Rapid simulation of protein functional motion

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Overview

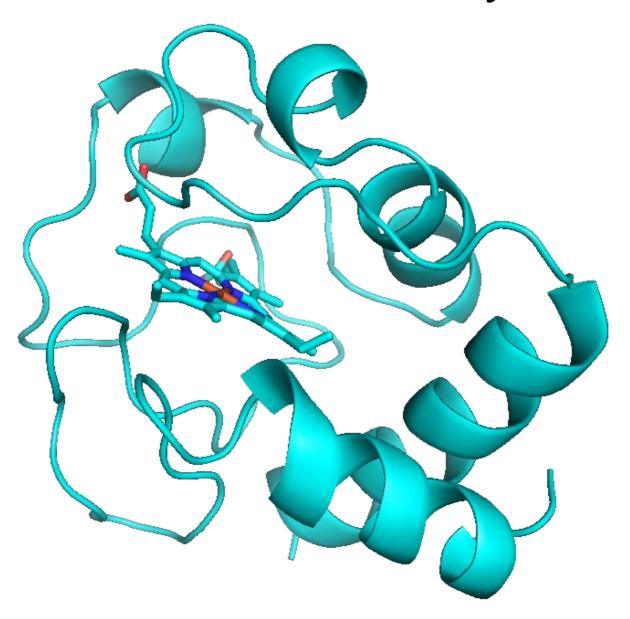
Protein Flexibility

Computational tools

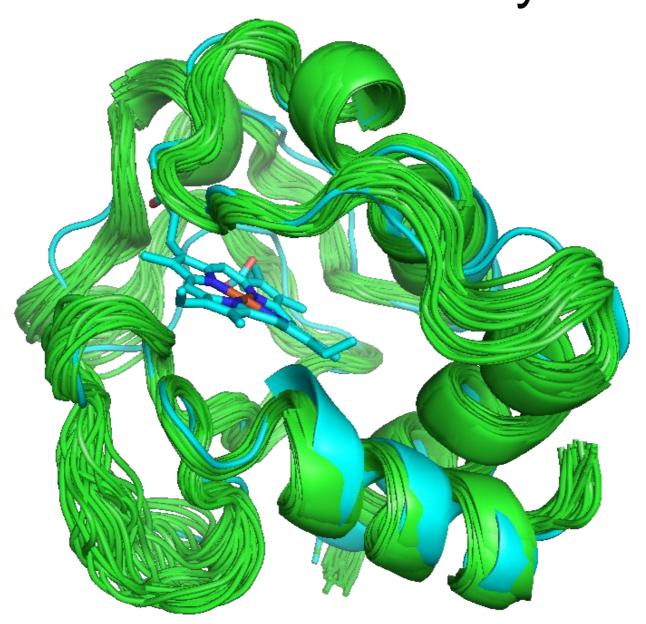
Rigidity analysis

Flexible motion

Proteins are intrinsically flexible



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A protein structure is never unique.

Each structure is a member of a flexible ensemble.

How can we investigate, understand, predict flexible motion?

Computational tools

- ·Molecular dynamics empirical potentials
 - · "Physical"
 - Individual trajectories
- Ab-initio/electronic structure
 - **ONETEP**
- .Coarse-grained MD
- Inference from structures
 - Elastic network model
 - Rigidity analysis: FIRST
- .Flexible motion
 - Geometric simulation: FRODA

Rigidity analysis

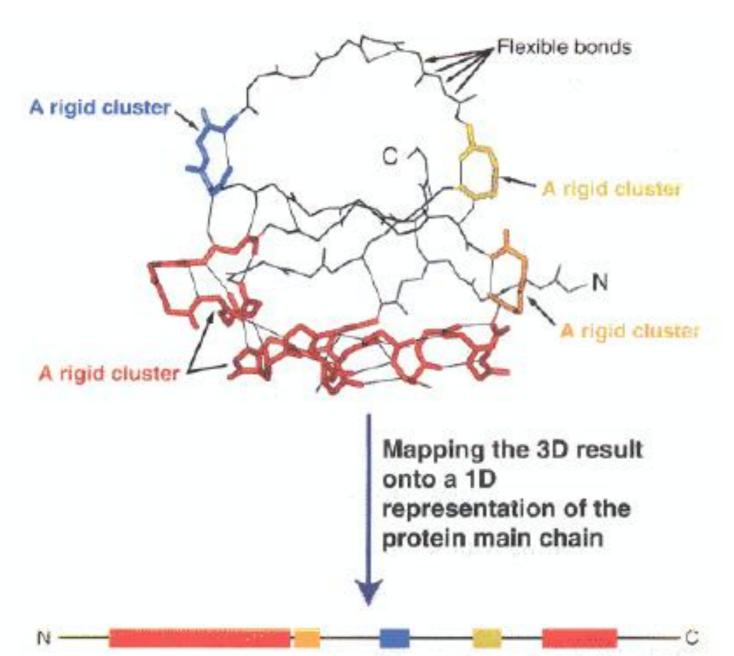
We can treat a protein as a molecular framework.

Atoms have degrees of freedom; bonds introduce constraints.

"Pebble game" algorithm matches DOF to constraints to identify rigid and flexible regions.

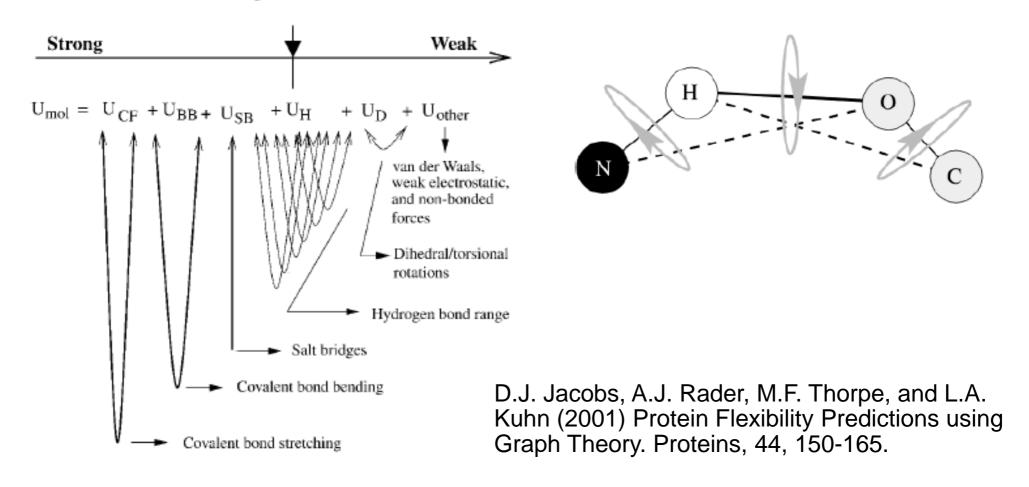
Software "FIRST" from Thorpe group at ASU (Flexible inclusions and rigid substructure topography).

Main-chain rigidity

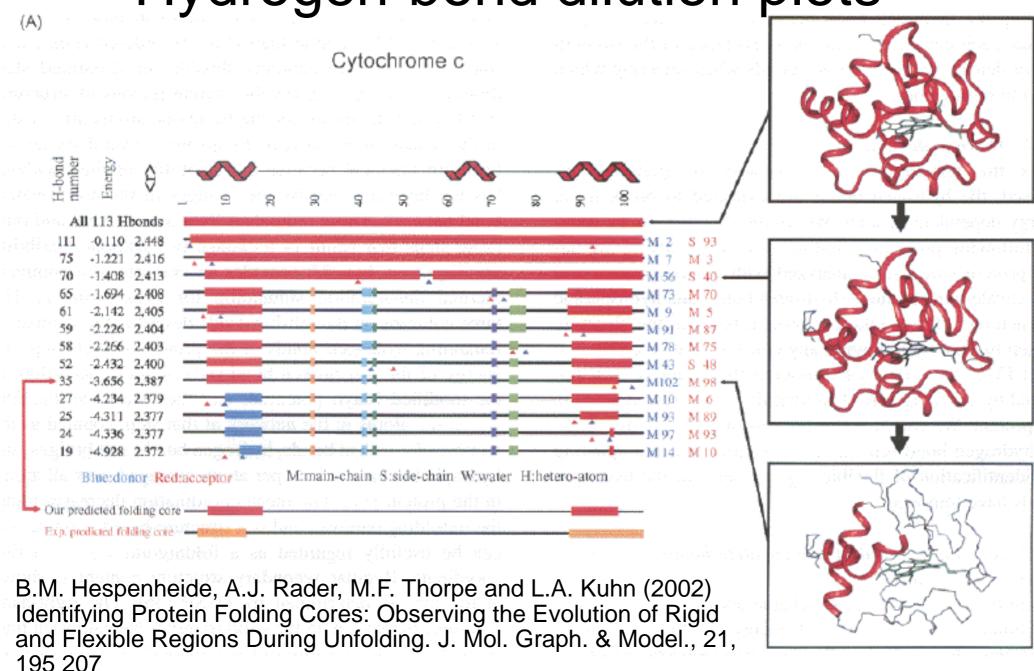


On hydrogen bonds

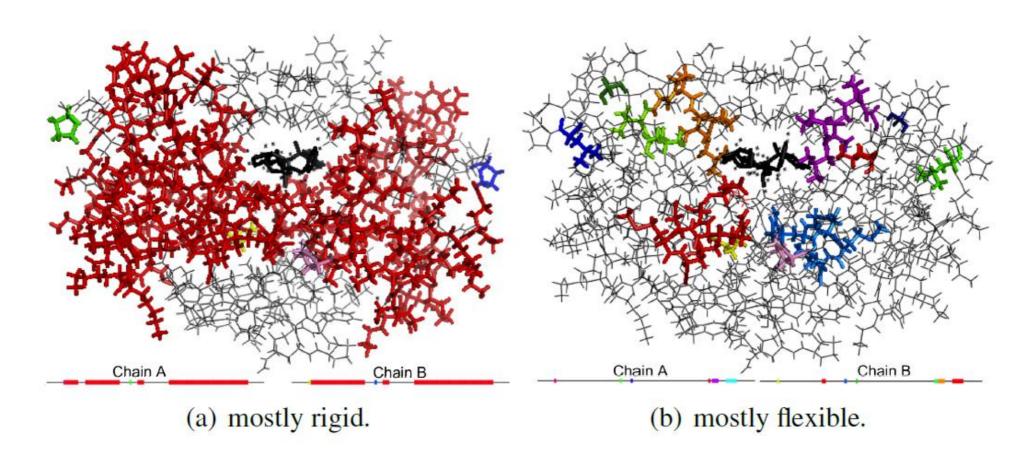
Microscopic Interactions



Hydrogen-bond dilution plots

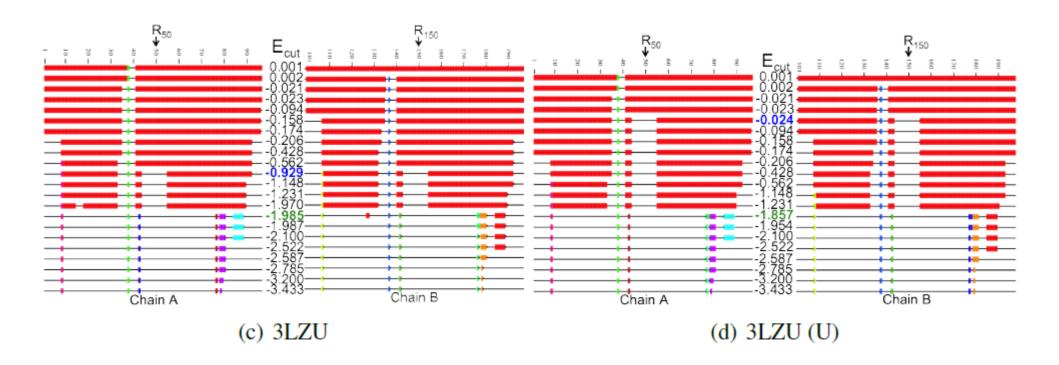


Rigidity of HIV protease



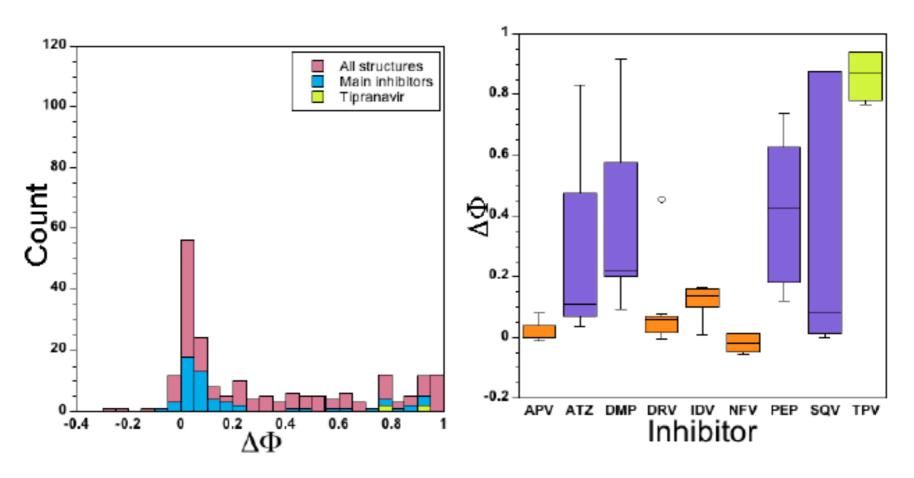
Crystal structure 3LZU at energy cutoffs of -1.970 kcal/mol (left) and of -1.985 kcal/mol (right), showing an abrupt rigidity transition.

Effect of inhibitor deletion



Deletion of the bound inhibitor (darunavir) makes the flap region around residue 50 flexible earlier in the rigidity dilution, but has little effect on the rigidity of the core and active site.

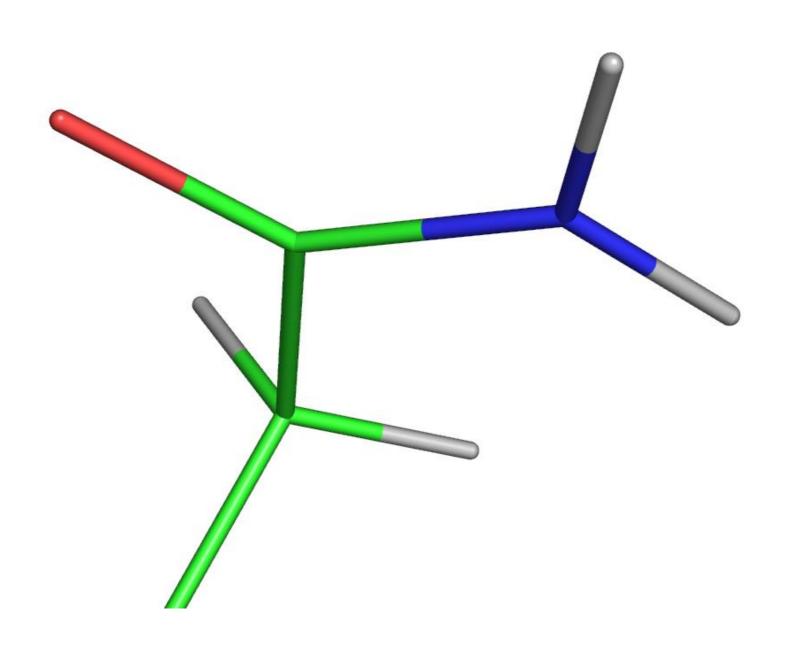
Two classes of inhibitor

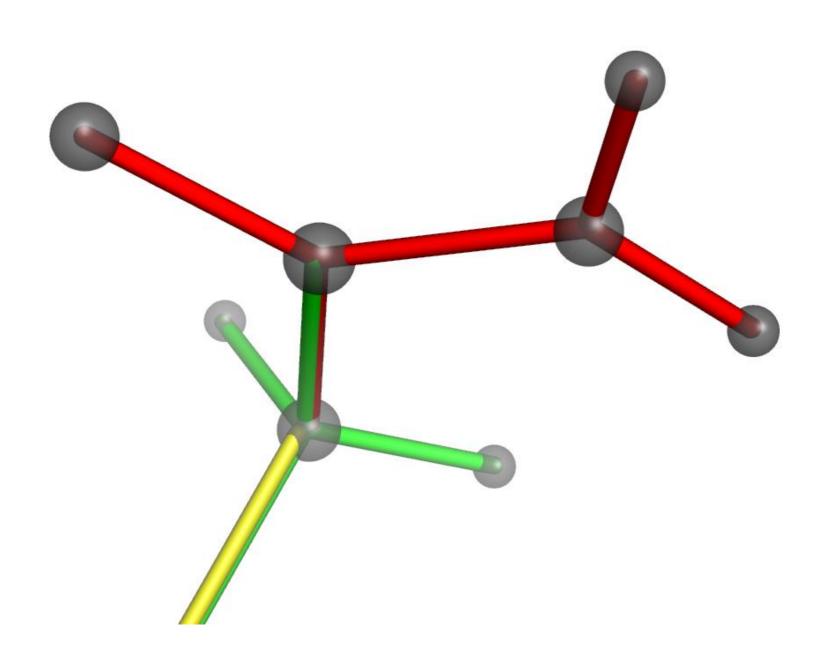


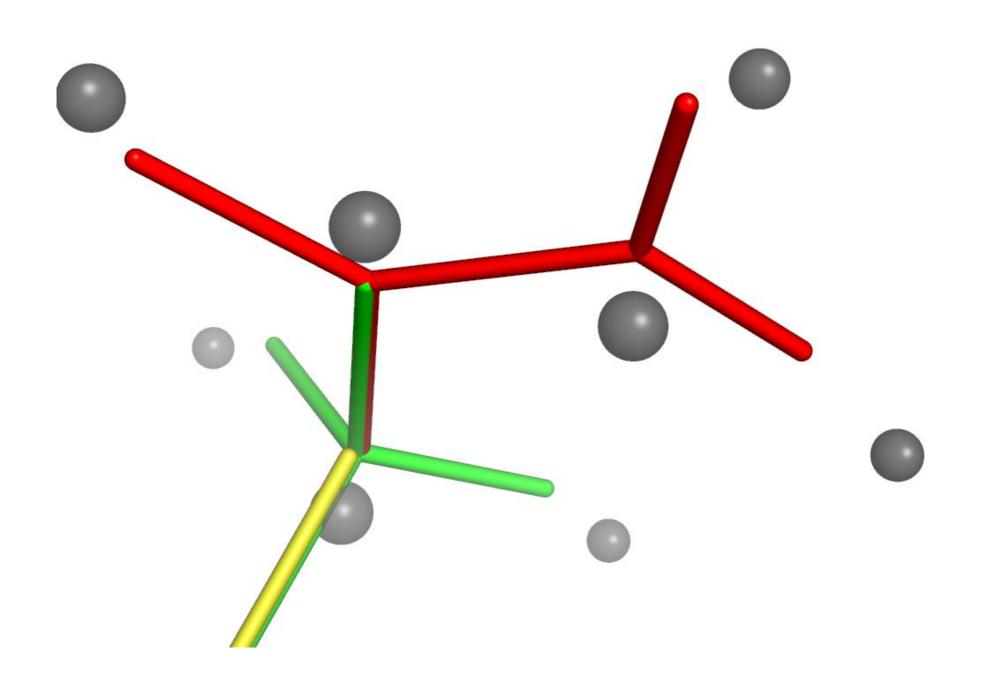
"Phi" is a ratio of energy cutoffs, indicating how flexible the flaps are relative to the core. "Delta-Phi" measures the rigidifying effect of inhibitors on the flaps. Major antiviral drug molecules fall into two classes: flap rigidifiers vs. active-site inhibitors.

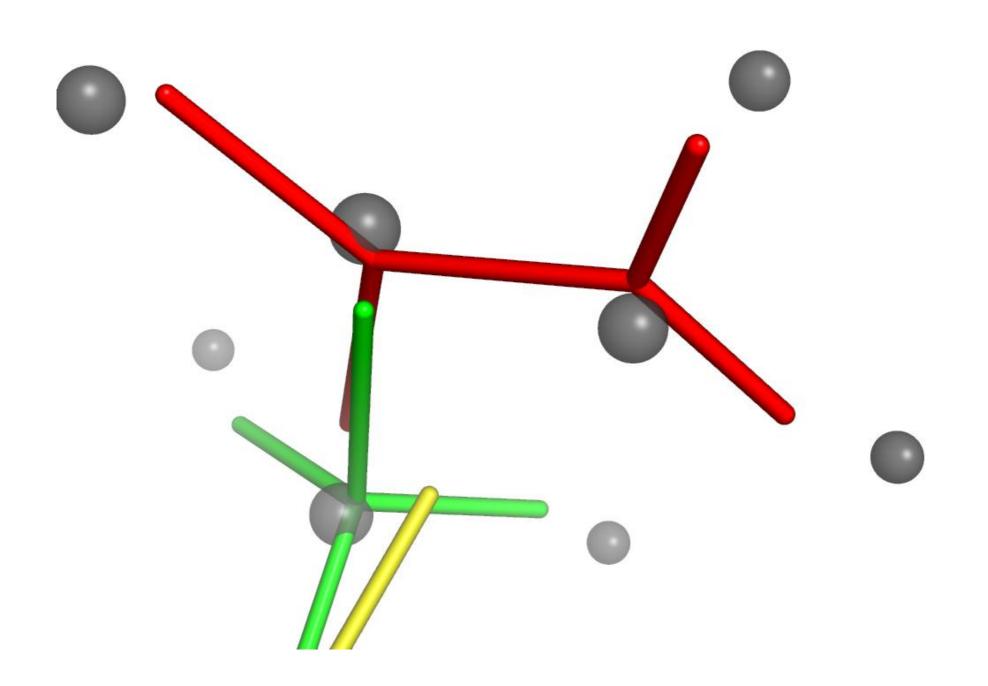
From rigidity to flexibility

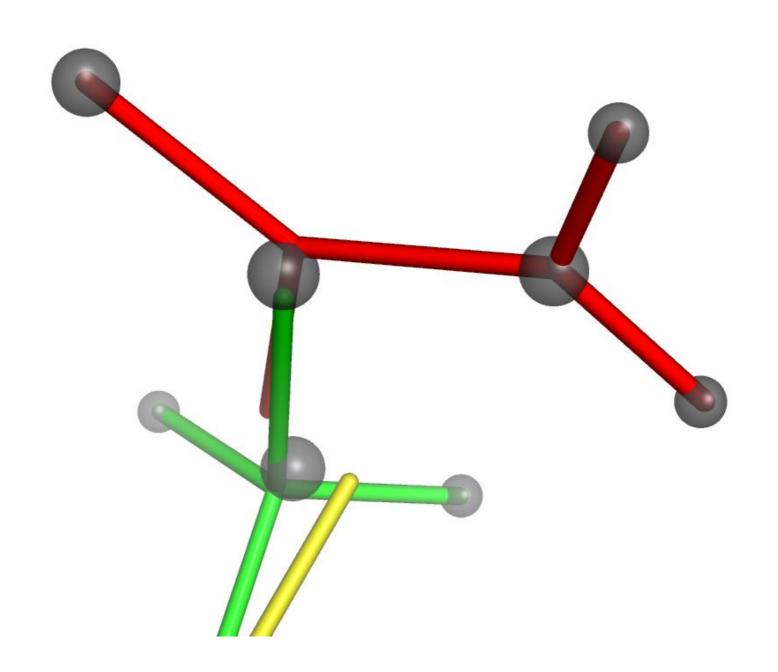
- Rigid clusters provide a basis to simulate flexible motion
- Motivations:
 - Connect static (crystal) and dynamic (NMR) structures
 - Explore large-scale motions
- Cheap and cheerful
- "FRODA" (Framework Rigidity Optimised Dynamic Algorithm) – included with FIRST

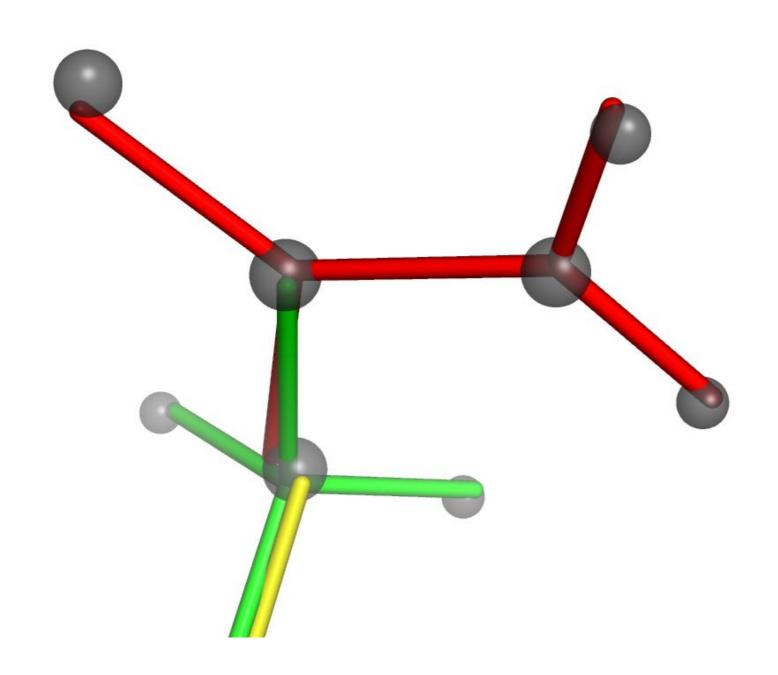


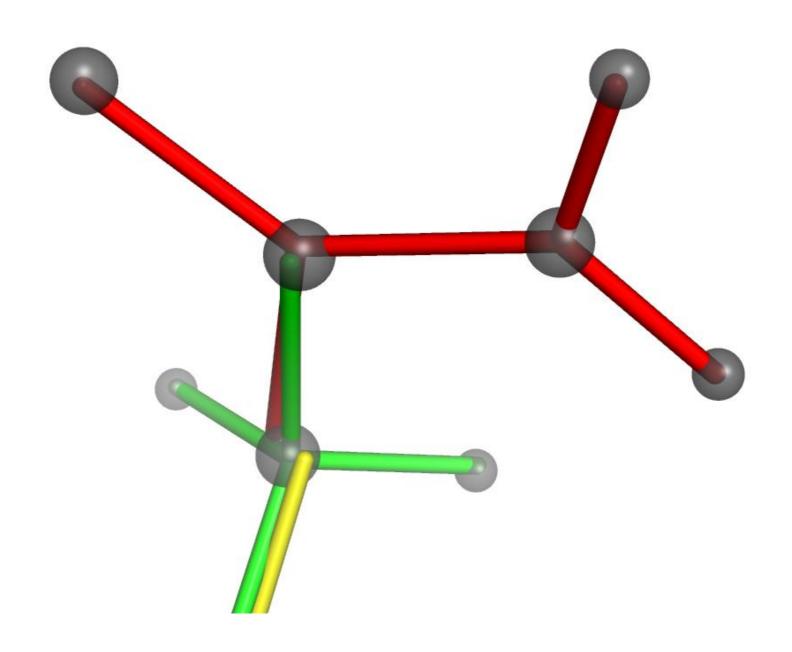


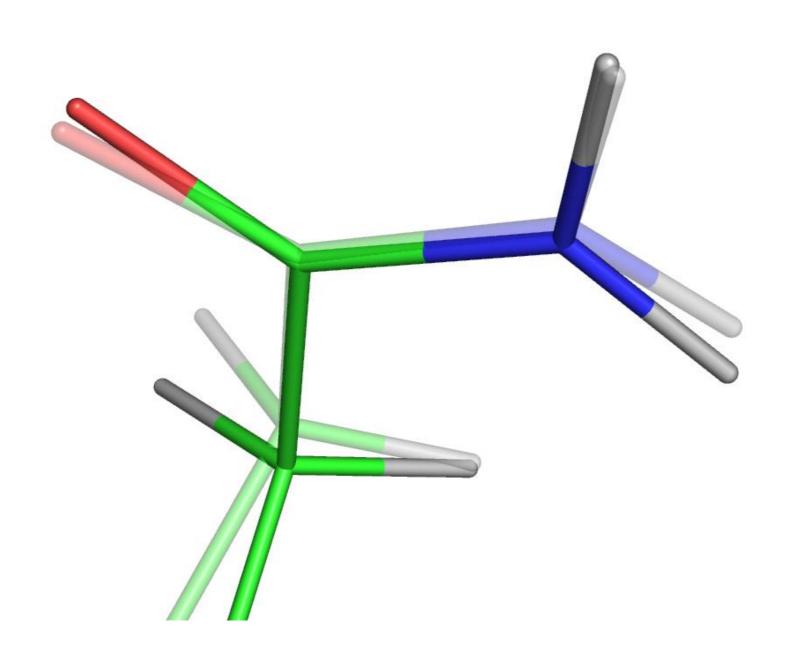












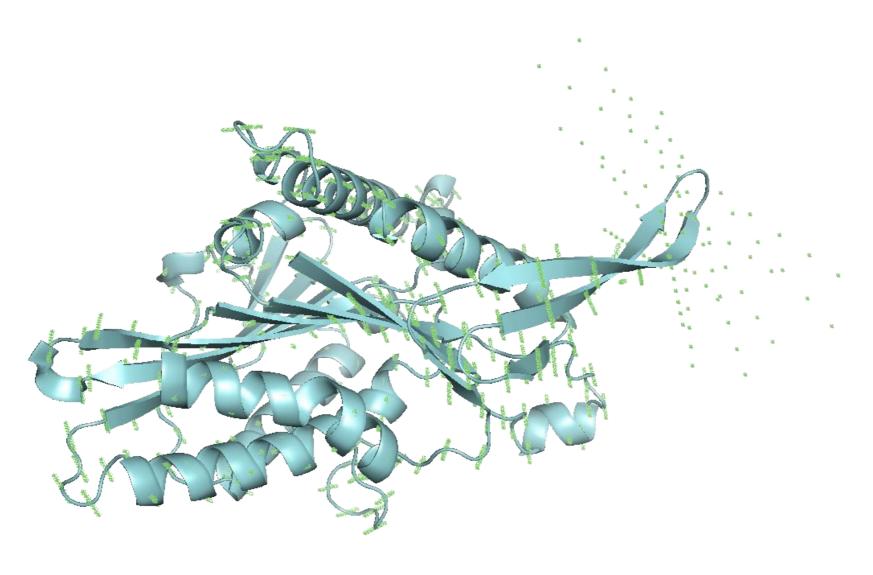
Elastic network models suggest directions for flexible motion

We use the "EINeMo" elastic network model to calculate mode eigenvectors.

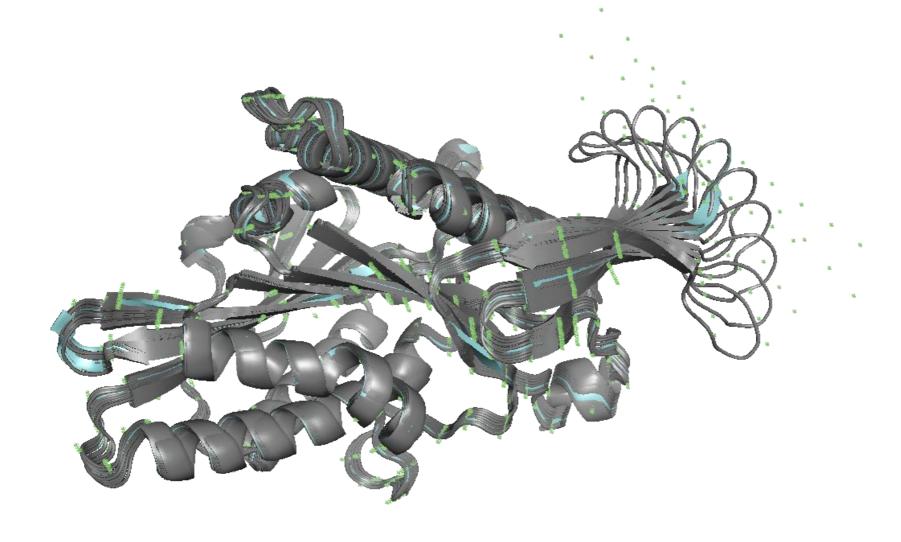
Eigenvectors are used to bias motion in FRODA.

FRODA investigates flexible motion while maintaining rigidity constraints.

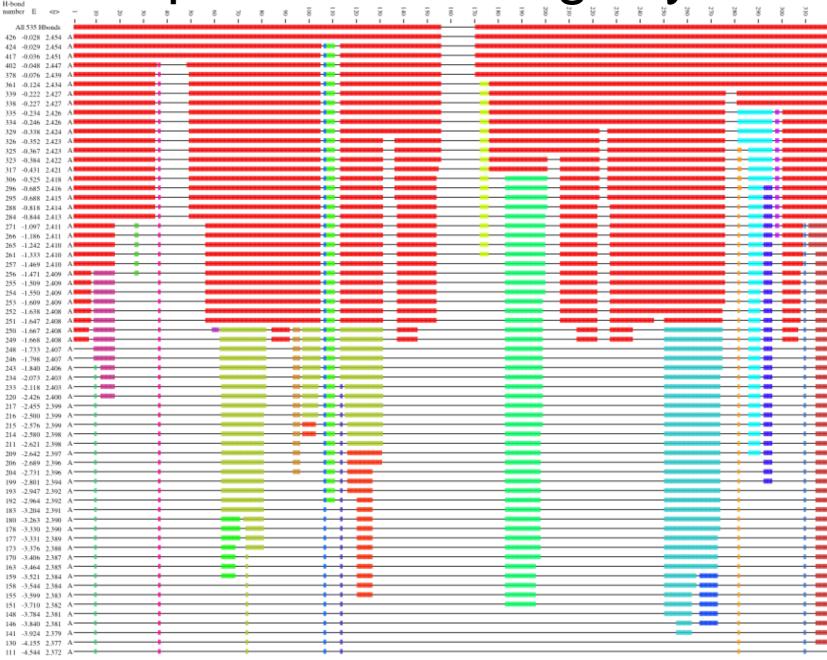
Internal kinesin motor domain (1RY6)



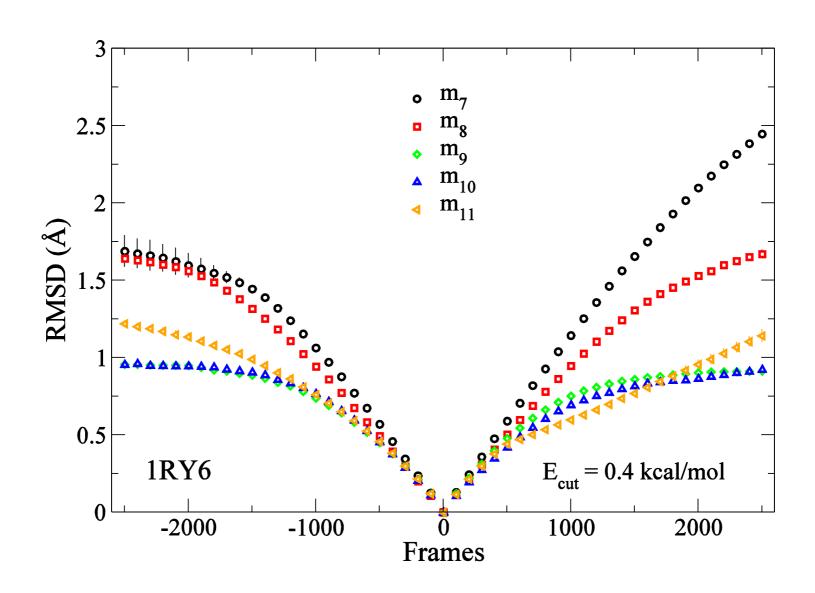
Internal kinesin motor domain (1RY6)



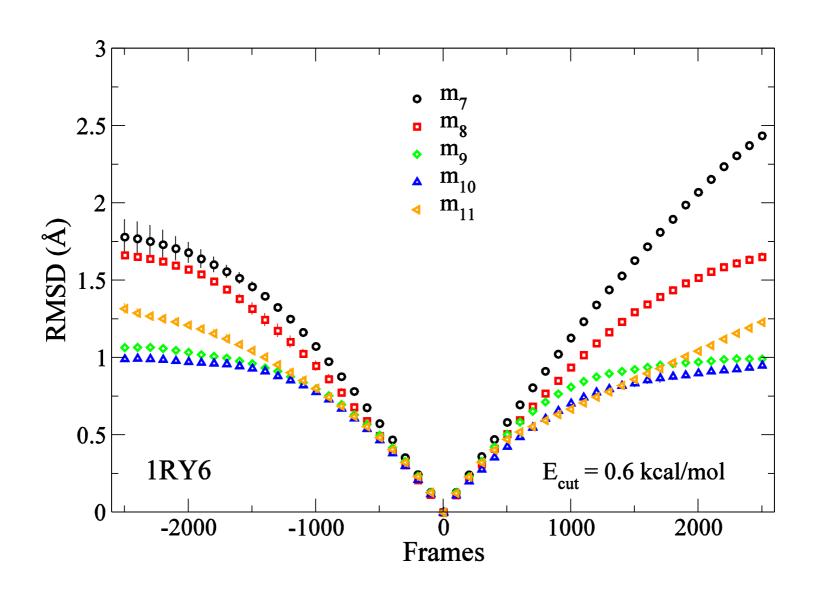
Dependence on rigidity?



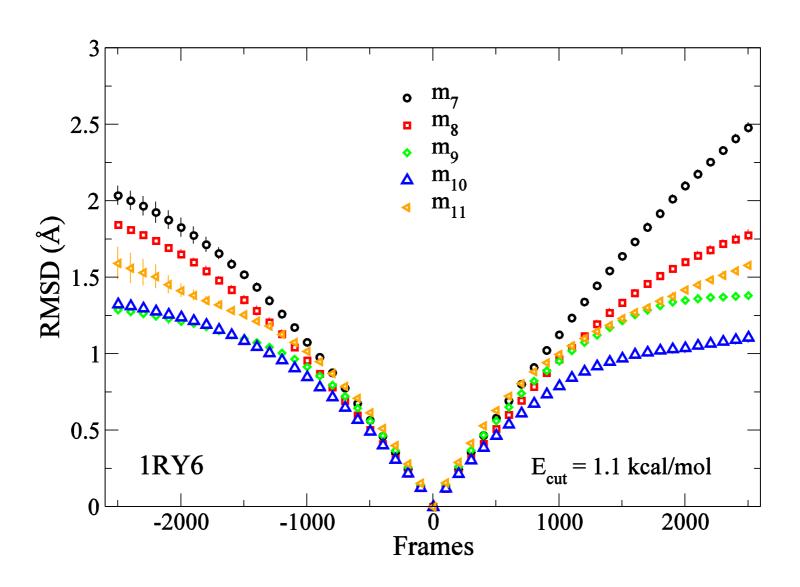
Dependence on energy cutoff



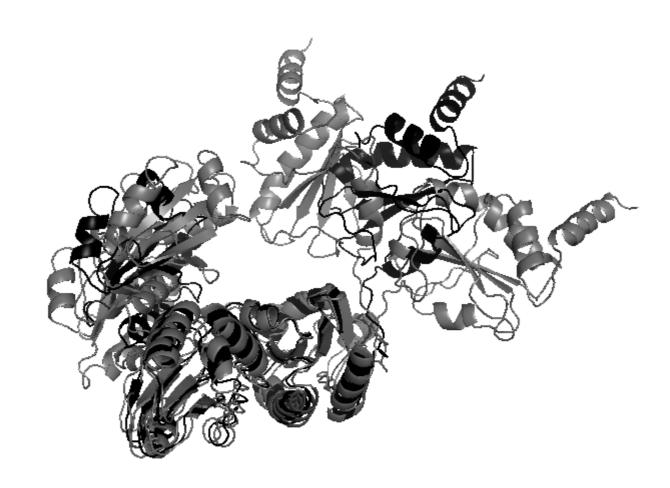
Dependence on energy cutoff



Dependence on energy cutoff

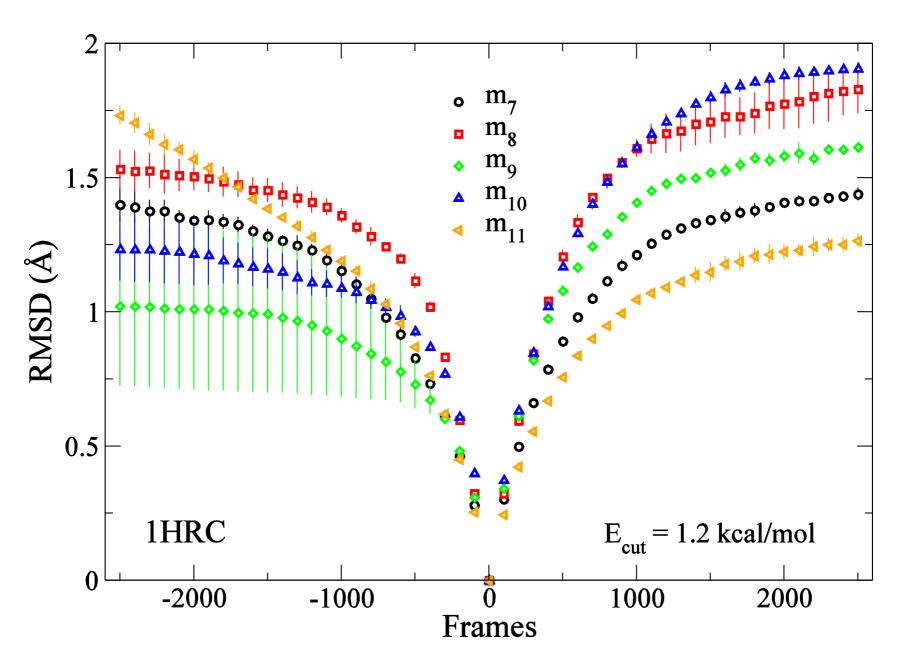


Amplitude exploration: PDI

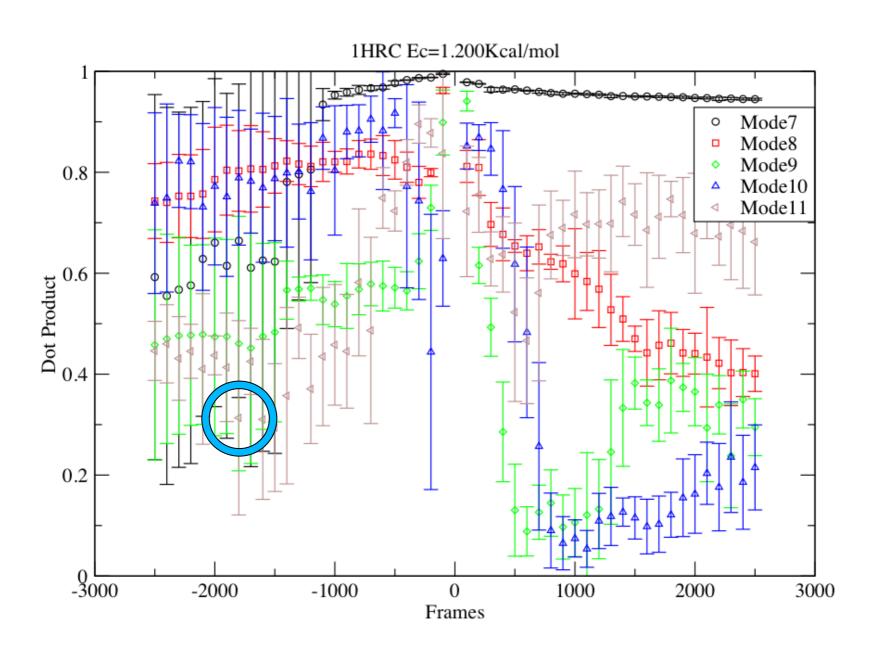


Emilio Jimenez and Moitrayee Bhattacharyya will have much more to say...

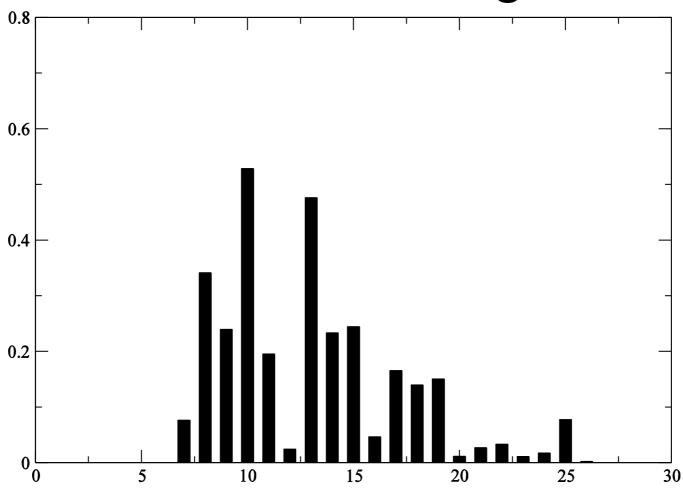
Persistence of modes? 1HRC



Persistence of mode eigenvector



Mode mixing



After projection over an RMSD ~ 1 A, the original mode 11 has a small overlap with the current mode 11 but has large overlaps with several flexible modes including 10, 13.

Modes for flexible motion

- Modes calculated from mass-and-spring model
- Harmonic oscillator: inertia-dominated
- Protein Reynolds number?

$$\frac{VL}{v} \sim \frac{(\mathring{A}/\mu s)(\mathring{A})}{10^{-6} m^2 s^{-1}} < 10^{-(4 \text{ to } 10)}$$

 Viscosity dominates: solvent pushes the protein along easy directions?

What is the point?

- To have scientific value our simulations must connect back to experiment!
- Understanding and interpretation of:
- NMR, crystal diffraction data, AFM, FRET, linkers, functional studies...

References

- FIRST/FRODA: http://flexweb.asu.edu (Thorpe et al.); pathways.asu.edu
- EINeMo: http://www.igs.cnrs-mrs.fr/elnemo/ (Tirion, Tama, Sanejouand et al.)
- SA Wells, S Menor, BM Hespenheide and MF Thorpe "Constrained geometric simulation of diffusive motion in proteins." Physical Biology 2, S127-S136 (2005).
- SA Wells, JE Jimenez and RA Roemer "Comparative analysis of rigidity across protein families." Physical Biology 6, 046005 (2009).
- JW Heal, SA Wells, JE Jimenez-Roldan, RB Freedman, RA Roemer "Rigidity analysis of HIV-1 protease." Proceedings of IOP Condensed Matter and Materials Physics "CMMP10". Journal of Physics: Conference Series 286 (2011).
- JE Jimenez-Roldan, SA Wells, RB Freedman, RA Roemer "Integration of FIRST, FRODA and NMM in a coarse grained method to study Protein Disulphide Isomerase conformational change." Proceedings of IOP Condensed Matter and Materials Physics "CMMP10". Journal of Physics: Conference Series 286 (2011).

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