

Mapping Conformational Changes in Proteins through Crystallographic Refinement and Computational Approaches

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Indo-UK Networking Meeting

17-19 May 2011

Laboratory Activities

Crystallography (Biochemical and Biophysical analysis) of *M. tuberculosis* proteins:

Hsp60, Hsp65, Hsp10

AhpC, Trx's, TrxR, Glutaredoxin

Chorismate mutase, toxin-antitoxin, CRP

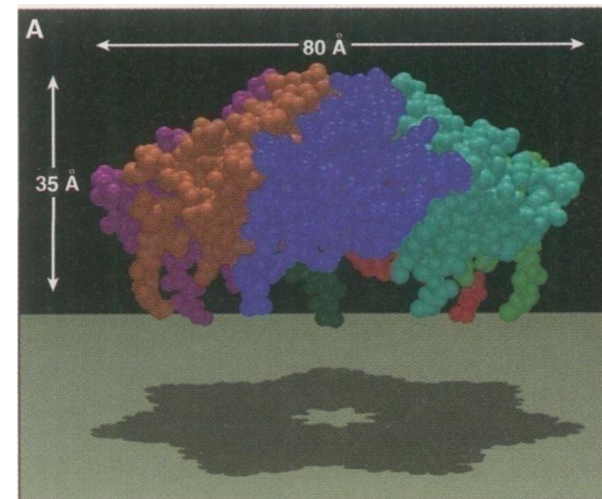
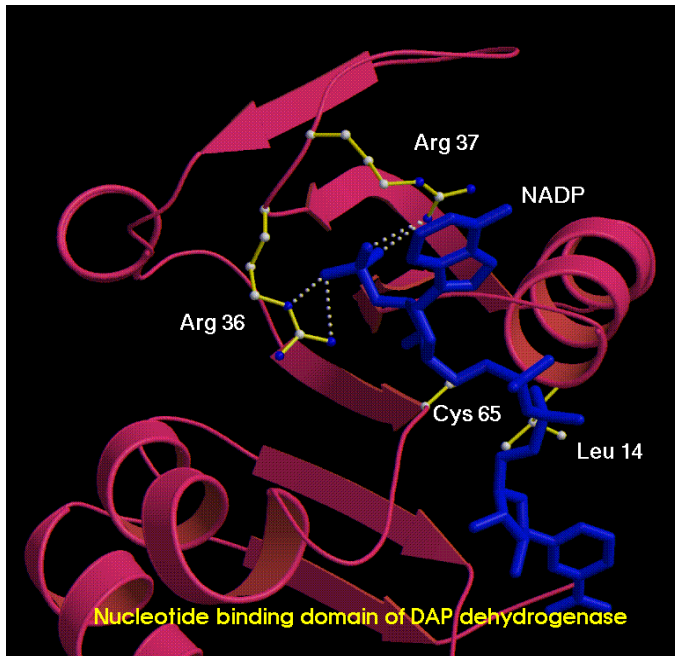
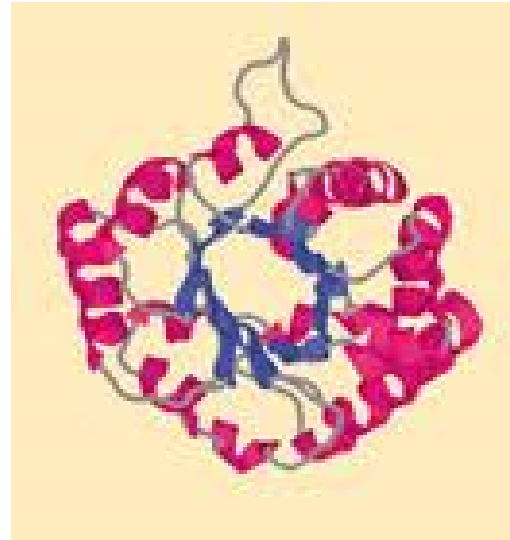
Networks

Genome-wide protein interaction networks

Dynamics of networks

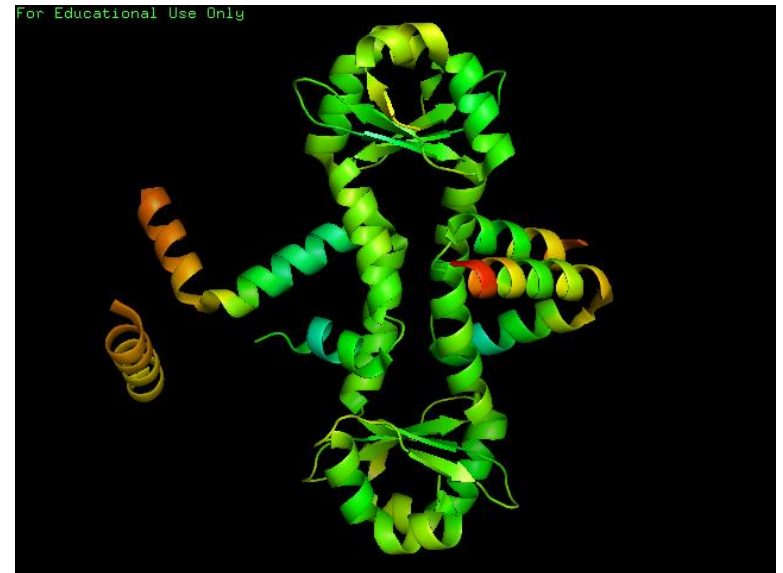
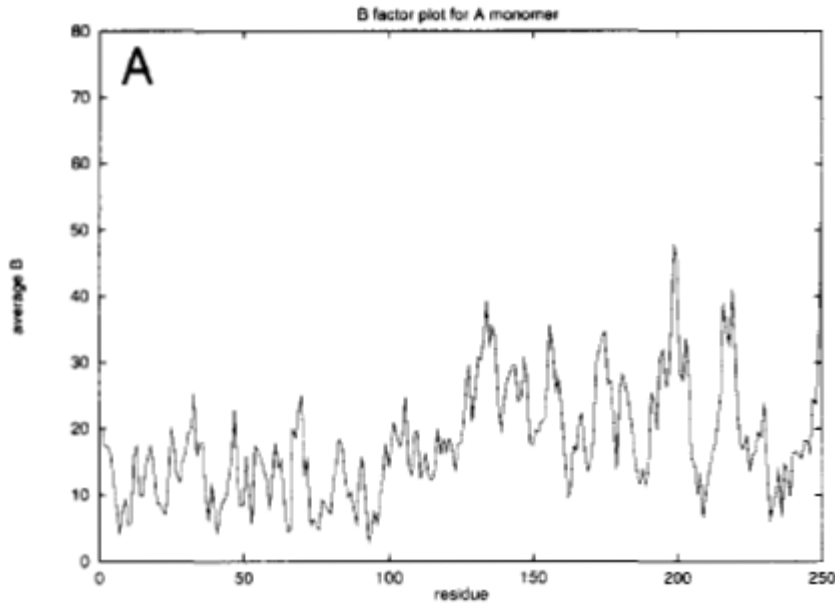
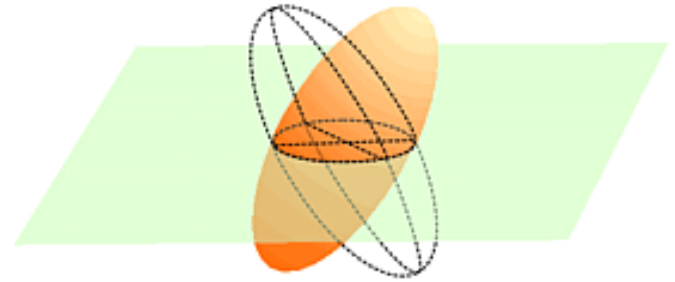
Prediction of genome-wide lethalties, synthetic lethalties

Protein Structures: Beyond pretty pictures



Thermal fluctuations in proteins: B-factor

$$B = 8 \pi^2 \langle u^2 \rangle$$



A typical B-factor represents isotropic movement of an atom around its mean position

Need to account for dynamic behaviour of proteins

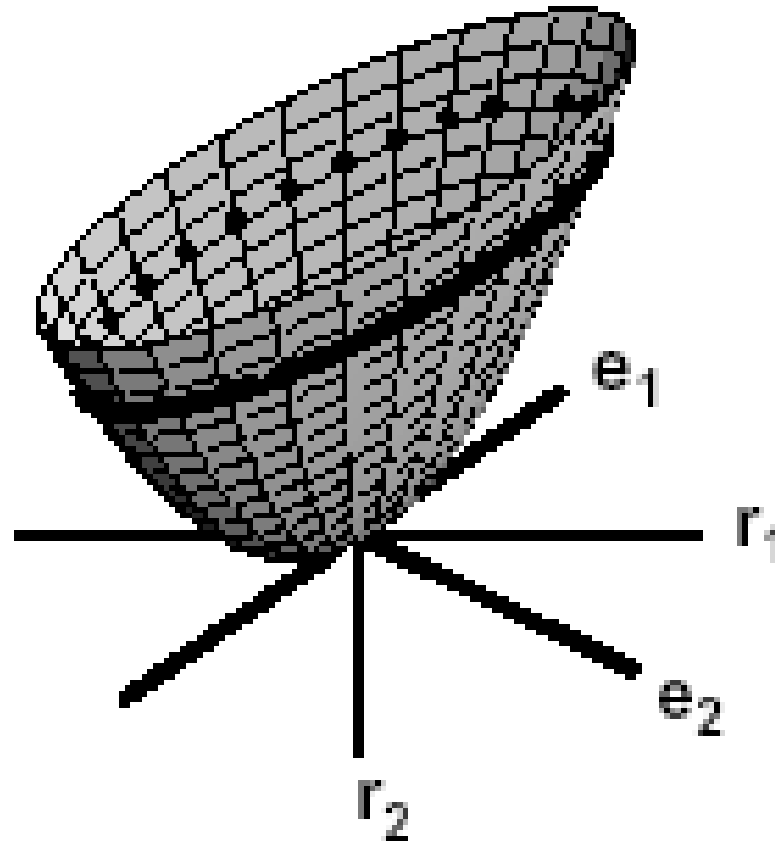
$$\begin{aligned}
 U(\chi) = & \sum_i (\chi_i - \chi_i^0) \left. \frac{\partial U}{\partial \chi_i} \right|_{\chi = \chi_0} + \\
 & \frac{1}{2!} \sum_{ij} \left. \frac{\partial^2 U}{\partial \chi_i \partial \chi_j} \right|_{\chi = \chi_0} (\chi_i - \chi_i^0)(\chi_j - \chi_j^0) + \\
 & \frac{1}{3!} \sum_{ijk} \left. \frac{\partial^3 U}{\partial \chi_i \partial \chi_j \partial \chi_k} \right|_{\chi = \chi_0} (\chi_i - \chi_i^0)(\chi_j - \chi_j^0)(\chi_k - \chi_k^0) + \dots
 \end{aligned}$$



Harmonic approximation

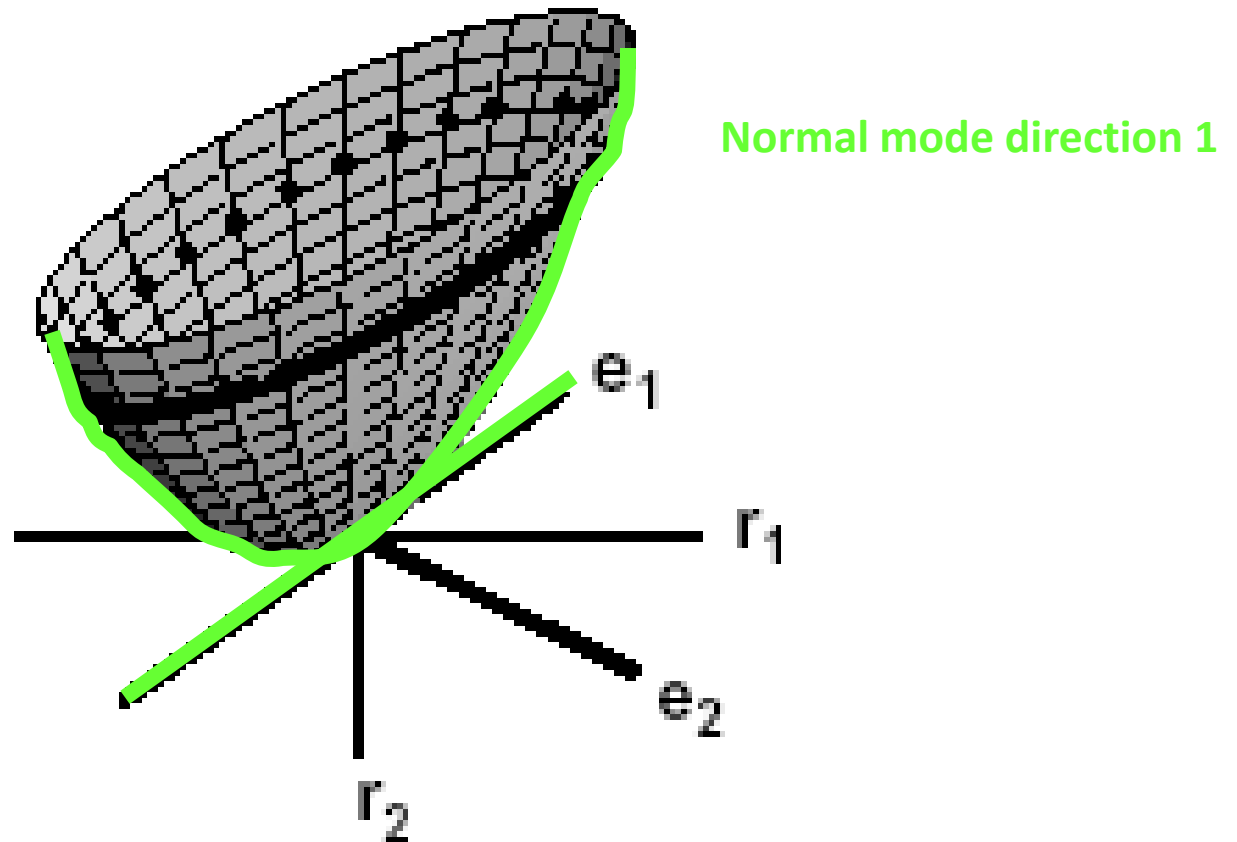
$$U(\chi) \cong \frac{1}{2} \sum_{ij} \left. \frac{\partial^2 U}{\partial \chi_i \partial \chi_j} \right|_{\chi = \chi_0} (\chi_i - \chi_i^0)(\chi_j - \chi_j^0)$$

NMA



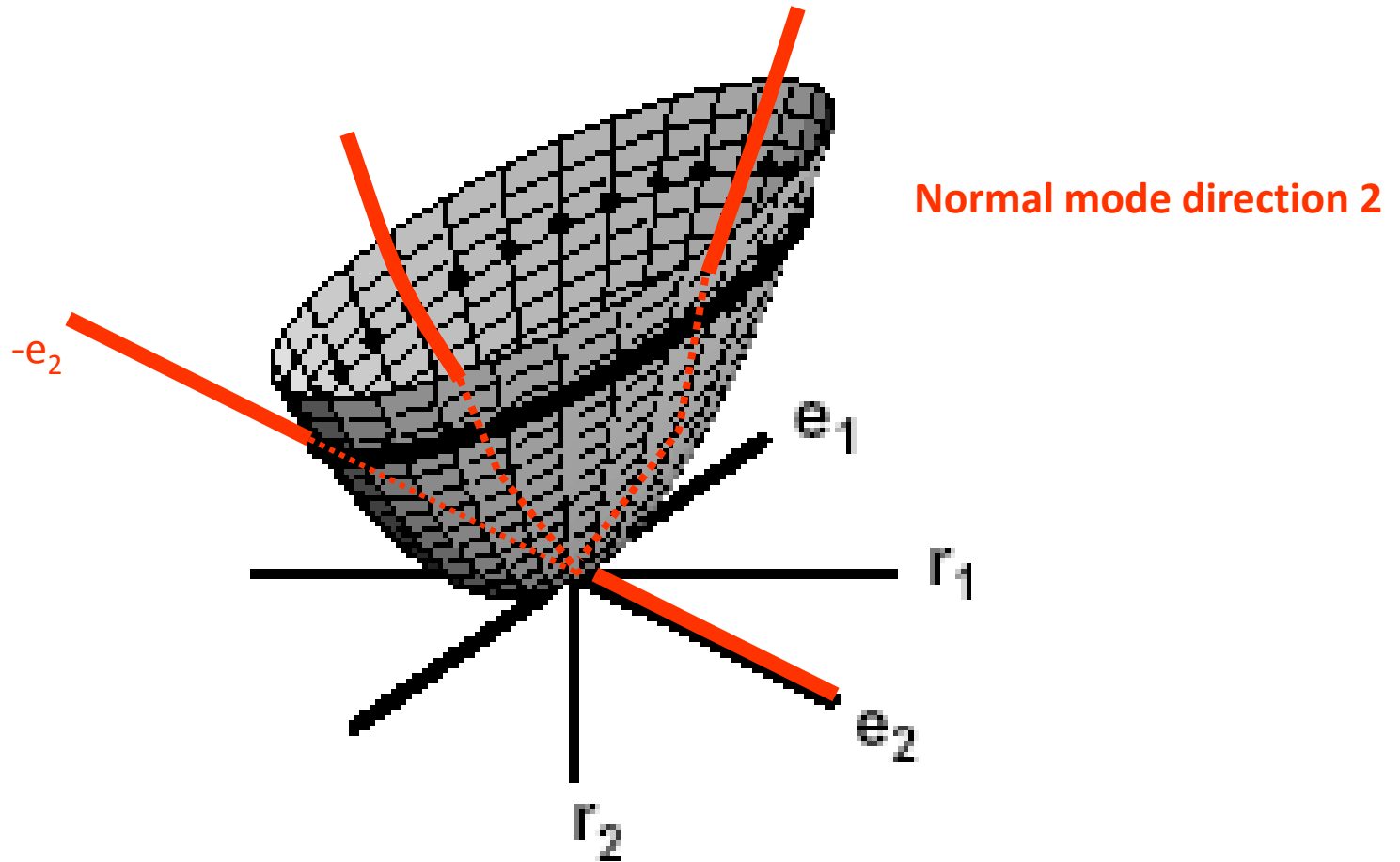
$$U(r) = 0.5 (r - R_{\min})' \cdot K(R_{\min}) \cdot (r - R_{\min})$$

NMA



$$U(r) = 0.5 (r - R_{\min})' \cdot K(R_{\min}) \cdot (r - R_{\min})$$

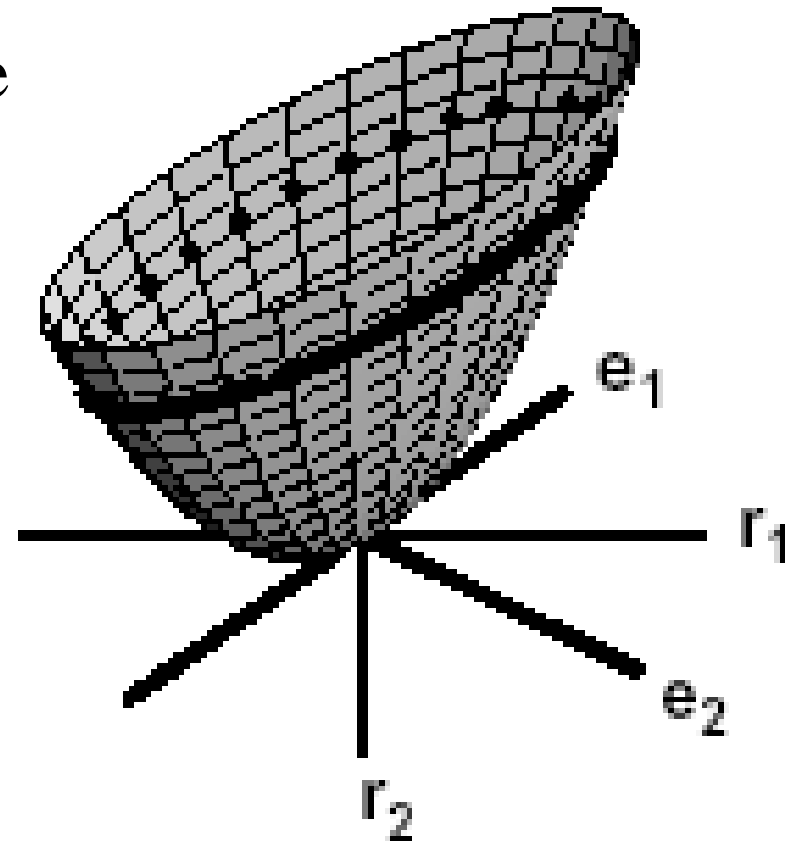
NMA



$$U(r) = 0.5 (r - R_{\min})' \cdot K(R_{\min}) \cdot (r - R_{\min})$$

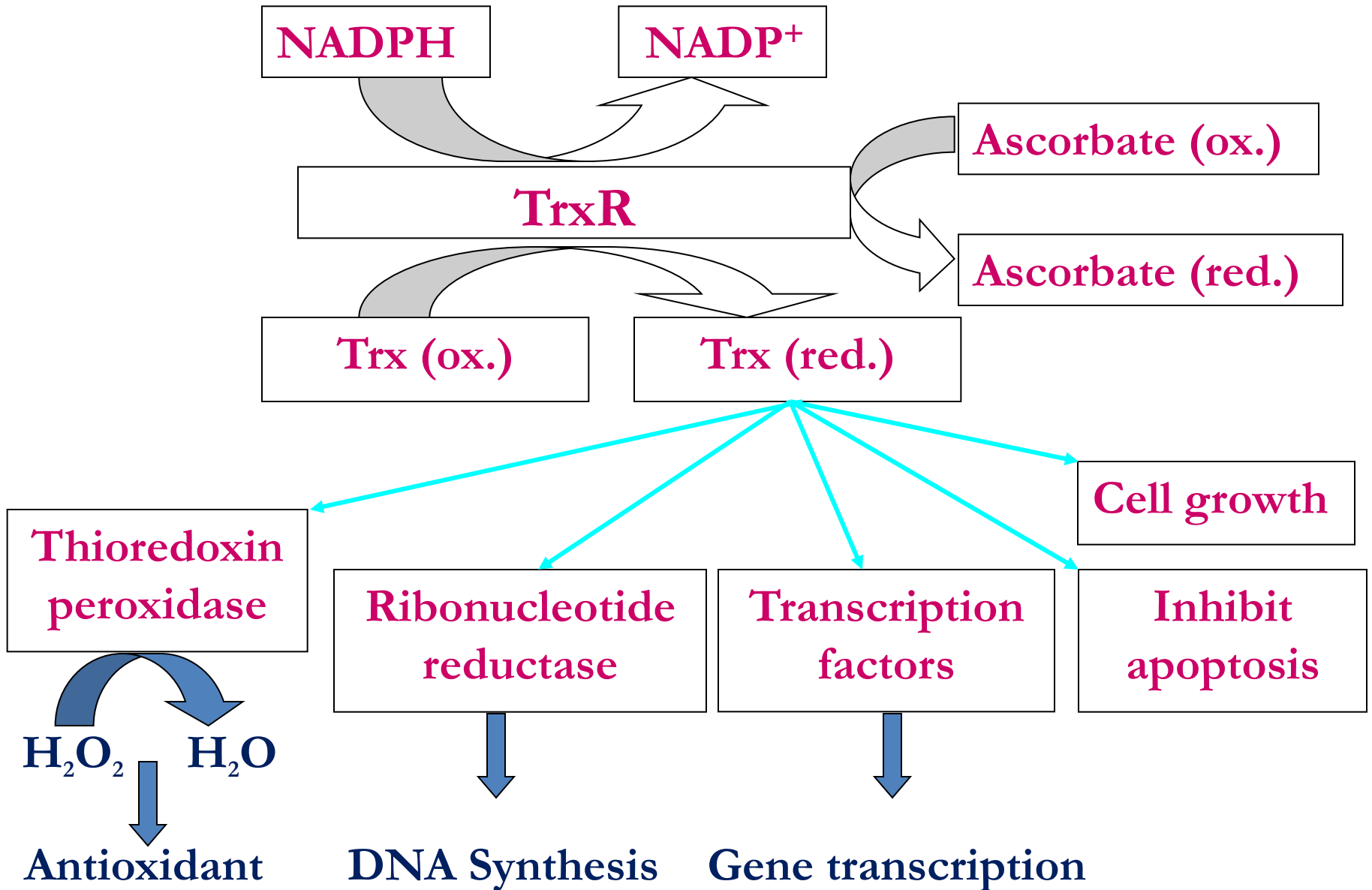
Properties of NMA

- The eigenvalues describe the energetic cost of displacing the system by one length unit along the eigenvectors.
- For a given amount of energy, the molecule can move more along the low frequency normal modes
- The first six eigenvalues are 0, corresponding to rigid body movements of the protein

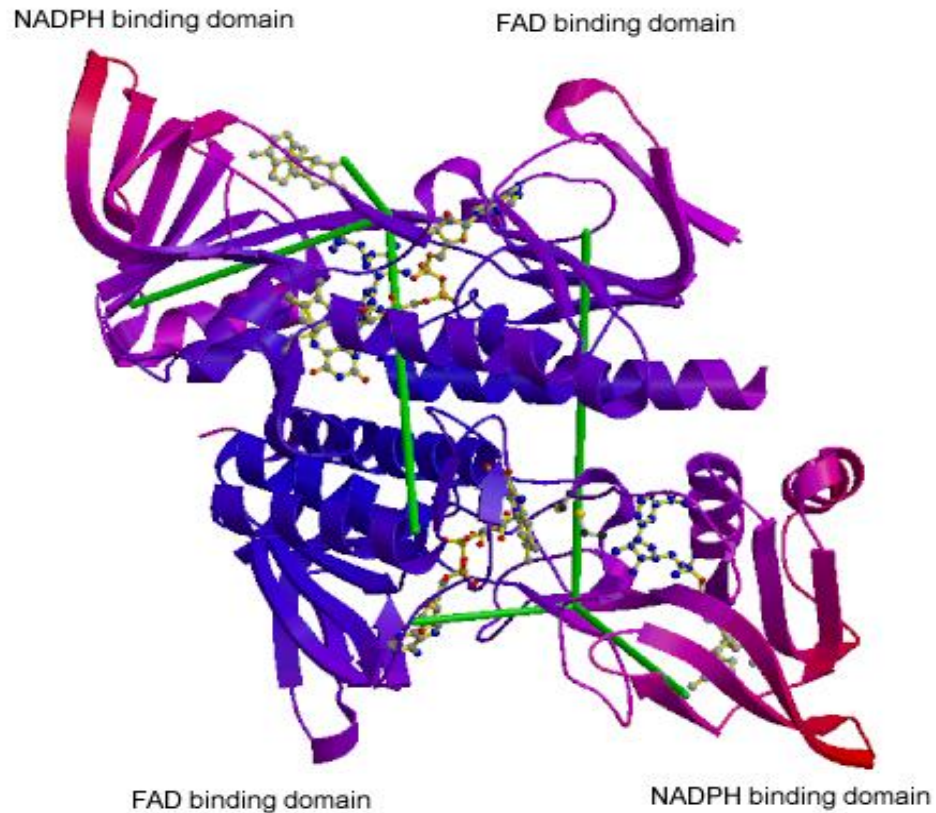


***M. tuberculosis* thioredoxin reductase**

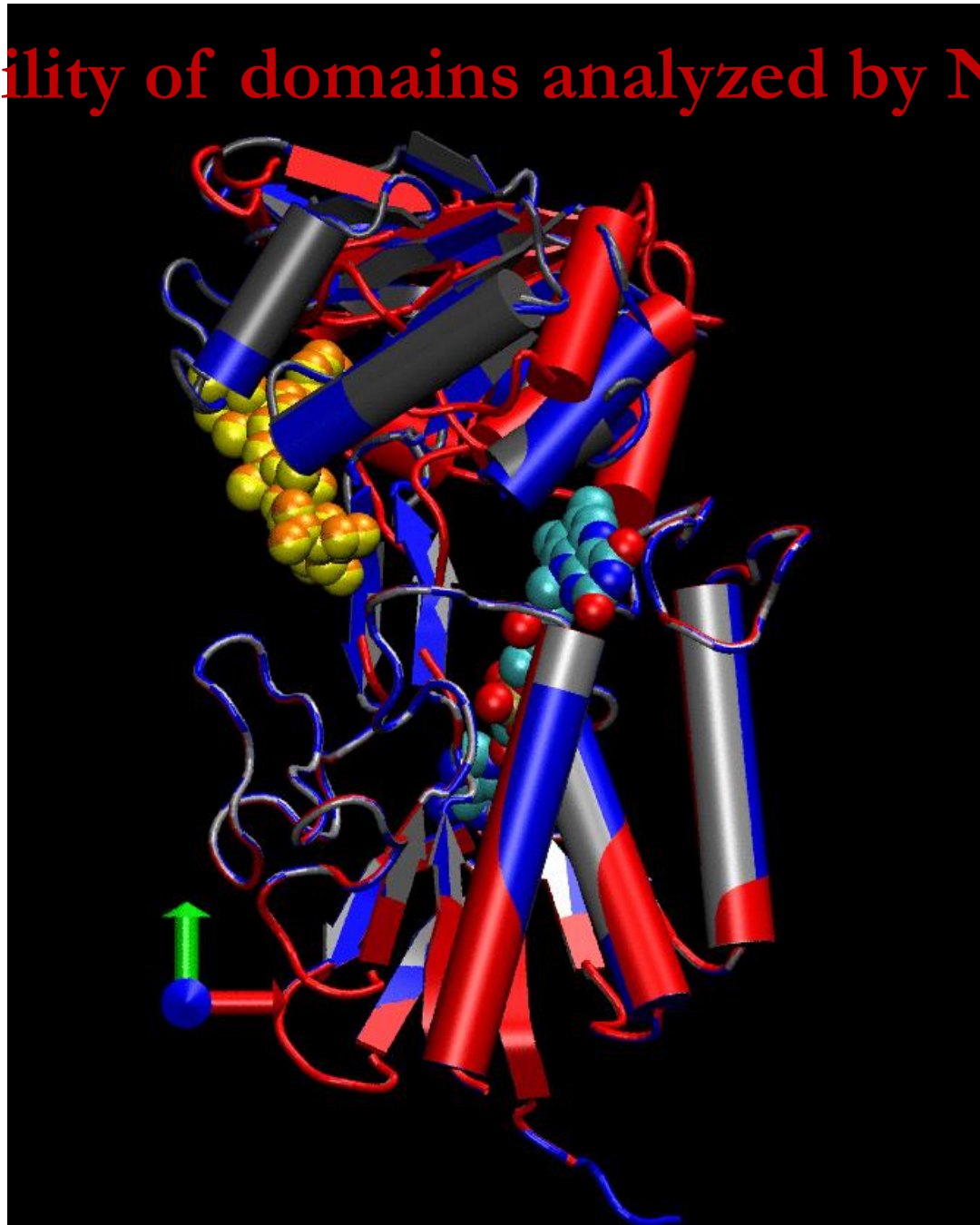
Functions of the Thioredoxin System



Domain Flexibility of *Mtb* TrxR

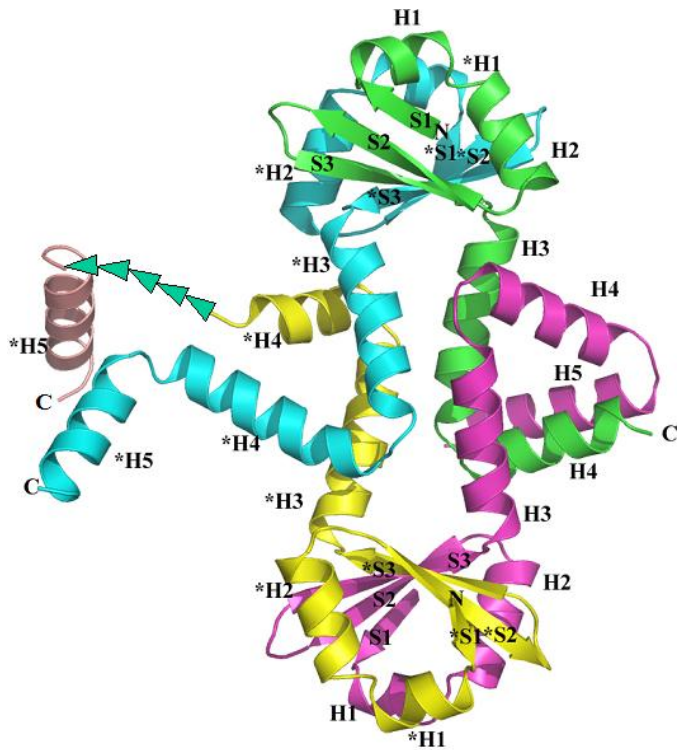


Flexibility of domains analyzed by NMA

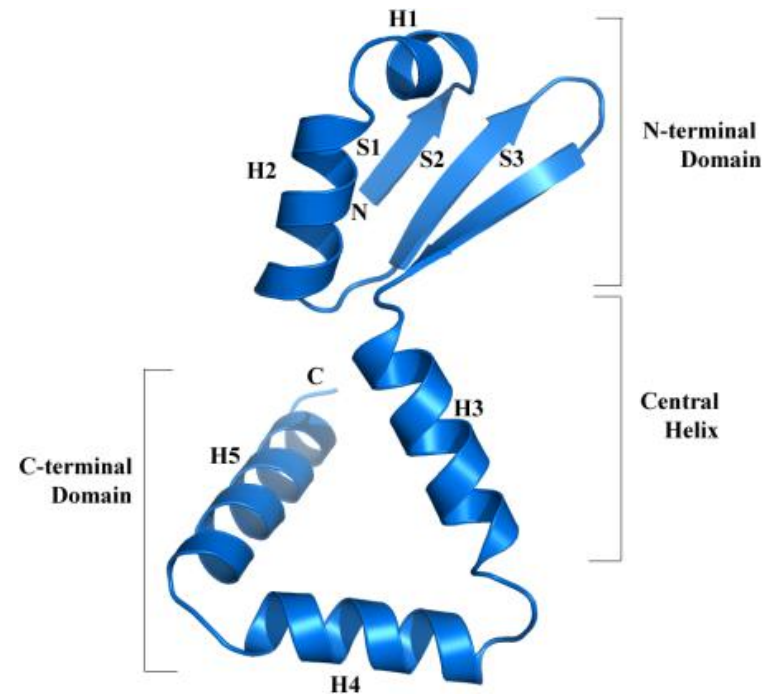


***M. tuberculosis* YefM anti-toxin**

Overall structure

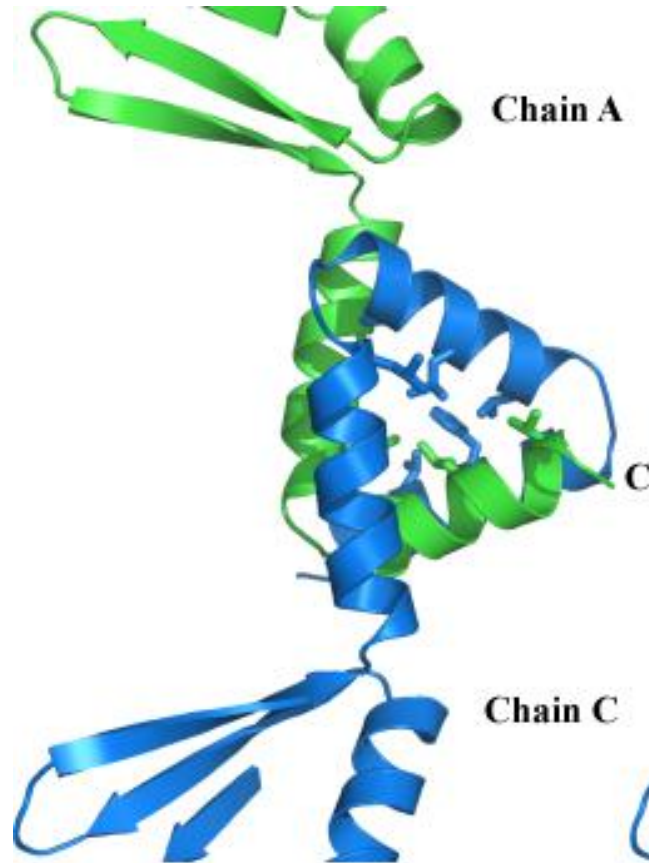


Tetramer



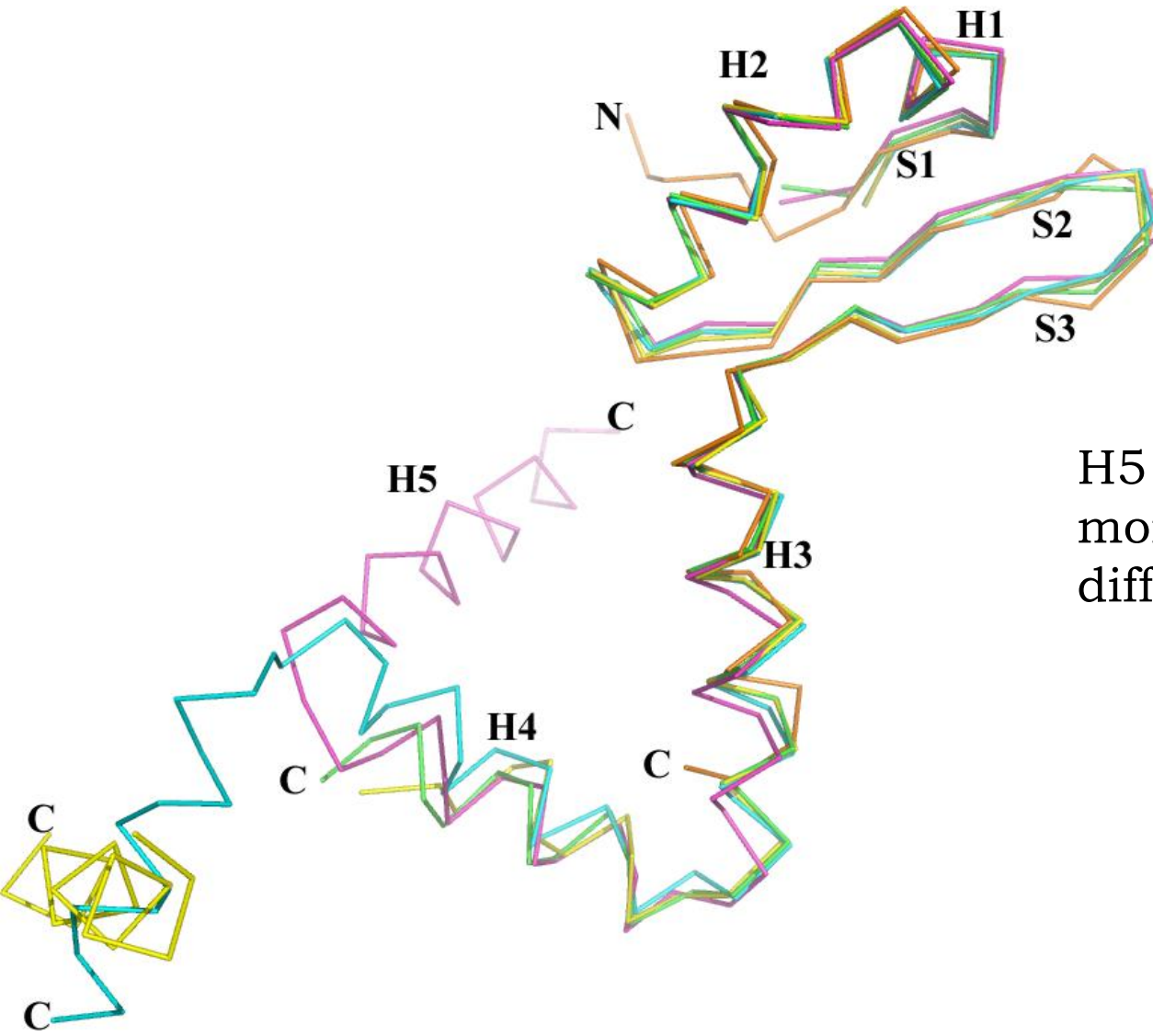
Monomer

Interactions within dimers

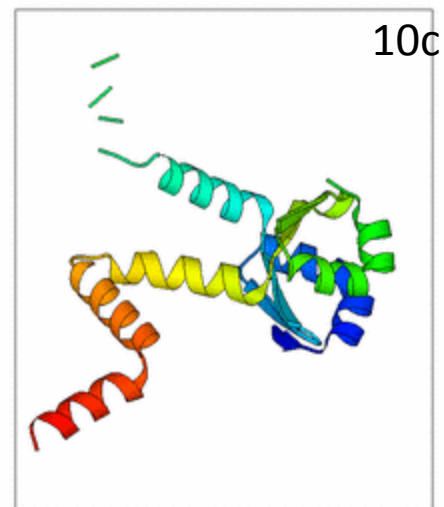
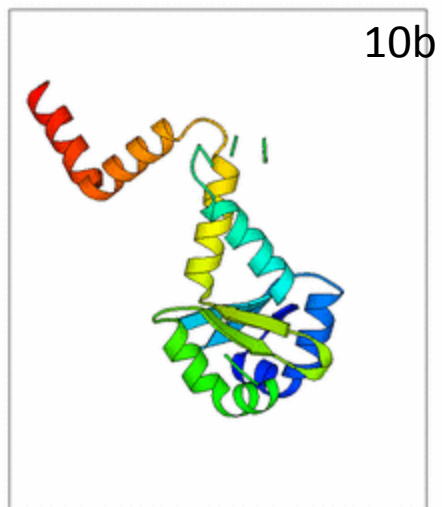
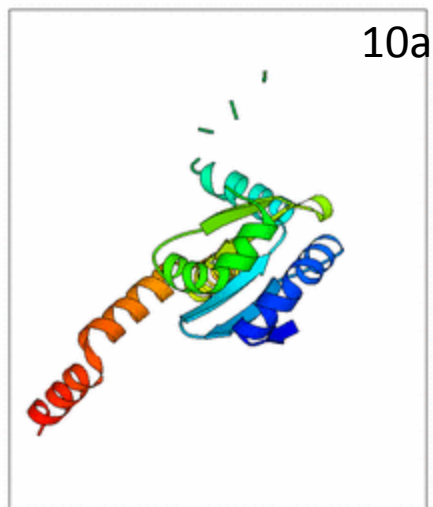
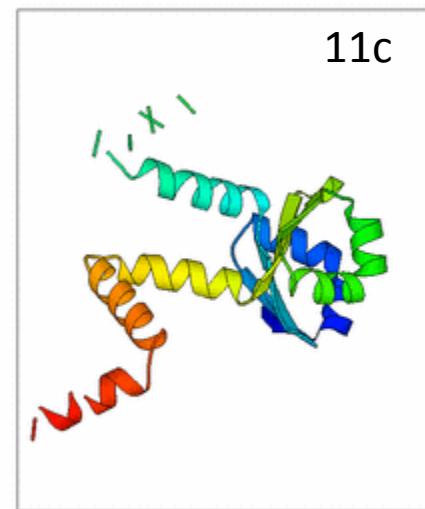
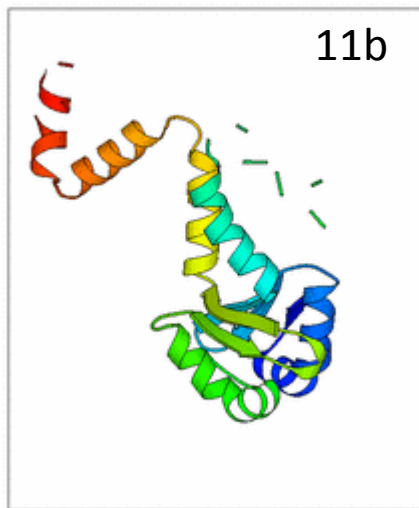
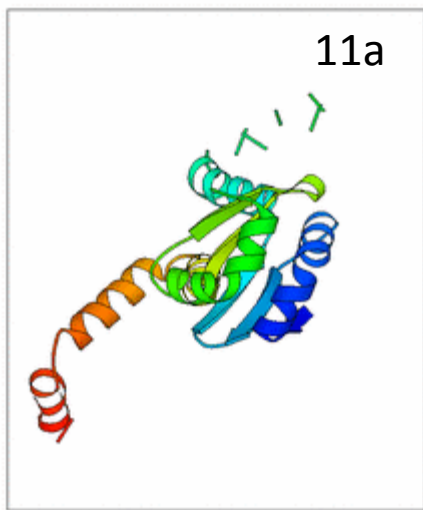


Buried area approximately 480 \AA^2

Conformational variability at the C-terminal

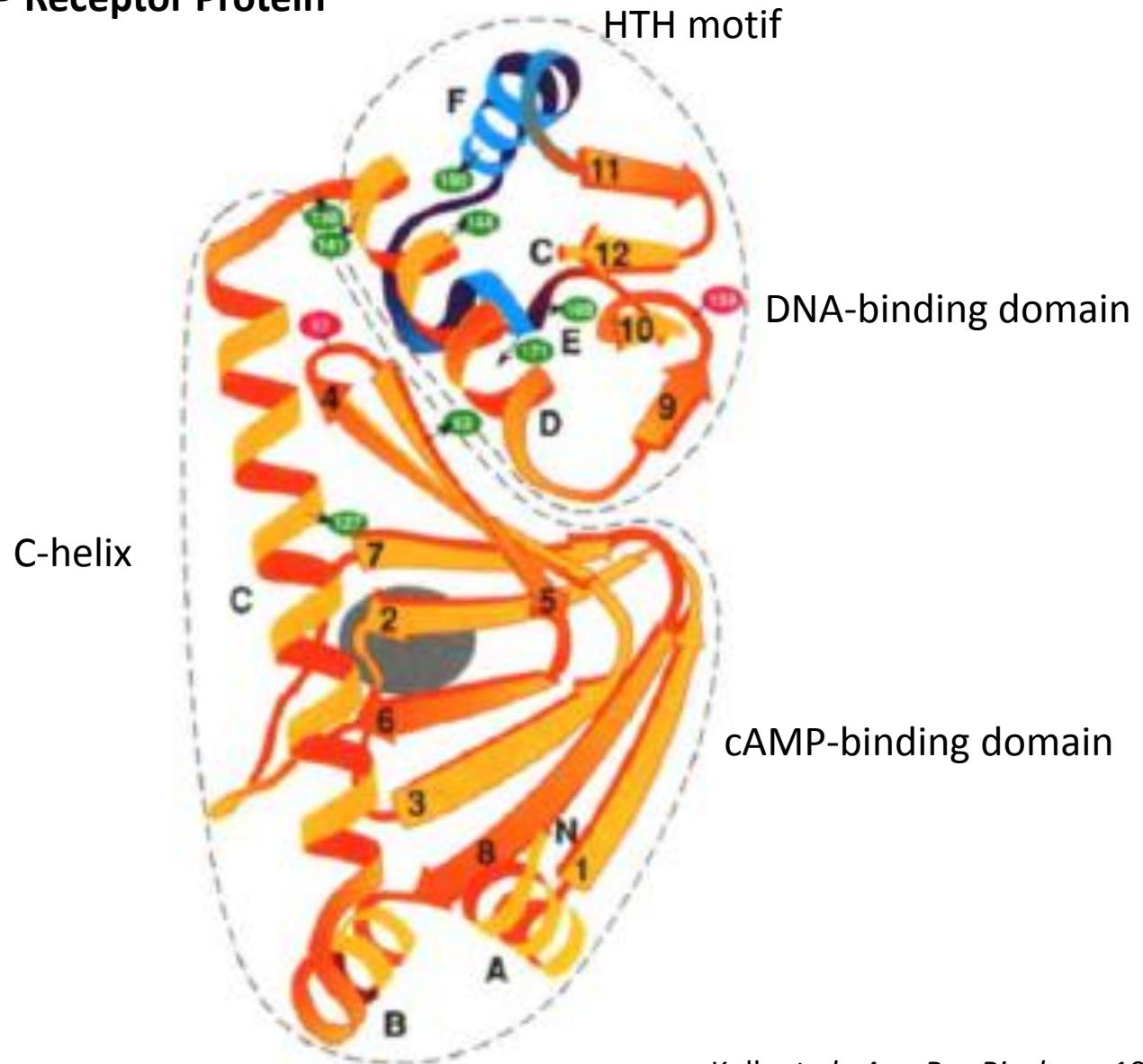


H5 helices in all the monomers adopt different conformations

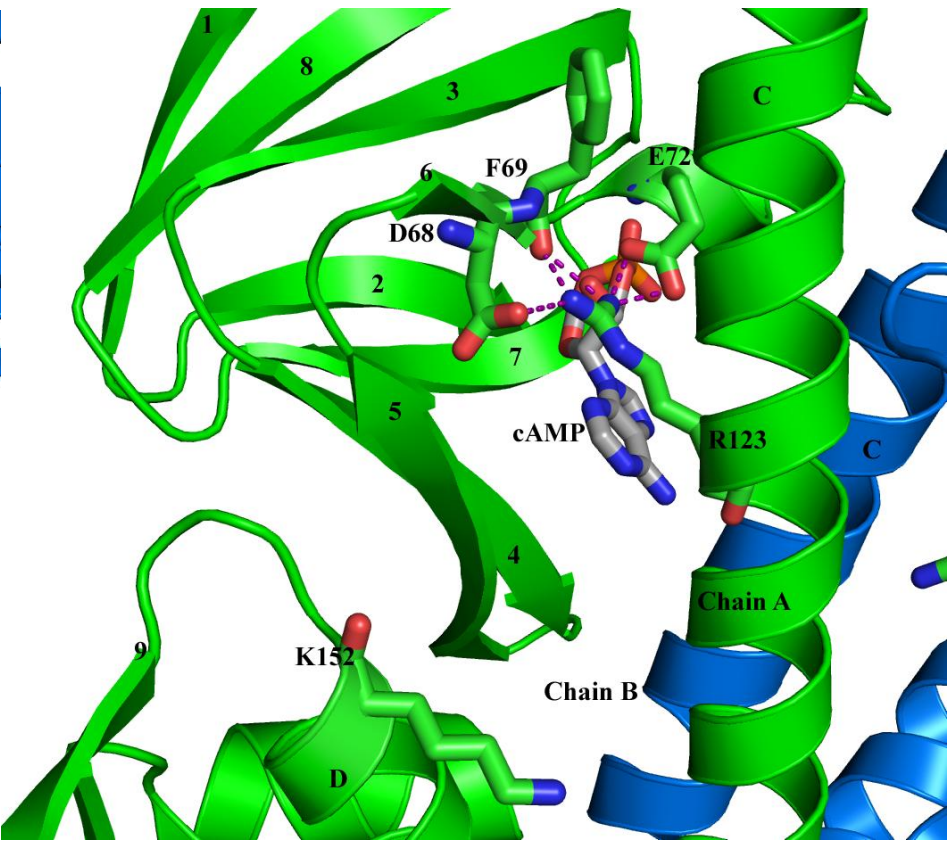
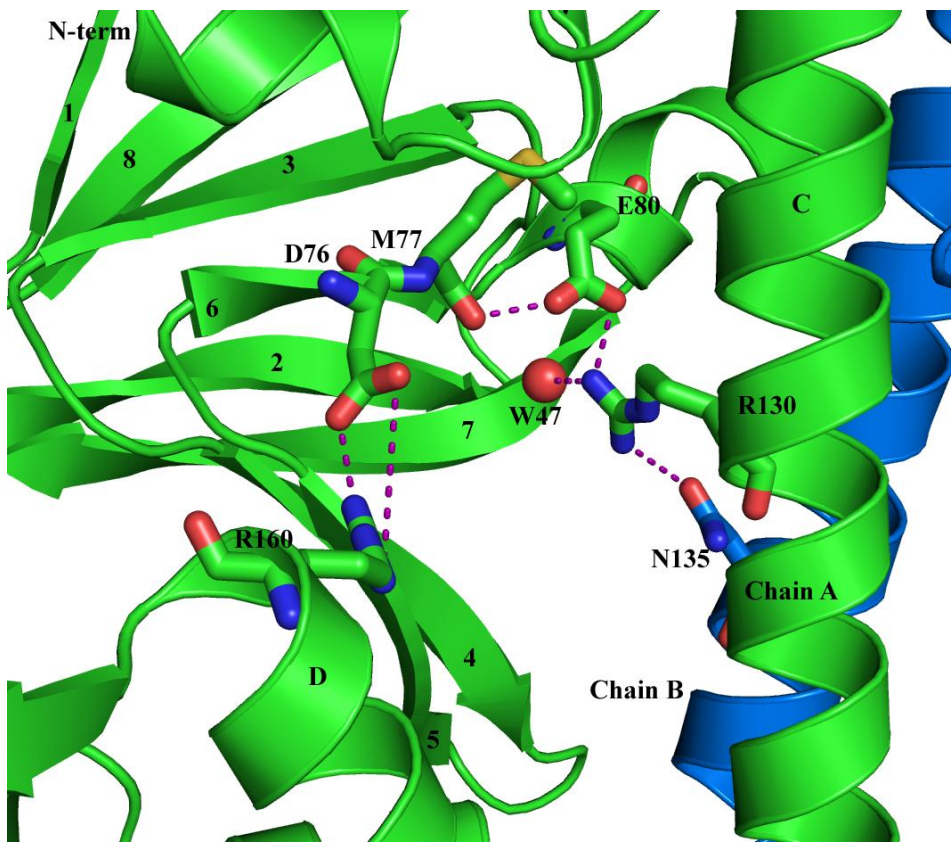


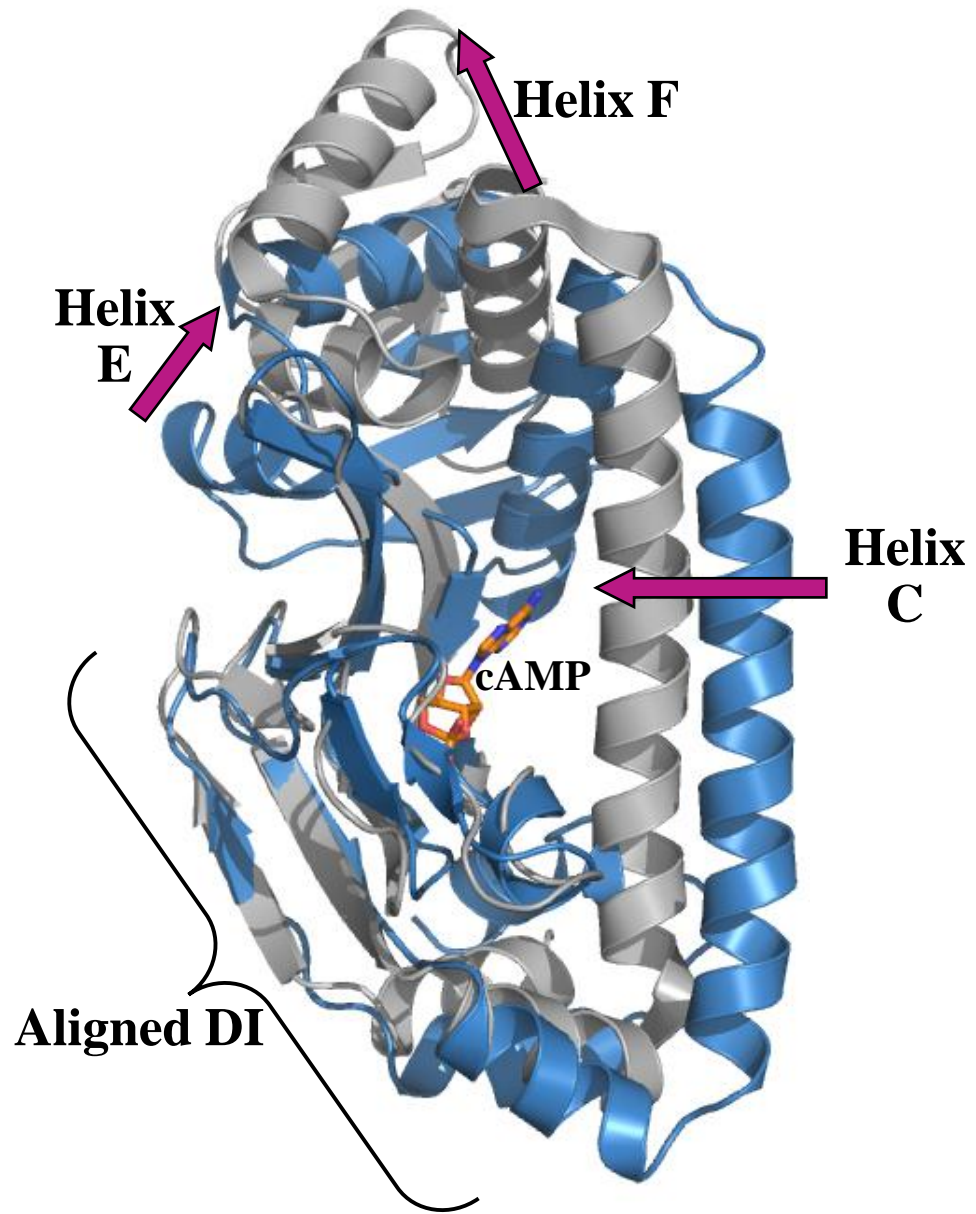
Conformational Transitions in the Cyclic AMP Receptor Protein

Structure of the cAMP Receptor Protein

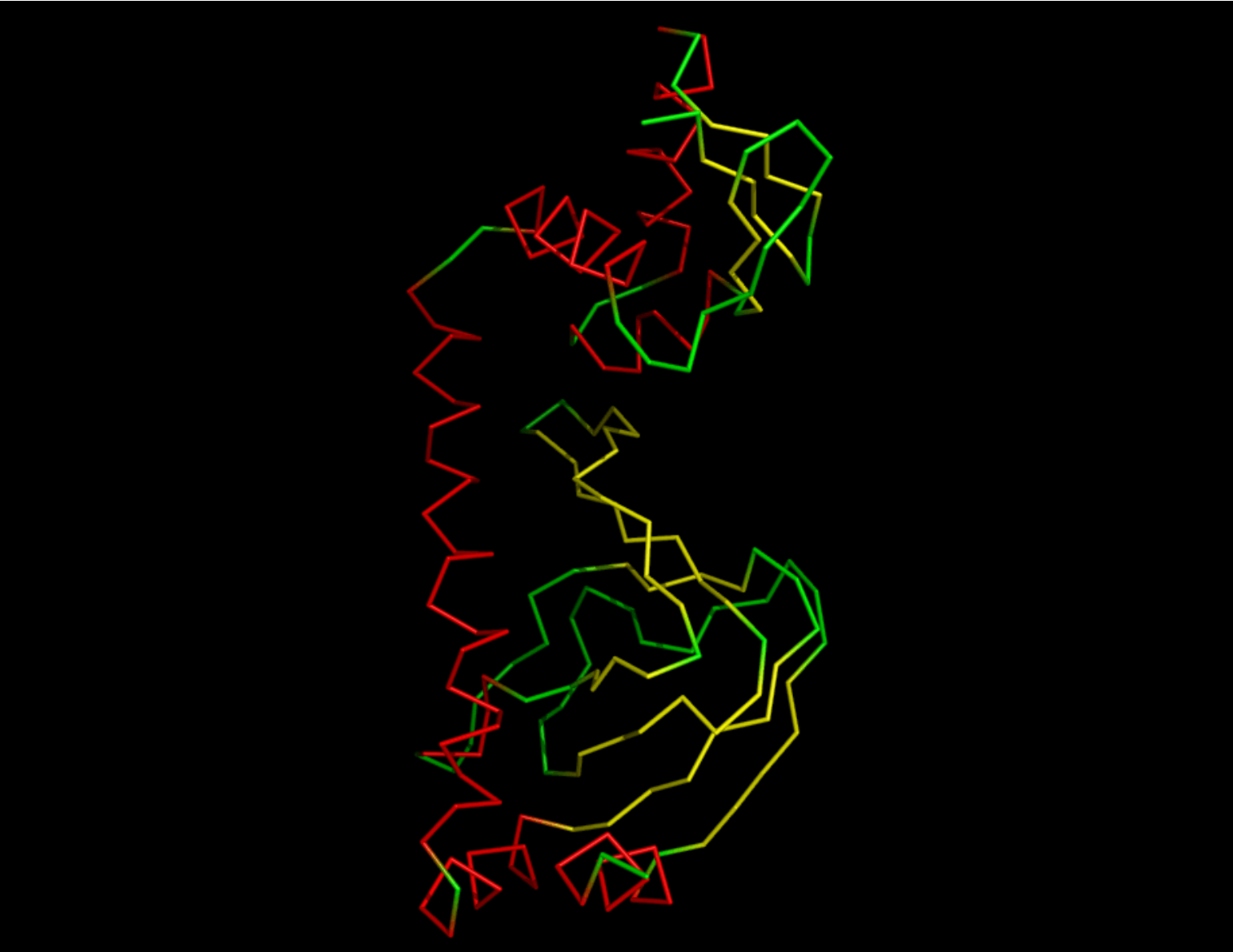


cAMP binding site comparison

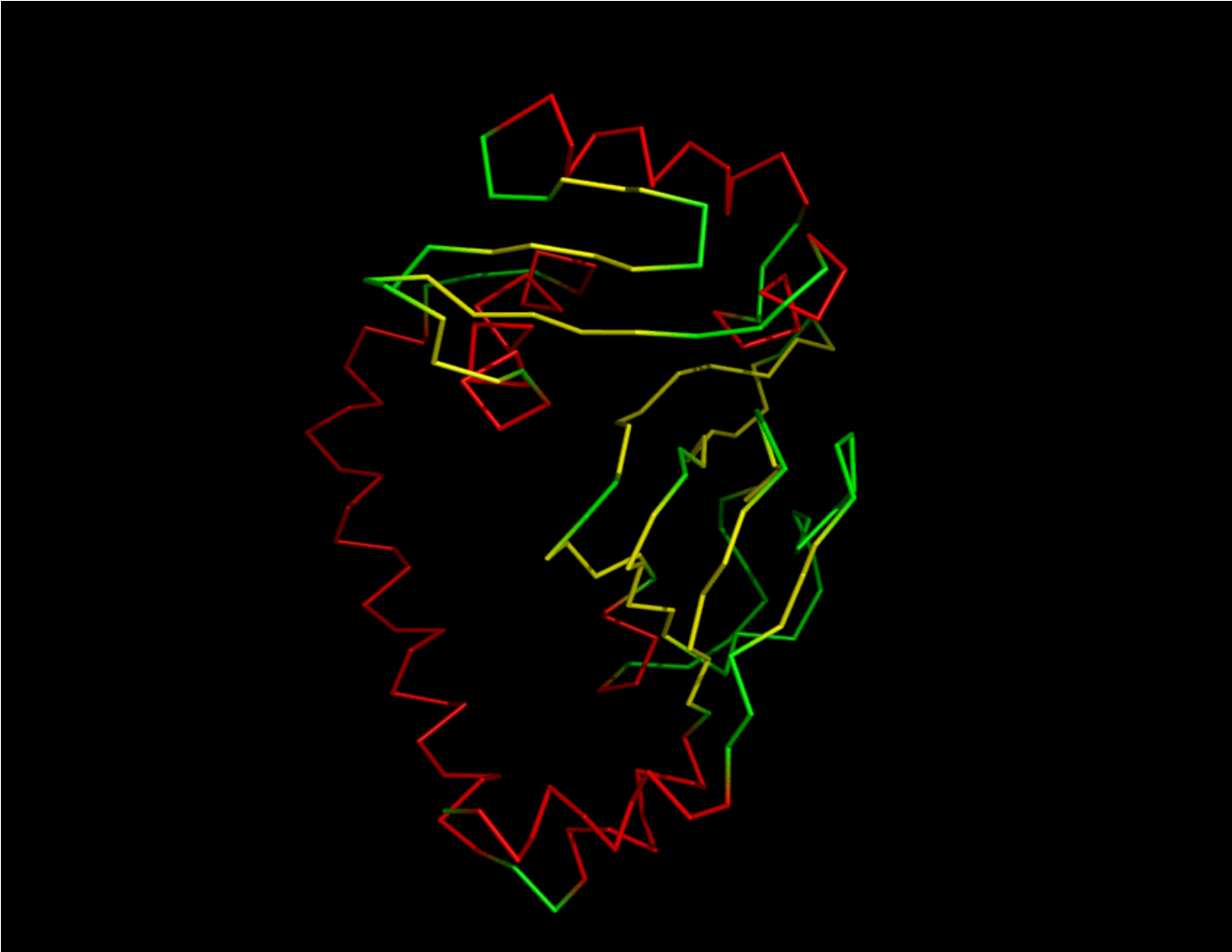


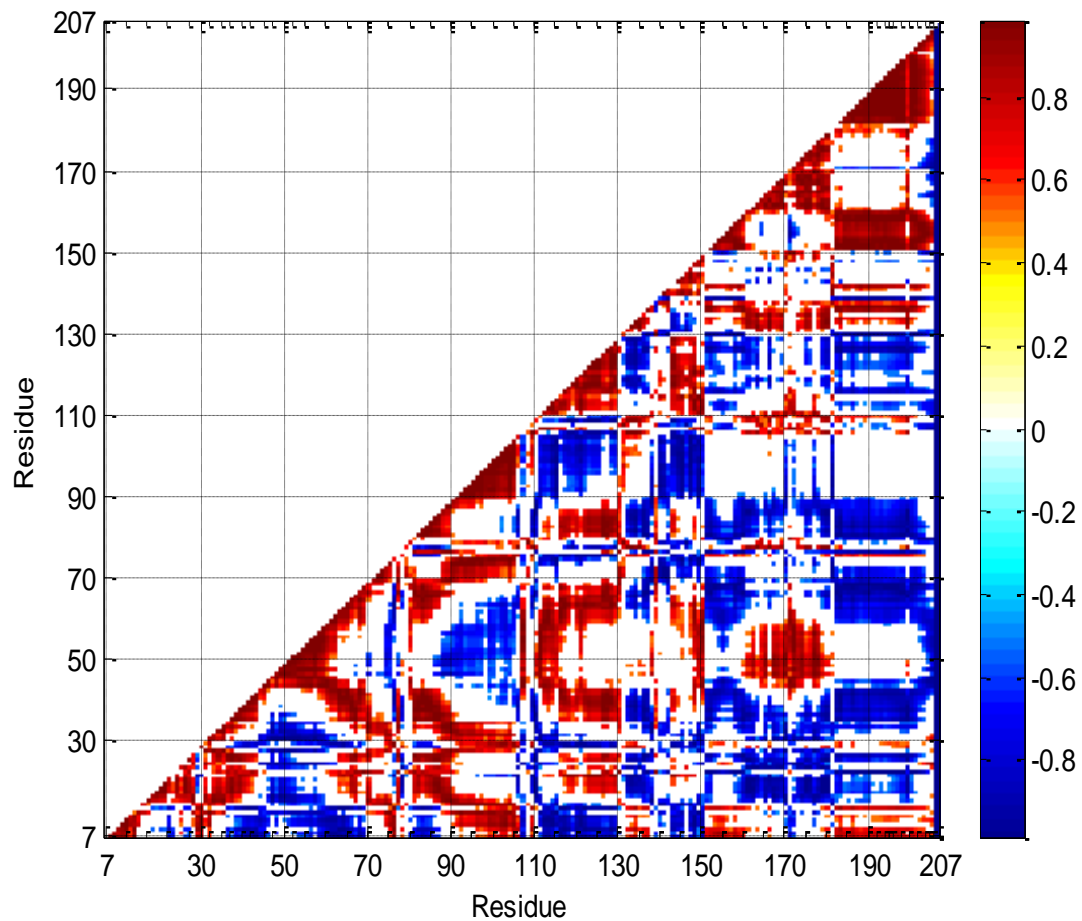


1G6N chain B

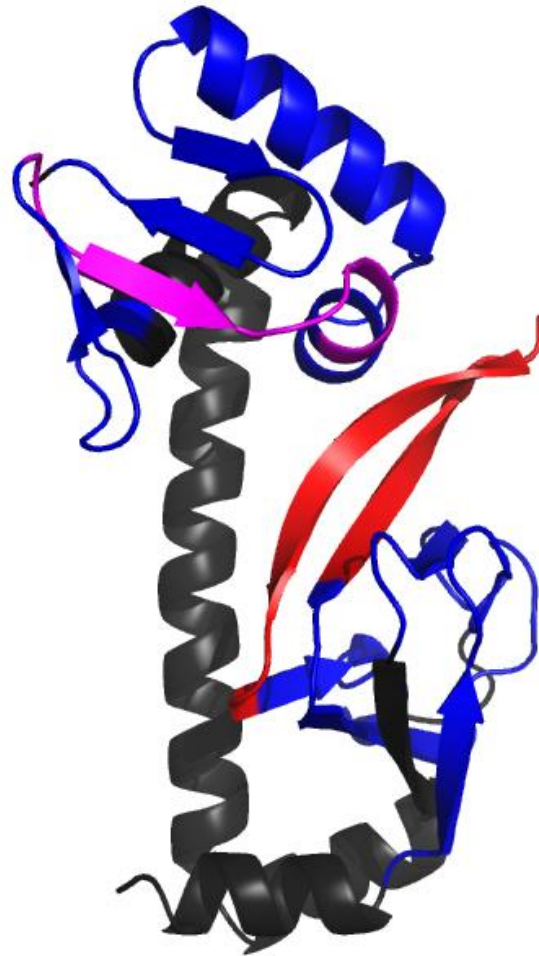


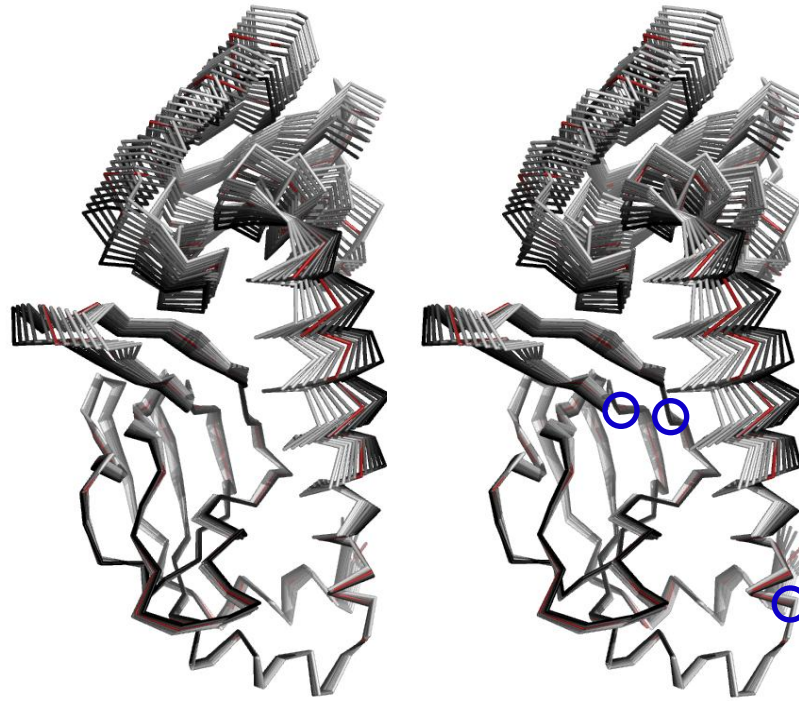
3H3U chain B





Dynamic Cross Correlation Map for 1G6N and model of *E. coli* cAMP-free CRP.





Normal Mode Analysis of 1G6N and *E.coli* model based on 3H3U structure as a reference. Mode 13 shows 57.9 % collectivity and 43.9 % overlap.

Summary of Overall Conformational Changes Effected by cAMP-binding

1. In absence of cAMP, the cAMP-binding and DNA-binding domains interact closely with each other, reducing mobility of the DNA-binding domain
2. The reduced mobility prevents sequence specific recognition of DNA
3. Binding of cAMP triggers reorientation of side chains (especially Arg 123) in the binding pocket of CRP
4. The cAMP-binding domain is drawn towards the C-helix closing over the bound cAMP
5. Conformational change in the cAMP-binding domain forces out the DNA-binding domain away from the C-helix
6. The DNA-binding domain remains sufficiently flexible, poised for sequence-specific DNA recognition

Acknowledgements

Colleagues:

Mohd Akif

Pramod Kumar

Dhananjay Joshi

Collaborators:

Chandra Verma

Karsten Suhre

Sayed E Hasnain

Funding:

Wellcome Trust, UK

DBT

CDFD

SUN Centre of Excellence