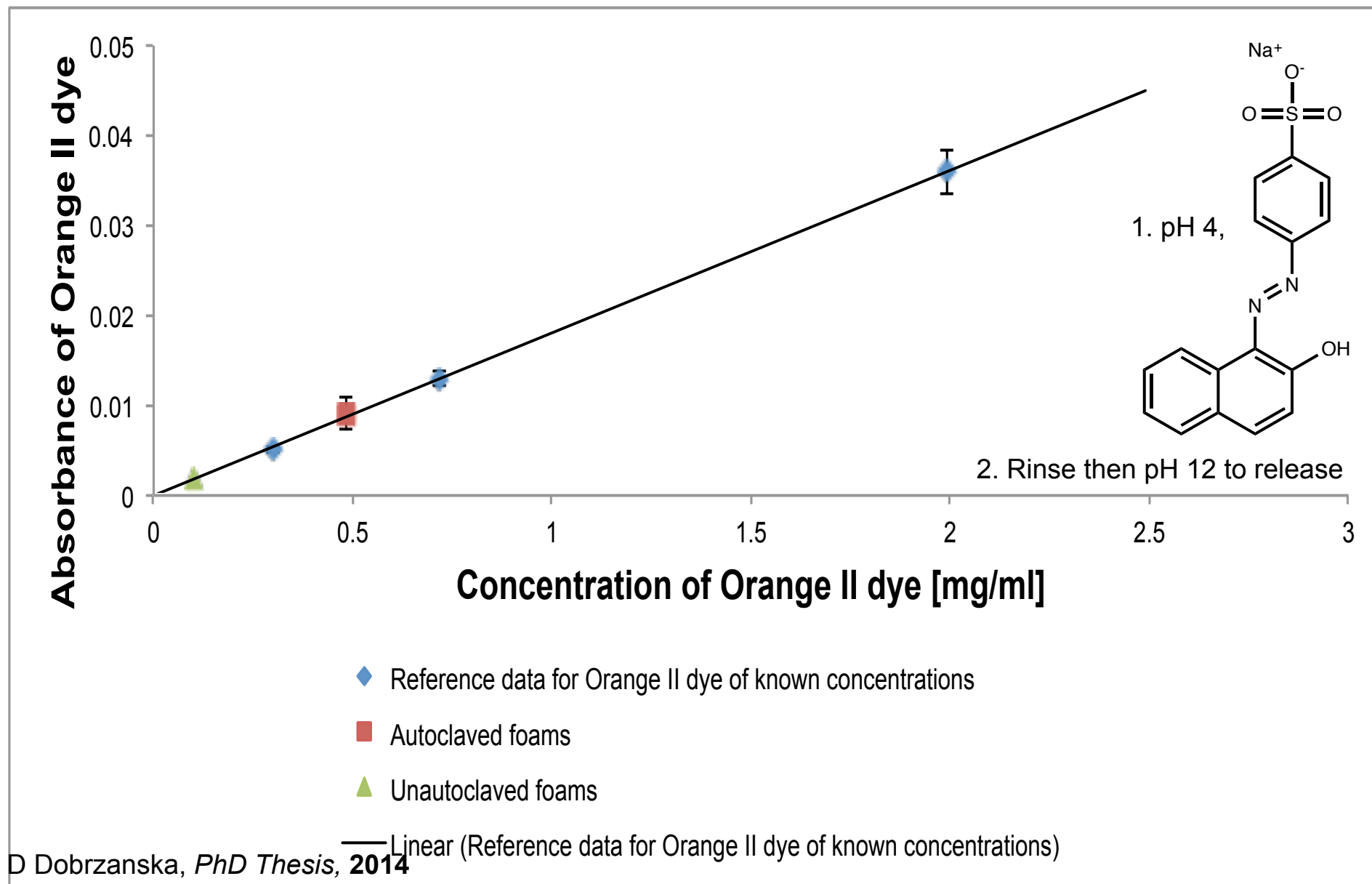


Autoclaved polyurethane surface chemistry



.... its why medical devices containing PU are not sterilised with heat!

Measuring amines exposed at surface



Summary

- Studied a wide range of substrates and derivatives at interfaces and demonstrated utility of tertiary amine *N*-oxide as a protein resistant functional group
- Found uncharged surfaces preferentially attract a prototype methanogen, *M. barkeri*, possibly interacting with glycoproteins set in a hydrophobic cell envelope
- PU foam attracts a unique set of microorganisms, including methanogens from a large scale anaerobic digester

Acknowledgements

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- John Griffin, Stephen Brown (Dept of Physics)
- Stephen Evans, Benjamin Johnson (University of Leeds)

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